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U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER
12660-002001TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

U.S. APPLICATION NO. (If Known, see 37 CFR 1.5)

09/720285

INTERNATIONAL APPLICATION NO.
PCT/JP99/03351INTERNATIONAL FILING DATE
23 June 1999PRIORITY DATE CLAIMED
24 June 1998

TITLE OF INVENTION

NOVEL HEMOPOIETIN RECEPTOR PROTEINS

APPLICANT(S) FOR DO/EO/US

Hitoshi Nomura and Masatsugu Maeda

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to promptly begin national examination procedures (35 U.S.C. 371(f)).
4. ☒ The US has been elected by the expiration of 19 months from the priority date (PCT Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US)
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2))
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made, however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made
8. ☐ An English language translation of amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 16 below concern other documents or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A FIRST preliminary amendment
☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☐ Other items or information:

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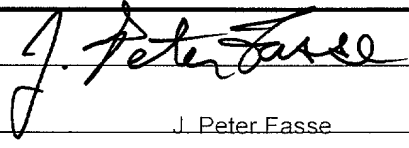
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I hereby certify under 37 CFR §1.10 that this correspondence is being deposited with the United States Postal Service as Express Mail Post Office to Addressee with sufficient postage on the date indicated below and is addressed to the Commissioner for Patents Washington, D.C. 20231

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Signature

Victor Maeda
Typed Name of
Person Signing

U.S. APPLICATION NO (IF KNOWN) 09/720285		INTERNATIONAL APPLICATION NO PCT/JP99/03351		ATTORNEY'S DOCKET NUMBER 12660-002001	
17. <input checked="" type="checkbox"/> The following fees are submitted: Basic National Fee (37 CFR 1.492(a)(1)-(5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1000 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO. \$710 International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4)..... \$690 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4)..... \$100 <div style="text-align: right;">ENTER APPROPRIATE BASIC FEE AMOUNT =</div>				CALCULATIONS PTO USE ONLY	
Surcharge of \$130 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e))				\$0.00	
Claims	Number Filed	Number Extra	Rate		
Total Claims	29 - 20 =	9	x \$18	\$162.00	
Independent Claims	2 - 3 =	0	x \$80	\$0.00	
MULTIPLE DEPENDENT CLAIMS(S) (if applicable)			+ \$270	\$0.00	
TOTAL OF ABOVE CALCULATIONS =				\$1,022.00	
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				\$0.00	
SUBTOTAL =				\$0.00	
Processing fee of \$130 for furnishing the English Translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f))				\$0.00	
TOTAL NATIONAL FEE =				\$0.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)) The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) \$40.00 per property +				\$0.00	
TOTAL FEES ENCLOSED =				1,022.00	
				Amount to be refunded:	\$
				Charged:	\$
a. <input checked="" type="checkbox"/> A check in the amount of \$1,022.00 to cover the above fees is enclosed. b. <input type="checkbox"/> Please charge my Deposit Account No. 06-1050 in the amount of \$0.00 to cover the above fees. A duplicate copy of this sheet is enclosed. c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 06-1050. A duplicate copy of this sheet is enclosed					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO					
J. Peter Fasse FISH & RICHARDSON P.C. 225 Franklin Street Boston, MA 02110-2804 (617) 542-5070 phone (617) 542-8906 facsimile			SIGNATURE  NAME J. Peter Fasse REGISTRATION NUMBER 32,983		



JC08 Rec'd PCT/PTO 1 2 FEB 2001

Attorney's Docket No.: 12660-002001 / 62-004PCT-US

09/720285

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Hitoshi Nomura et al.

Art Unit : Unknown

Serial No. : 09/720,285

Examiner : Unknown

Filed : December 21, 2000

Title : NOVEL HEMOPOIETIN RECEPTOR PROTEINS

Commissioner for Patents

Washington, D.C. 20231

5F00

SECOND PRELIMINARY AMENDMENT

Prior to examination, please amend the application as follows.

In the Specification:

At page 31, lines 10-13, delete "Figure 16 shows a comparison between amino acid sequences of human and mouse NR8 β . The amino acid sequences where the two coincide are shadowed. Also, cysteine residues conserved in other hemopoietin receptors are displayed in boldface type within the sequence."

At page 31, lines 14-18, delete "Figure 17 shows a comparison between amino acid sequences of human and mouse NR8 γ . The amino acid sequences where the two coincide are shadowed. Also, cysteine residues conserved in other hemopoietin receptors and the WSXWS-Box are displayed in boldface type within the sequence."

At page 31, line 19, replace "18" with --16--.

At page 31, line 23, replace "19" with --17--.

At page 55, lines 29-30, delete "Figure 16 shows a comparison between human and mouse NR8 β amino acid sequences."

At page 55, lines 30-31, delete "Figure 17 shows a comparison between human and mouse NR8 γ amino acid sequences."

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2/6/01

Signature

Susan R. Jones

Typed or Printed Name of Person Signing Certificate

At page 57, line 3, replace "18" with --16--.

At page 58, line 1, replace "19" with --17--.

In the Claims:

Please amend claims 24, 25, 28 to 32, 34 to 36, and 43 as follows.

24. (Amended) The polypeptide of claim 23, wherein the polypeptide is human NR8-alpha comprising the amino acid sequence from the [1st amino acid Met] 20th amino acid Cys to the 361st amino acid Ser of SEQ ID NO:1, or a modified human NR8-alpha polypeptide comprising the amino acid sequence of said human NR8-alpha polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said human NR8-alpha polypeptide.

25. (Amended) The polypeptide of claim 23, wherein the polypeptide is human NR8-beta comprising the amino acid sequence from the [1st amino acid Met] 20th amino acid Cys to the 144th amino acid Leu of SEQ ID NO:3, or a modified human NR8-beta polypeptide comprising the amino acid sequence of said human NR8-beta polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said human NR8-beta polypeptide.

28. (Amended) The polypeptide of claim 23, wherein the polypeptide is human NR8-gamma comprising the amino acid sequence from the [1st amino acid Met] 20th amino acid Cys to the 538th amino acid Ser of SEQ ID NO:7, or a modified human NR8-gamma polypeptide comprising the amino acid sequence of said human NR8-gamma polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said human NR8-gamma polypeptide.

29. (Amended) The polypeptide of claim 23, wherein the polypeptide is mouse NR8-beta comprising the amino acid sequence from the [1st amino acid Met] 20th amino acid Cys to the 144th amino acid Leu of SEQ ID NO:19, or a modified mouse NR8-beta polypeptide comprising the amino acid sequence of said mouse NR8-beta polypeptide with one or more amino acids

NO:20, said polypeptide being functionally equivalent to a mouse NR8-beta polypeptide comprising the amino acid sequence from the [1st amino acid Met] 20th amino acid Cys to the 144th amino acid Leu of SEQ ID NO: 19.

36. (Amended) The polypeptide of claim 23, wherein the polypeptide is encoded by a nucleic acid that hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO: 22, said polypeptide being functionally equivalent to a mouse NR8-gamma polypeptide comprising the amino acid sequence from the [1st amino acid Met] 20th amino acid Cys to the 538th amino acid Ser of SEQ ID NO:21.

43. (Amended) The nucleic acid of claim 38, wherein the nucleic acid comprises the nucleotide sequence from the [441st nucleotide A] 498th nucleotide T to the 1523rd nucleotide C in SEQ ID NO:2; the nucleotide sequence from the [441st nucleotide A] 498th nucleotide T to the 872nd nucleotide A in SEQ ID NO:4; the nucleotide sequence from the 659th nucleotide A to the 1368th nucleotide C in SEQ ID NO:6; the nucleotide sequence from the [441st nucleotide A] 498th nucleotide T to the 2054th nucleotide C in SEQ ID NO:8; the nucleotide sequence from the [439th nucleotide A] 496th nucleotide T to the 870th nucleotide A in SEQ ID NO:20; or the nucleotide sequence from the [439th nucleotide A] 496th nucleotide T to the 2052nd nucleotide C in SEQ ID NO:22.

Please add new claim 51 as follows.

51. (New) A hemopoietin receptor NR8 polypeptide produced by the method of claim 46.

In the Drawings:

Please delete Figure 16.

Please delete Figure 17.

Please renumber Figure 18 as Figure 16, as shown in red in the attached copy.

Please renumber Figure 19 as Figure 17, as shown in red in the attached copy.

Applicant : Hitoshi Nomura et al.
Serial No. : 09/720,285
Filed : December 21, 2000
Page : 5

Attorney's Docket No.: 12660-002001 / C2-004PCT-
US

At the center top of each figure, please replace “/19” with --/17-- (for example, in Figure 12, replace “12/19” with --12/17--). In addition, please apply this scheme to renumbered Figures 16 and 17 (that is, “16/17” for renumbered Figure 16 and “17/17” for renumbered Figure 17).

REMARKS

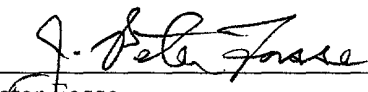
Applicants submit that all of the claims are now in condition for examination and allowance, which actions are requested.

The claim amendments are supported by the application as filed, and thus do not introduce new matter. In particular, the new claims recite polypeptide sequences without the 19 amino acid long signal sequences. Such polypeptides are described in the application at page 12, lines 13 to 19. The nucleic acids in claim 43 also remove the 57 nucleotides encoding the signal sequence.

Included herewith is a check to cover the fees for new claims added in this amendment. No other fees are believed due. However, please apply any charges or credits to Deposit Account No. 06-1050, referencing Attorney Docket No. 12660-002001.

Respectfully submitted,

Date: 2-6-01



J. Peter Fasse
Reg. No. 32,983

Fish & Richardson P.C.
225 Franklin Street
Boston, MA 02110-2804
Telephone: (617) 542-5070
Facsimile: (617) 542-8906

09/720285

Attorney's Docket No.: 12660-002001 / C2-004PCT-US

JC01 Rec'd PCT/PTO 2 1 DEC 2000

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Hitoshi Nomura et al. Art Unit : Unknown
Serial No. : New Examiner : Unknown
Filed : December 21, 2000
Title : NOVEL HEMOPOIETIN RECEPTOR PROTEINS

Commissioner for Patents
Washington, D.C. 20231

PRELIMINARY AMENDMENT

Prior to examination, please amend the application as follows.

In the Specification:

At page 1, line 10, delete "Art".

At page 3, line 4, replace "Disclosure" with --Summary--.

At page 7, after line 27, insert, at the left margin,

--Detailed Description--.

At page 31, line 29, replace "Best Mode for Carrying Out the Invention" with

--Examples--.

In the Claims:

Please cancel claims 1 to 21 without prejudice and add new claims 22 to 50 as follows.

22. (New) A purified hemopoietin receptor NR8 polypeptide.

23. (New) The polypeptide of claim 22, wherein the polypeptide is a human or mouse NR8 alpha, beta, or gamma polypeptide.

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December 21, 2000

Date of Deposit

Vicior Mahoney

Signature

Vicior Mahoney

Typed or Printed Name of Person Signing Certificate

24. (New) The polypeptide of claim 23, wherein the polypeptide is human NR8-alpha comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO:1, or a modified human NR8-alpha polypeptide comprising the amino acid sequence of said human NR8-alpha polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said human NR8-alpha polypeptide.

25. (New) The polypeptide of claim 23, wherein the polypeptide is human NR8-beta comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO:3, or a modified human NR8-beta polypeptide comprising the amino acid sequence of said human NR8-beta polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said human NR8-beta polypeptide.

26. (New) The polypeptide of claim 23, wherein the polypeptide is membrane-bound NR8-beta.

27. (New) The polypeptide of claim 26, wherein the polypeptide is human NR8-beta comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO:5, or a modified human NR8-beta polypeptide comprising the amino acid sequence of said human NR8-beta polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said human NR8-beta polypeptide.

28. (New) The polypeptide of claim 23, wherein the polypeptide is human NR8-gamma comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO:7, or a modified human NR8-gamma polypeptide comprising the amino acid sequence of said human NR8-gamma polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said human NR8-gamma polypeptide.

29. (New) The polypeptide of claim 23, wherein the polypeptide is mouse NR8-beta comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO:19, or a modified mouse NR8-beta polypeptide comprising the amino acid sequence of said mouse NR8-beta polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said mouse NR8-beta polypeptide.

30. (New) The polypeptide of claim 23, wherein the polypeptide is mouse NR8-gamma comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO:21, or a modified mouse NR8-gamma polypeptide comprising the amino acid sequence of said mouse NR8-gamma polypeptide with one or more amino acids deleted, added, or substituted with a different amino acid, and being functionally equivalent to said mouse NR8-gamma polypeptide.

31. (New) The polypeptide of claim 23, wherein the polypeptide is encoded by a nucleic acid that hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:2, said polypeptide being functionally equivalent to a human NR8-alpha polypeptide comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO:1.

32. (New) The polypeptide of claim 23, wherein the polypeptide is encoded by a nucleic acid that hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:4, said polypeptide being functionally equivalent to a human NR8-beta polypeptide comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO:3.

33. (New) The polypeptide of claim 23, wherein the polypeptide is encoded by a nucleic acid that hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:6, said polypeptide being functionally equivalent to a human membrane-bound NR8-beta polypeptide comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO:5.

34. (New) The polypeptide of claim 23, wherein the polypeptide is encoded by a nucleic acid that hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:8, said polypeptide being functionally equivalent to a human NR8-gamma polypeptide comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO:7.

35. (New) The polypeptide of claim 23, wherein the polypeptide is encoded by a nucleic acid that hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:20, said polypeptide being functionally equivalent to a mouse NR8-beta polypeptide comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO:19.

36. (New) The polypeptide of claim 23, wherein the polypeptide is encoded by a nucleic acid that hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO: 22, said polypeptide being functionally equivalent to a mouse NR8-gamma polypeptide comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO:21.

37. (New) A purified fusion polypeptide comprising the polypeptide of claim 22 and a second polypeptide or peptide.

38. (New) An isolated nucleic acid encoding a hemopoietin receptor NR8 polypeptide.

39. (New) The nucleic acid of claim 38, wherein the nucleic acid hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:2, 4, 6, or 8.

40. (New) The nucleic acid of claim 38, wherein the nucleic acid comprises the nucleotide sequence of SEQ ID NO:2, 4, 6, or 8.

41. (New) The nucleic acid of claim 38, wherein the nucleic acid hybridizes to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:20 or 22.

42. (New) The nucleic acid of claim 38, wherein the nucleic acid comprises the nucleotide sequence of SEQ ID NO:20 or 22, or is complementary to the nucleotide sequence of SEQ ID NO:20 or 21.

43. (New) The nucleic acid of claim 38, wherein the nucleic acid comprises the nucleotide sequence from the 441st nucleotide A to the 1523rd nucleotide C in SEQ ID NO:2; the nucleotide sequence from the 441st nucleotide A to the 872nd nucleotide A in SEQ ID NO:4; the nucleotide sequence from the 659th nucleotide A to the 1368th nucleotide C in SEQ ID NO:6; the nucleotide sequence from the 441st nucleotide A to the 2054th nucleotide C in SEQ ID NO:8; the nucleotide sequence from the 439th nucleotide A to the 870th nucleotide A in SEQ ID NO:20; or the nucleotide sequence from the 439th nucleotide A to the 2052nd nucleotide C in SEQ ID NO:22.

44. (New) A vector comprising the nucleic acid of claim 38.

45. (New) A cell comprising the nucleic acid of claim 43 and an expression regulating nucleotide sequence.

46. (New) A method of producing a hemopoietin receptor NR8 polypeptide, the method comprising culturing the cell of claim 45.

47. (New) A method of obtaining a compound that binds to the hemopoietin receptor NR8 polypeptide of claim 22, the method comprising

- (a) contacting a test sample with said polypeptide;
- (b) detecting a compound in the test sample that binds to said polypeptide; and
- (c) isolating the compound, thereby obtaining a compound that binds to said polypeptide.

48. (New) An antibody that specifically binds to the polypeptide of claim 22.

49. (New) A method of detecting a hemopoietin receptor NR8 polypeptide in a test sample, the method comprising
contacting a test sample with the antibody of claim 48 under conditions that enable formation of an antibody-NR8 polypeptide complex;
detecting an antibody-NR8 polypeptide complex, if present, wherein the presence of the complex indicates a hemopoietin receptor NR8 polypeptide in the test sample.

50. (New) A nucleic acid comprising a nucleotide sequence of at least 15 nucleotides and specifically hybridizing to a nucleic acid comprising a nucleotide sequence of any one of SEQ ID NOs:2, 4, 6, 8, 20, 22, 23, 24, 25, 26, or 27.

REMARKS

Applicants have replaced original claims 1 to 21 with new claims 22 to 50 to place the claims into a format more typical for United States prosecution and to avoid multiple dependent claims. Applicants submit that all of the claims are now in condition for examination, which action is requested. Applicants have submitted a filing fee that includes the excess claims fees required by the present preliminary amendment. No other fees are believed due. However, please apply any charges or credits to Deposit Account No. 06-1050, referencing Attorney Docket No. 12660-002001.

Respectfully submitted,

Date: December 21, 2000

J. Peter Fasse
J. Peter Fasse
Reg. No. 32,983

Fish & Richardson P.C.
225 Franklin Street
Boston, MA 02110-2804
Telephone: (617) 542-5070
Facsimile: (617) 542-8906

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09/720285

JCO1 Rec'd PCT/PTO 21 DEC 2000

DESCRIPTION

NOVEL HEMOPOIETIN RECEPTOR PROTEINS

5 Technical Field

The present invention relates to novel hemopoietin receptor proteins, the encoding genes, and methods of production and uses thereof.

10 Background Art

A large number of cytokines are known as humoral factors that are involved in the proliferation/differentiation of various cells, or activation of differentiated mature cells, and also cell death. These cytokines have their own specific receptors, which are categorized into several families based on their structural similarities (Hilton D.J., in "Guidebook to Cytokines and Their Receptors" edited by Nicola N.A. (A Sambrook & Tooze Publication at Oxford University Press), 1994, p8-16).

Compared to similarities between receptors, primary-structure homology is quite low between cytokines, and a significant amino acid homology cannot be seen even among cytokine members that belong to the same receptor family. This explains the functional specificity of each cytokine, as well as similarities of cellular reactions induced by each cytokine.

Representative examples of the above-mentioned receptor families are the tyrosine kinase receptor family, hemopoietin receptor family, tumor necrosis factor (TNF) receptor family, and transforming growth factor β (TGF β) receptor family. Different signal transduction pathways have been reported to be involved in each of these families. Among these receptor families, many receptors of especially the hemopoietin receptor family are expressed in blood cells and immunocytes, and their ligands, cytokines, are often termed as hemopoietic factors or interleukins. Some of these hemopoietic factors or interleukins exist within blood and are thought to be involved in a systemic humoral regulation of hemopoietic or immune functions.

This contrasts with the belief that cytokines belonging to other families are often involved in only topical regulations. Some of

these hemopoietins can be taken as hormone-like factors, and conversely, representative peptide hormones such as the growth hormone, prolactin, or leptin receptors also belong to the hemopoietin receptor family. Because of these hormone-like systemic regulatory features, it is anticipated that hemopoietin administration would be applied in the treatment of various diseases.

Among the large number of cytokines, those that are actually being clinically applied are, erythropoietin, G-CSF, GM-CSF, and IL-2. Combined with IL-11, LIF, and IL-12 that are being considered for clinical trials, and the above-mentioned peptide hormones such as growth hormone and prolactin, it can be envisaged that by searching among the above-mentioned various receptor families for a novel cytokine that binds to hemopoietin receptors, it is possible to find a cytokine that can be clinically applied with a higher efficiency.

As mentioned above, cytokine receptors have structural similarities between the family members. Using these similarities, many investigations are being carried out aiming at finding novel receptors. Regarding the tyrosine kinase receptor especially, many receptors have already been cloned using its highly conserved sequence at the catalytic site (Matthews W. et al., Cell, 1991, 65 (7) p1143-52). Compared to this, hemopoietin receptors do not have a tyrosine kinase-like enzyme activity domain in their cytoplasmic regions, and their signal transductions are known to be mediated through associations with other tyrosine kinase proteins existing freely in the cytoplasm.

Though the binding site on receptors associating with these cytoplasmic tyrosine kinases (JAK kinases) is conserved between family members, the homology is not very high (Murakami M. et al., Proc. Natl. Acad. Sci. USA, 1991, 88, 11349-11353). On one hand, the sequence that characterizes these hemopoietin receptors most well exists in the extracellular region, and especially the five amino acid Trp-Ser-Xaa-Trp-Ser (where Xaa is an arbitrary amino acid) motif is conserved in almost all of the hemopoietin receptors. Therefore, novel receptors are expected to be obtained by searching novel family members using this sequence. In fact, this approach has already identified the IL-11 receptor (Robb, L. et al., J. Biol. Chem., 1996, 271 (23) 13754-13761), leptin receptor (Gainsford T. et al., Proc. Natl. Acad. Sci. USA, 1996, 93 (25) p14564-8) and the IL-13 receptor

(Hilton D.J. et al., Proc. Natl. Acad. Sci. USA, 1996, 93 (1) p497-501).

Disclosure of the Invention

5 The present invention provides a novel hemopoietin receptor protein, and the encoding DNA. The present invention also provides, a vector into which the DNA has been inserted, a transformant harboring the DNA, and a method of producing a recombinant protein using the transformant. It also provides a method of screening a compound that
10 binds to the protein.

 Until now, the inventors have been trying to search for a novel receptor using an oligonucleotide encoding the Trp-Ser-Xaa-Trp-Ser motif as a probe by plaque hybridization, RT-PCR method, and so on. However, because of reasons such as the oligonucleotide tggag (t/c)
15 nnntggag (t/c) (where n is an arbitrary nucleotide) that encodes the motif being short having just 15 nucleotides, and the g/c being high, it was extremely difficult to strictly select only those in which the 15 nucleotides have completely hybridized under the usual hybridization conditions.

20 Also, a similar sequence is contained within cDNA encoding proteins other than hemopoietin receptors, starting with various collagens that are thought to be widely distributed and also have high expression amounts, which makes the screening by the above-mentioned plaque hybridization and RT-PCR highly inefficient.

25 To solve these problems, and to estimate how many different hemopoietic receptor genes actually exist on the human genome, the inventors computer-searched sequences that completely coincided with each probe using all capable oligonucleotide sequences encoding the above-mentioned Trp-Ser-Xaa-Trp-Ser motif as probes.

30 Next, among the clones identified by the above search, the nucleotide sequence around the probe sequence of human genome-derived clones (cosmid, BAC, PAC) was converted to the amino acid sequence and compared with the amino acid sequence of known hemopoietin receptors to select human genes thought to encode hemopoietin receptor
35 family members.

 From the above search, two clones thought to be hemopoietin receptor genes were identified. One of these was the known GM-CSF β receptor gene (derived from the 22q12.3-13.2 region of chromosome

no. 22), and the other (BAC clone AC002303 derived from the 16p12 region of chromosome no. 16) was presumed to encode a novel hemopoietin receptor protein, and this human gene was named "NR8."

Next, the cDNA thought to encode NR8 was found within the human fetal liver cell cDNA library by RT-PCR using a specific primer designed based on the obtained nucleotide sequence. Furthermore, using this cDNA library as the template, the full-length cDNA NR8 α encoding a transmembrane receptor comprising 361 amino acids was ultimately obtained by 5'-RACE method and 3'-RACE method.

In the primary structure of NR8 α , a cysteine residue and a proline rich motif conserved between other family members, were well conserved in the extracellular region, and in the intracellular region, the Box 1 motif thought to be involved in signal transduction was well conserved, and therefore, NR8 α was thought to be a typical hemopoietin receptor.

Furthermore, the inventors revealed the presence of two genes named NR8 β and NR8 γ as selective splicing products of NR8 α .

The inventors next attempted the isolation of the mouse gene corresponding to NR8 gene. First, using an oligonucleotide primer designed within human NR8 cDNA sequence and a mouse brain cDNA library as the template, xenogeneic cross PCR cloning was done to isolate the mouse partial nucleotide sequence of the above receptor. Furthermore, based on the obtained partial sequence, an oligonucleotide primer was designed, and using this, the inventors succeeded in isolating the full-length ORF of the mouse homologous gene corresponding to NR8 by the 5'-RACE method and 3'-RACE method. As a result of determining the whole nucleotide sequence of the obtained cDNA clone, alike NR8, the presence of mouse NR8 γ encoding a transmembrane receptor protein comprising 538 amino acids, and mouse NR8 β encoding a secretory, soluble receptor-like protein comprising 144 amino acids were confirmed by the difference of transcripts derived from the splice variant. When the amino acid sequences encoded by these receptor genes were compared between human and mouse, a high homology of 98.9% was observed for NR8 γ , and on the other hand, a homology of 97.2% was seen for NR8 β as well. Furthermore, the inventors succeeded in isolating the objective positive clones by plaque screening against a mouse genomic DNA library using the obtained mouse NR8 β cDNA fragment as the probe.

Therefore, the present invention provides:

- 5 (1) a protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1;
- 10 (2) a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3;
- 15 (3) a protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5;
- 20 (4) a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7;
- 30 (5) a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being
- 35

functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19;

- 5 (6) a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21;
- 10 (7) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 2, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1;
- 15 (8) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 4, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3;
- 20 (9) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 6, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5;
- 25 (10) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 8, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7;
- 30 (11) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 20, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19;
- 35 (12) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 22, said protein being

functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21;

- 5 (13) a fusion protein comprising the protein of any one of (1) to (12) and another peptide or polypeptide;
- (14) a DNA encoding the protein of any one of (1) to (13);
- (15) a vector comprising the DNA of (14);
- (16) a transformant harboring the DNA of (14) in an expressible manner;
- 10 (17) a method of producing the protein of any one of (1) to (13), comprising the step of culturing the transformant of (16);
- (18) a method of screening a compound that binds to the protein of any one of (1) to (13) comprising the steps of,
 - 15 (a) contacting a test sample with the protein of any one of (1) to (13), and
 - (b) selecting a compound that comprises an activity to bind to the protein of any one of (1) to (13);
- (19) an antibody that specifically binds to the protein of any one of (1) to (12);
- 20 (20) a method of detecting or measuring the protein of any one of (1) to (13) comprising the steps of contacting a test sample presumed to contain said protein with the antibody of (19), and detecting or measuring the formation of the immune complex between the antibody and the protein; and
- 25 (21) a DNA specifically hybridizing to a DNA comprising the nucleotide sequence of any one of SEQ ID NOS: 2, 4, 6, 8, 20, and 22 to 27, and comprising at least 15 nucleotides.

The present invention relates to the novel hemopoietin receptor "NR8." 5'-RACE and 3'-RACE analyses, NR8 genome sequence analysis, and plaque screening analysis revealed the presence of NR8 α , NR8 β , and NR8 γ . The structures of these NR8 genes are shown in Fig. 13. Among the NR8 genes, NR8 β is an alternative splicing product lacking the 5th exon, and can encode two different proteins, a soluble protein in which the CDS ends with a stop codon on the 6th exon that results from a frame shift following direct coupling to the 4th exon, and a membrane-bound protein lacking the signal sequence starting from the ATG upon the 4th exon.

Since the soluble protein comprises the same sequence as NR8 α up

to the 4th exon, it may function as a soluble receptor. On the other hand, NR8 γ encodes a protein containing a 177 amino acid insertion derived from the NR8 9th intron close to the C terminus of the NR8 α as a result of selective splicing.

5 Both NR8 α and NR8 γ encode transmembrane-type hemopoietin receptors. Among the sequences conserved between other hemopoietin receptors that are thought to be involved in signal transduction, a motif resembling Box 1 exists in the intracellular domain of NR8 α and NR8 γ adjacent to the cell membrane. Though low in the degree of
10 conservation, a sequence that is similar to Box 2 also exists, and therefore, NR8 is thought to be a type of receptor in which the signal is transduced by a homodimer.

The amino acid sequences of the NR8 proteins included in the proteins of the present invention are shown in SEQ ID NO: 1 (NR8 α),
15 SEQ ID NO: 3 (soluble NR8 β), SEQ ID NO: 5 (membrane-bound NR8 β), and SEQ ID NO: 7 (NR8 γ), and the nucleotide sequences of cDNA encoding these proteins are shown in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, and SEQ ID NO: 8, respectively.

Northern blot analysis for the spleen, thymus, peripheral
20 leucocytes, and lung showed two to three bands in the 5kb and 3 to 4kb regions. Similar sized bands were observed for cell lines HL60 and Raji also, but no expression was seen for other tumor cell lines (HeLa, SW480, A549, G361) and leukemia cell lines (K562, MOLT4).

The above results suggest that NR8 is specifically expressed on
25 hemopoietic cell lines, especially on granulocytic lines, and B cell lines.

The above NR8 protein is expected to be applied in medicine. NR8 is expressed in fetal liver, spleen, thymus, and some leukemic cell lines, suggesting the possibility that it might be a receptor of an
30 unknown hemopoietic factor. Therefore, NR8 protein would be a useful material for obtaining this unknown hemopoietic factor.

Furthermore, it is possible that NR8 is specifically expressed in limited cell populations within these hemopoietic tissues, and therefore, anti NR8 antibody may be useful as a means of separating
35 these cell populations. Thus separated cell populations can be applied for cell transplant therapy. Anti NR8 antibody is also expected to be applied for the diagnosis and treatment of leukemic diseases represented by leukemia.

On the other hand, the soluble protein including the extracellular domain of NR8 protein, or NR8 β , a splicing variant of NR8, may be applied as a decoy-type receptor that is an inhibitor of the NR8 ligand, and is anticipated to be applied in the treatment of diseases in which NR8 is involved, starting with leukemia.

The inventors also isolated mouse NR8 cDNA corresponding to the human-derived NR8 cDNA above-mentioned, by using the xenogeneic cross PCR cloning method. The amino acid sequences of the proteins named mouse NR8, which are included in the protein of the present invention are shown in SEQ ID NO: 19 (soluble mouse NR8 β) and SEQ ID NO: 21 (mouse NR8 γ), and the nucleotide sequences of the cDNA encoding these proteins are shown in SEQ ID NO: 20 and SEQ ID NO: 22, respectively.

As a result of structural analysis of the obtained mouse cDNA clones, alike human-derived NR8, the presence of mouse NR8 γ encoding a transmembrane receptor protein comprising 538 amino acids and mouse NR8 β encoding a secretory soluble receptor-like protein comprising 144 amino acids which were confirmed by the difference of transcripts derived the splice variant, was confirmed. When the amino acid sequences encoded by these receptor genes were compared between human and mouse, a high homology of 98.9% was observed for NR8 γ , while a homology of 97.2% was seen for NR8 β as well.

Northern blot analysis and RT-PCR analysis showed that although there were deviations in expression levels, mouse NR8 gene expression was seen in all organs analyzed, and seemed to be widely distributed compared to human NR8, for which a strong expression was seen only in immunocompetent and hemopoietic tissues. This also suggests the possibility that molecular functions of mouse NR8 may span a broad range of physiological regulatory mechanisms of the body.

The present invention also encompasses a protein that is functionally equivalent to the above-mentioned human or mouse NR8 protein. Herein "functionally equivalent" means having an equivalent biological activity to the above-mentioned NR8 proteins. Hemopoietic factor receptor protein activity can be given as an example of a biological activity. Such proteins can be obtained by the method of introducing a mutation to the amino acid sequence of a protein. For example, site-specific mutagenesis using a synthetic oligonucleotide primer, can be used to introduce a desired mutation into the amino acid sequence of a protein (Kramer, W. and Fritz, H.J.,

Methods in Enzymol., 1987, 154, 350-367). This could also be done by a PCR-mediated site-specific mutagenesis system (GIBCO-BRL). Using these methods, the amino acid sequence of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, or SEQ ID NO: 21 can be modified to obtain a protein functionally equivalent to the NR8 protein, in which one or more amino acids in the amino acid sequence of the protein have been deleted, added, and/or substituted by another amino acid without affecting the biological activity of the protein.

- 10 As a protein functionally equivalent to the NR8 protein of the invention, the following are given: one in which one or two or more, preferably, two to 30, more preferably, two to ten amino acids are deleted in any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, or SEQ ID NO: 21; one in which one or two or more, preferably, two to 30, more preferably, two to ten amino acids have been added into any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, or SEQ ID NO: 7; or one in which one or two or more, preferably, two to 30, more preferably, two to ten amino acids have been substituted with other amino acids in any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, or SEQ ID NO: 7.

- It is already known that a protein comprising a modified amino acid sequence of a certain amino acid sequence in which one or more amino acid residues have been deleted, added, and/or substituted with another amino acid, still maintains its biological activity (Mark, D. F. et al., Proc. Natl. Acad. Sci. USA, 1984, 81, 5662-5666; Zoller, M. J. & Smith, M., Nucleic Acids Research, 1982, 10, 6487-6500; Wang, A. et al., Science, 224, 1431-1433; Dalbadie-McFarland, G. et al., Proc. Natl. Acad. Sci. USA, 1982, 79, 6409-6413).

- For example, a fusion protein can be given as a protein in which one or more amino acid residues have been added to the NR8 protein of the present invention. A fusion protein is made by fusing the NR8 protein of the present invention with another peptide or protein and is encompassed in the present invention. A fusion protein can be prepared by ligating DNA encoding the NR8 protein of the present invention with DNA encoding another peptide or protein so as the frames match, introducing this into an expression vector, and expressing

the fusion gene in a host. Methods commonly known can be used for preparing such a fusion gene. There is no restriction as to the other peptide or protein that is fused to the protein of this invention.

For example, FLAG (Hopp, T.P. et al., Biotechnology, 1988, 6, 1204-1210), 6x His constituting six histidine (His) residues, 10x His, Influenza agglutinin (HA), human c-myc fragment, VSV-GP fragment, p18HIV fragment, T7-tag, HSV-tag, E-tag, SV40T antigen fragment, lck tag, α -tubulin fragment, B-tag, Protein C fragment, and such well-known peptides can be used. Examples of proteins are, glutathione-S-transferase (GST), Influenza agglutinin (HA), immunoglobulin constant region, β -galactosidase, maltose-binding protein (MBP), etc. Commercially available DNAs encoding these may also be used to prepare fusion proteins.

The protein of the invention can also be encoded by a DNA that hybridizes under stringent conditions to a DNA comprising any one of the nucleotide sequences of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 20, and SEQ ID NO: 22 to 27. Such a protein also includes a protein functionally equivalent to the above-mentioned NR8 protein. Stringent conditions can be suitably selected by one skilled in the art, and for example, low-stringent conditions can be given. Low-stringent conditions are, for example, 42°C, 2x SSC, and 0.1% SDS, and preferably, 50°C, 2x SSC, and 0.1% SDS. More preferable are highly stringent conditions, for example, 65°C, 2x SSC, and 0.1% SDS. Under these conditions, the higher the temperature is raised, the higher the homology of the obtained DNA will be.

The present invention also includes a protein that is functionally equivalent to the above NR8 protein, which has also a homology with a protein comprising any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, or SEQ ID NO: 21. A protein having a homology means, a protein having at least 70%, preferably at least 80%, more preferably at least 90%, even more preferably, at least 95% homology to any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, and SEQ ID NO: 7. The homology of a protein can be determined by the algorithm in "Wilbur, W.J. and Lipman, D.J. Proc. Natl. Acad. Sci. USA, 1983, 80, 726-730."

In the protein of the invention, the amino acid sequence, molecular

weight, isoelectric point, the presence or absence of the sugar chain, and its form differ according to the producing cells, host, or purification method described below. However, as long as the obtained protein comprises a hemopoietic factor receptor protein activity, it is included in the present invention.

For example, if the protein of the present invention is expressed in prokaryotic cells such as *E. coli*, a methionine residue is added at the N-terminus of the amino acid sequence of the expressed protein. If the protein of the present invention is expressed in eukaryotic cells such as mammalian cells, the N-terminal signal sequence is removed. The protein of the present invention includes these proteins.

For example, as a result of analyzing the protein of the invention based on the method in "Von Heijne, G., *Nucleic Acids Research*, 1986, 14, 4683-4690," it was presumed that the signal sequence is from the 1st Met to the 19th Gly in the amino acid sequence of SEQ ID NO: 1. Therefore, the present invention encompasses a protein comprising the sequence from the 20th Cys to 361st Ser in the amino acid sequence of SEQ ID NO: 1.

To produce the protein of the invention, the obtained DNA is incorporated into an expression vector in a manner that the DNA is expressible under the regulation of an expression regulatory region, for example, an enhancer or promoter. Next, host cells are transformed by this expression vector to express the protein.

Specifically, the protein can be produced as follows. When mammalian cells are used, DNA comprising a commonly used useful promoter/enhancer, DNA encoding the protein of the invention, and the poly A signal that is functionally bound to the 3' side downstream of the protein-encoding DNA, or a vector containing it, is constructed. For example, as the promoter/enhancer, human cytomegalovirus immediate early promoter/enhancer can be given.

Also, as other promoters/enhancers that can be used for protein expression, viral promoters/enhancers of retroviruses, polyomaviruses, adenoviruses, simian virus 40 (SV40), and such, and promoters/enhancers derived from mammalian cells, such as that of human elongation factor 1 α (HEF1 α) can be used.

For example, a protein can be easily expressed by following the method of Mulligan et al. (*Nature*, 1979, 277, 108) when using the

SV40 promoter/enhancer, and the method of Mizushima et al. (Nucleic Acids Res., 1990, 18, 5322) when using the HEFl α promoter/enhancer.

When using *E. coli*, well-used useful promoters, the signal sequence for polypeptide secretion, and genes to be expressed, may
 5 be functionally bound to express the desired gene. For example, lacZ promoter and araB promoter may be used as promoters. When using the lacZ promoter, the method of Ward et al. (Nature, 1098, 341, 544-546; FASEB J., 1992, 6, 2422-2427), and when using the araB promoter, the method of Better et al. (Science, 1988, 240, 1041-1043) may be
 10 followed.

When producing the protein into the periplasm of *E. coli*, the pelB (Lei, S. P. et al., J. Bacteriol., 1987, 169, 4379) signal sequence may be used as a protein secretion signal.

A replication origin derived from SV40, polyomavirus, adenovirus,
 15 bovine papillomavirus (BPV), and such may be used. To amplify gene copies in host cell lines, the expression vector may include an aminoglycoside transferase (APH) gene, thymidine kinase (TK) gene, *E. coli* xanthine guanine phosphoribosyl transferase (Ecogpt) gene, dihydrofolate reductase (dhfr) gene, and such as a selective marker.

20 The expression vector used to produce the protein of the invention may be any, as long as it's an expression vector that is suitably used for the present invention. Mammalian expression vectors, for example, pEF and pCDM8; insect-derived expression vectors, for example, pBacPAK8; plant-derived expression vectors, for example,
 25 pMH1 and pMH2; animal virus-derived expression vectors, for example, pHSV, pMV, and pAdexLcw; retrovirus-derived expression vectors, for example, pZIpneo; yeast-derived expression vectors, for example, pNV11 and SP-Q01; *Bacillus subtilis*-derived expression vectors, for example, pPL608 and pKTH50; *E. coli*-derived expression vectors, for
 30 example, pQE, pGEAPP, pGEMEAPP, and pMALp2 can be given as expression vectors of this invention.

Not only vectors that produce the protein of the invention *in vivo* and *in vitro*, but also those that are used for gene therapy of mammals, for example humans, are also included as vectors of the present
 35 invention.

When introducing the expression vector of the present invention constructed above into a host cell, well-known methods, for example the calcium phosphate method (Virology, 1973, 52, 456-467),

electroporation (EMBO J., 1982, 1, 841-845), and such may be used.

In the present invention, an arbitrary production system may be used to produce the protein. *In vitro* and *in vivo* production systems are known as production systems for producing proteins. Production
 5 systems using eukaryotic cells and prokaryotic cells may be used as *in vitro* production systems.

When using eukaryotic cells, production systems using, for example, animal cells, plant cells, and fungal cells are known. As animal cells used, for example, mammalian cells such as CHO (J. Exp. Med., 1995,
 10 108, 945), COS, myeloma, baby hamster kidney (BHK), HeLa, or Vero, amphibian cells such as *Xenopus* oocytes (Valle, et al., Nature, 1981, 291, 358-340), insect cells such as sf9, sf21, or Tn5, are known. As CHO cells, especially DHFR gene-deficient CHO cell, dhfr-CHO (Proc. Natl. Acad. Sci. USA, 1980, 77, 4216-4220), and CHO K-1 (Proc. Natl.
 15 Acad. Sci. USA, 1968, 60, 1275) can be suitably used.

Nicotiana tabacum-derived cells are well known as plant cells, and these can be callus cultured. As fungal cells, yeasts such as the *Saccharomyces* genus, for example, *Saccharomyces cerevisiae*, filamentous bacteria such as the *Aspergillus* genus, for example,
 20 *Aspergillus niger* are known.

Bacterial cells may be used as prokaryotic production systems. As bacterial cells, *E. coli* and *Bacillus subtilis* are known.

Proteins can be obtained by transforming these cells with the objective DNA, and culturing the transformed cells *in vitro* according
 25 to well-known methods. For example, DMEM, MEM, RPMI1640, and IMDM can be used as culture media. At that instance, fetal calf serum (FCS) and such serum supplements may be added in the above media, or a serum-free culture medium may be used. The pH is preferably about 6 to 8. Culture is usually done at about 30°C to 40°C, for about 15
 30 to 200 hr, and medium changes, aeration, and stirring are done as necessary.

On the other hand, production systems using animals and plants may be given as *in vivo* production systems. The objective gene is introduced into the plant or animal, and the protein is produced within
 35 the plant or animal, and recovered. "Host" as used in the present invention encompasses such animals and plants as well.

When using animals, mammalian and insect production systems can be used. As mammals, goats, pigs, sheep, mice, and cattle may be used

(Vicki Glaser, SPECTRUM Biotechnology Applications, 1993). Transgenic animals may also be used when using mammals.

For example, the objective DNA is inserted within a gene encoding a protein produced intrinsically into milk, such as goat β casein, to prepare a fusion gene. The DNA fragment containing the fusion gene is injected into a goat's embryo, and this embryo is implanted in a female goat. The protein is collected from the milk of the transgenic goats produced from the goat that received the embryo, and descendents thereof. To increase the amount of protein-containing milk produced from the transgenic goat, a suitable hormone/hormones may be given to the transgenic goats (Ebert, K.M. et al., Bio/Technology, 1994, 12, 699-702).

Silk worms may be used as insects. When using the silk worm, it is infected with a baculovirus to which the objective DNA has been inserted, and the desired protein is obtained from the body fluids of the silk worm (Susumu, M. et al., Nature, 1985, 315, 592-594).

When using plants, for example, tobacco can be used. In the case of tobacco, the objective DNA is inserted into a plant expression vector, for example pMON 530, and this vector is introduced into a bacterium such as *Agrobacterium tumefaciens*. This bacterium is infected to tobacco, for example *Nicotiana tabacum*, to obtain the desired polypeptide from tobacco leaves (Julian, K.-C. Ma et al., Eur. J. Immunol., 1994, 24, 131-138).

The thus-obtained protein of the invention is isolated from within and without cells, or from hosts, and can be purified as a substantially pure homogenous protein. The separation and purification of the protein is not limited to any specific method and can be done using ordinary separation and purification methods used to purify proteins. For example, chromatography, filtration, ultrafiltration, salting out, solvent precipitation, solvent extraction, distillation, immunoprecipitation, SDS-polyacrylamide gel electrophoresis, isoelectric focusing, dialysis, recrystallization, and such may be suitably selected, or combined to separate/purify the protein.

As chromatographies, for example, affinity chromatography, ion exchange chromatography, hydrophobic chromatography, gel filtration, reversed-phase chromatography, adsorption chromatography, and such can be exemplified (Strategies for Protein Purification and

Characterization: A Laboratory Course Manual. Ed Daniel R. Marshak et al., Cold Spring Harbor Laboratory Press, 1996). These chromatographies can be done by liquid chromatography such as HPLC, FPLC, and the like. The present invention encompasses proteins
5 highly purified by using such purification methods.

Proteins can be arbitrarily modified, or peptides may be partially excised by treating the proteins with appropriate modification enzymes prior to or after the purification. Trypsin, chymotrypsin, lysyl endopeptidase, protein kinase, glucosidase, and such are used
10 as protein modification enzymes.

The present invention includes a partial peptide comprising the active center of a protein comprising any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, and SEQ ID NO: 21. A partial peptide of the protein
15 of the present invention is, for example, a partial peptide of the molecules of the protein, which contains one or more regions of the hydrophilic region and hydrophobic region presumed by hydrophobicity plot analysis. These partial peptides may contain the whole hydrophilic region or a part of it, and may contain the whole
20 hydrophobic region or a part of it. For example, soluble proteins and proteins comprising extracellular regions of the protein of the invention, are also encompassed in the invention.

The partial peptides of the protein of the invention may be produced by genetic engineering techniques, well-known peptide synthesizing
25 methods, or by excising the protein of the invention by a suitable peptidase. As peptide synthesizing methods, the solid-phase synthesizing method, and the liquid-phase synthesizing method may be used.

The present invention also relates to a DNA encoding the protein
30 of the invention. A cDNA encoding the protein of the invention may be obtained by, for example, screening a human cDNA library using the probe described herein.

Using the obtained cDNA or cDNA fragment as a probe, cDNA can also be obtained from other cells, tissues, organs, or species by further
35 screening cDNA libraries. cDNA libraries may be prepared by, for example, the method of Sambrook, J. et al., Molecular Cloning, Cold Spring Harbor Laboratory Press (1989), or commercially available cDNA libraries may be used.

By determining the nucleotide sequence of the obtained cDNA, the translation region encoded by it can be determined, and the amino acid sequence of the protein of the present invention can be obtained. Furthermore, genomic DNA can be isolated by screening the genomic
 5 DNA library using the obtained cDNA as a probe.

Specifically, this can be done as follows. First, mRNA is isolated from cells, tissues, and organs expressing the protein of the invention. For this mRNA isolation, whole RNA is prepared using well-known methods, for example, guanidine ultracentrifugation
 10 method (Chirgwin, J.M. et al., Biochemistry, 1979, 18, 5294-5299), the AGPC method (Chomczynski, P. and Sacchi, N., Anal. Biochem., 1987, 162, 156-159), and such, and purified using the mRNA Purification Kit (Pharmacia), etc. mRNA may be directly prepared using the QuickPrep mRNA Purification Kit (Pharmacia).

15 cDNA is synthesized using reverse transcriptase from the obtained mRNA. cDNA can be synthesized using the AMV Reverse Transcriptase First-strand cDNA Synthesis Kit (SEIKAGAKU CORPORATION), etc. Also, cDNA synthesis and amplification may also be done using the probe described herein by following the 5'-RACE method (Frohman, M.A. et
 20 al., Proc. Natl. Acad. Sci. U.S.A., 1988, 85, 8998-9002; Belyavsky, A. et al., Nucleic Acids Res., 1989, 17, 2919-2932) using the polymerase chain reaction (PCR) and the 5'-Ampli FINDER RACE KIT (Clontech).

The objective DNA fragment is prepared from the obtained PCR
 25 product and ligated with vector DNA. Thus, a recombination vector is created, introduced into *E.coli*, etc. and colonies are selected to prepare the desired recombination vector. The nucleotide sequence of the objective DNA may be verified by known methods, for example, the dideoxy nucleotide chain termination method.

30 In the DNA of the invention, a sequence with a higher expression efficiency can be designed by considering the codon usage frequency of hosts used for the expression (Grantham, R. et al., Nucleic Acids Research, 1981, 9, p43-p74). The DNA of the invention may also be modified using commercially available kits and known methods. For
 35 example, digestion by restriction enzymes, insertion of synthetic oligonucleotides and suitable DNA fragments, addition of linkers, insertion of a start codon (ATG) and/or stop codon (ATT, TGA, or TAG), and such can be given.

The DNA of the present invention encompasses DNA comprising the nucleotide sequence from the 441st nucleotide A to the 1523rd nucleotide C in the nucleotide sequence of SEQ ID NO: 2, DNA comprising the nucleotide sequence from the 441st nucleotide A to the 872nd nucleotide A in the nucleotide sequence of SEQ ID NO: 4, DNA comprising the nucleotide sequence from the 659th nucleotide A to the 1368th nucleotide C in the nucleotide sequence of SEQ ID NO: 6, DNA comprising the nucleotide sequence from the 441st nucleotide A to the 2054th nucleotide C in the nucleotide sequence of SEQ ID NO: 8, DNA comprising the nucleotide sequence from the 439th nucleotide A to the 870th nucleotide A in the nucleotide sequence of SEQ ID NO: 20, and DNA comprising the nucleotide sequence from the 439th nucleotide A to the 2052nd nucleotide C in the nucleotide sequence of SEQ ID NO: 22.

The DNA of the present invention encompasses DNA that hybridizes under stringent conditions to the DNA comprising any one of the nucleotide sequences of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 20, and SEQ ID NO: 22 to 27, which also includes a DNA encoding a protein functionally equivalent to the NR8 protein.

Stringent conditions can be suitably selected by one skilled in the art, and for example, low-stringent conditions can be given. Low-stringent conditions are, for example, 42°C, 2x SSC, and 0.1% SDS, and preferably 50°C, 2x SSC, and 0.1% SDS. More preferable are highly stringent conditions, for example, 65°C, 2x SSC, and 0.1% SDS. Under these conditions, the higher the temperature is raised, the higher the homology of the obtained DNA will be. The above DNA is preferably natural DNA such as cDNA and chromosomal DNA.

As shown in Examples, the mRNA of the gene hybridizing to cDNA encoding the protein of the invention was distributed in various human tissues. Therefore, the above-mentioned natural DNA may be, for example, genomic DNA and cDNA derived from tissues in which the mRNA that hybridizes to the cDNA encoding the protein of the invention is detected in Examples. The DNA encoding the protein of the invention may be cDNA, genomic DNA, or synthetic DNA.

The protein of the invention is useful in screening a compound that binds to it. Namely, the protein of the invention is used in the screening method that comprises the steps of contacting a test sample expected to contain a compound that binds to the protein of the invention with the protein of the invention, and selecting the

compound that comprises an activity to bind to the protein of the invention.

As methods for screening a compound that comprises an activity to bind to the protein of the invention, numerous methods usually used by those skilled in the art can be employed. The protein of the invention that is used for the screening of the invention may be a recombinant, natural, or partial peptide. A compound comprising an activity to bind to the protein of the invention may be a protein comprising a binding activity, or it may be a chemically synthesized compound having a binding activity.

As a test sample that is used in the screening method of the present invention, for example, peptides, purified or crudely purified proteins, non-peptide compounds, synthetic compounds, microbial fermentation products, extracts of marine organisms, plant extracts, cell extracts, animal tissue extracts, and such can be given. These test samples may be novel compounds, or well-known compounds.

A protein that binds to the protein of the invention can be screened by, for example, using the West-western blotting method (Skolnik, E.Y. et al., Cell, 1991, 65, 83-90). cDNA is isolated from cells, tissues, and organs presumed to express the protein binding to the protein of the invention, this is inserted into phage vectors, for example, λ gt11, ZAPII, and such, to make a cDNA library, expressed on a plate containing a culture medium, the proteins expressed are fixed on a filter, this filter is reacted with the labeled, purified protein of the invention, and plaques expressing the protein bound to the protein of the invention are detected by the labels. As methods to label the protein of the invention, the method that uses the binding ability of avidin and biotin, the method of using an antibody that specifically binds to the protein of the invention or the peptide or polypeptide fused to the protein of the invention, the method of using radioisotopes, or fluorescence, and such can be given.

A ligand that binds specifically to the protein of the invention can be screened by, preparing a chimeric receptor by ligating the extracellular domain of the protein of the invention with the intracellular domain containing the transmembrane domain of a hemopoietin receptor protein comprising a known signal transduction ability, expressing this chimeric receptor on the cell surface of a suitable cell line, preferably, a cell line that can survive and

proliferate under the presence of a suitable growth factor (a growth factor-dependent cell line), and culturing the cell line by adding a material that is expected to contain various growth factors, cytokines, or hemopoietic factors. This method uses the fact that

5 the above-mentioned growth factor-dependent cell line survives and proliferates only when a ligand that specifically binds to the extracellular domain of the protein of the invention exists within the test material. Known hemopoietic receptors are, for example, the thrombopoietin receptor, erythropoietin receptor, G-CSF receptor,

10 gp130, etc. However, the partner of the chimeric receptor used in the screening of the invention is not limited to these known hemopoietic receptors, and any may be used as long as a structure needed for the signal transduction activity is contained in the cytoplasmic domain. Growth factor-dependent cell lines are for

15 example, IL-3-dependent cell lines starting with BaF3 and FDC-P1.

As a ligand that specifically binds to the protein of the invention, the possibility of not only soluble proteins, but also cell membrane-binding proteins can be envisaged, though rare. In such cases, screening can be done by labeling the protein containing only

20 the extracellular domain of the protein of the invention, or a fusion protein in which the partial sequence of another soluble protein has been added to this extracellular domain, and measuring the binding with cells expected to express the ligand. As examples of proteins containing only the extracellular domain of the protein of the

25 invention, for example, a soluble receptor protein artificially made by inserting a stop codon to the N terminal side of the transmembrane domain, or NR8 β soluble protein may be used. On the other hand, as a fusion protein in which the partial sequence of another soluble protein has been added to the extracellular domain of the protein

30 of the invention, for example, proteins prepared by adding immunoglobulin Fc site, FLAG peptide, etc. to the C terminus of the extracellular domain can be used. These soluble labeled proteins can be used in the detection in the above-described West-western blotting method.

35 A protein that binds to the protein of the invention can be screened by using the two-hybrid system (Fields, S. and Sternglanz, R., Trends. Genet., 1994, 10, 286-292).

In the two-hybrid system, an expression vector containing DNA

encoding the fusion protein between the protein of the invention and one subunit of a heterodimeric transcriptional regulatory factor, and an expression vector containing DNA made by ligating DNA encoding the other subunit of the heterodimeric transcriptional regulatory factor and a desired cDNA used as a test sample are introduced into
 5 cells and expressed. If the protein encoded by the cDNA binds with the protein of the invention and the transcriptional regulatory factor forms a heterodimer, a reporter gene constructed in the cell beforehand will be expressed. Therefore, a protein binding to the
 10 protein of the invention can be selected by detecting or measuring the expression level of the reporter gene.

Specifically, the DNA encoding the protein of the invention and the gene encoding the DNA binding domain of LexA are ligated so as the frames match to prepare an expression vector. Next, the desired
 15 cDNA and the gene encoding GAL4 transcription activation domain are ligated to prepare an expression vector.

Cells into which the HIS3 gene has been incorporated (the transcription of HIS3 gene is regulated by the promoter having a LexA binding motif) are transformed by the above two-hybrid system
 20 expression plasmids, and then incubated on a histidine-free synthetic culture medium. Herein, cells only grow when a protein interaction is present. Thus, the increase in reporter gene expression can be examined by the growth rate of the transformant.

Other than the HIS3 gene, for example, the luciferase gene, plasminogen activator inhibitor type1 (PAI-1) gene, ADE2 gene, LacZ
 25 gene, CDC25H gene, and such can be used as reporter genes.

The two-hybrid system may be constructed according to the usual methods, or a commercially available kit may be used. As commercially available two-hybrid system kits, the MATCHMARKER Two-Hybrid System,
 30 Mammalian MATCHMARKER Two-Hybrid Assay Kit (both by CLONTEC), HybriZAP Two-Hybrid Vector System (Stratagene), and CytoTrap two-hybrid system (Stratagene) can be given.

A protein binding to the protein of the invention can be screened by affinity chromatography. Namely, the protein of the invention is
 35 immobilized onto a carrier of an affinity column, and a test sample presumed to express a protein binding to the protein of the invention is applied to the column. As this test sample, a cell culture supernatant, cell extract, cell lysate, and such may be used. After

applying the test sample, the column is washed to obtain the protein binding to the protein of the invention.

5 The compound isolated by the screening method of the invention is a candidate drug for promoting or inhibiting the activity of the protein of the invention. The compound obtained by using the screening method of the invention encompasses a compound resulting from modifying the compound having an activity to bind to the protein of the invention by adding, deleting, and/or replacing a part of the structure.

10 When using the compound obtained by the screening method of the invention as drugs for humans and other mammals such as, mice, rats, guinea pigs, rabbits, chicken, cats, dogs, sheep, pigs, cattle, monkeys, sacred baboons, and chimpanzees, the drug may be administered using ordinary means.

15 For example, according to the need, the drugs can be taken orally as sugar-coated tablets, capsules, elixirs, and microcapsules, or parenterally in the form of injections of sterile solutions or suspensions with water or any other pharmaceutically acceptable liquid. For example, the compounds comprising the activity to bind to the protein of the invention can be mixed with physiologically acceptable carriers, flavoring agents, excipients, vehicles, preservatives, stabilizers, and binders, in a unit dose form required for generally accepted drug implementation. The amount of active ingredients in these preparations makes a suitable dosage within the indicated range acquirable.

25 Examples of additives that can be mixed to tablets and capsules are, binders such as gelatin, corn starch, tragacanth gum, and arabic gum; excipients such as crystalline cellulose; swelling agents such as cornstarch, gelatin, and alginic acid; lubricants such as magnesium stearate; sweeteners such as sucrose, lactose, or saccharin; and flavoring agents such as peppermint, Gaultheria adeno-thrix oil, and cherry. When the unit dosage form is a capsule, a liquid carrier, such as oil, can also be included in the above additives. Sterile compositions for injections can be formulated following usual drug implementations using vehicles such as distilled water used for injections.

35 For example, physiological saline and isotonic liquids including glucose or other adjuvants, such as D-sorbitol, D-mannose, D-mannitol,

and sodium chloride, can be used as aqueous solutions for injections. These can be used in conjunction with suitable solubilizers, such as alcohol, specifically ethanol, polyalcohols such as propylene glycol and polyethylene glycol, non-ionic surfactants, such as
5 Polysorbate 80 (TM) and HCO-50.

Sesame oil or soy-bean oil can be used as a oleaginous liquid and may be used in conjunction with benzyl benzoate or benzyl alcohol as a solubilizer; may be formulated with a buffer such as phosphate buffer and sodium acetate buffer; a pain-killer such as procaine
10 hydrochloride; a stabilizer such as benzyl alcohol and phenol; and an anti-oxidant. The prepared injection is usually filled into a suitable ampule.

Although the dosage of the compound that has the activity to bind to the protein of the invention varies according to symptoms, the
15 daily dose is generally about 0.1 to about 100 mg, preferably about 1.0 to about 50 mg, and more preferably about 1.0 to about 20 mg, when administered orally to an adult (body weight 60 kg).

When given parenterally, the dose differs according to the patient, target organ, symptoms, and method of administration, but the daily
20 dose is usually about 0.01 to about 30 mg, preferably about 0.1 to about 20 mg and more preferably about 0.1 to about 10 mg for an adult (body weight 60 kg) when given as an intravenous injection. Also, in the case of other animals too, it is possible to administer an amount converted to 60 kg of body-weight.

25 The antibody of the present invention can be obtained as a monoclonal antibody or a polyclonal antibody using well-known methods.

The antibody that specifically binds to the protein of the invention can be prepared by using the protein of the invention as
30 a sensitizing antigen for immunization according to usual immunizing methods, fusing the obtained immunized cells with known parent cells by ordinary cell fusion methods, and screening for antibody producing cells using the usual screening techniques.

Specifically, a monoclonal or polyclonal antibody that binds to
35 the proteins of the invention may be prepared as follows.

For example, the protein of the invention that is used as a sensitizing antigen for obtaining the antibody is not restricted by the animal species from which it is derived, but is preferably a

protein derived from mammals, for example, humans, mice, or rats, especially from humans. Proteins of human origin can be obtained by using the nucleotide sequence or amino acid sequence disclosed herein.

- 5 The protein that is used as a sensitizing antigen in the present invention can be a protein that comprises the biological activity of all the proteins described herein. Partial peptides of the proteins may also be used. As partial peptides of the proteins, for example, the amino (N) terminal fragment of the protein, and the
10 carboxy (C) terminal fragment can be given. "Antibody" as used herein means an antibody that specifically reacts with the full-length or fragment of the protein.

- A gene encoding the protein of the invention or a fragment thereof is inserted into a well-known expression vector, and after
15 transforming the host cells described herein, the objective protein or a fragment thereof is obtained from within and without the host cell, or from the host using well-known methods, and this protein can be used as a sensitizing antigen. Also, cells expressing the protein, cell lysates, or chemically synthesized protein of the
20 invention may be used as a sensitizing antigen.

- The mammals that are immunized by the sensitizing antigen are not restricted, but it is preferable to select the animal by considering the adaptability with the parent cells used in cell fusion. Generally, an animal belonging to Rodentia, Lagomorpha, or Primates is used.
25 As animals belonging to Rodentia, for example, mice, rats, hamsters, and such are used. As animals belonging to Lagomorpha, for example rabbits, as Primates, for example monkeys, are used. As monkeys, monkeys of the infraorder Catarrhini (Old World Monkeys), for example, cynomolgus monkeys, rhesus monkeys, sacred baboons,
30 chimpanzees, etc., are used.

- To immunize animals with the sensitizing antigen, well-known methods may be used. For example, the sensitizing antigen is generally injected into mammals intraperitoneally or subcutaneously. Specifically, the sensitizing antigen is suitably diluted, suspended
35 in physiological saline or phosphate-buffered saline (PBS), mixed with a suitable amount of a general adjuvant if desired, for example, with Freund's complete adjuvant, emulsified and injected into the mammal. Thereafter, the sensitizing antigen suitably mixed with

Freund's incomplete adjuvant is preferably given several times every four to 21 days. A suitable carrier can also be used when immunizing an animal with the sensitizing antigen. After the immunization, the elevation in the serum antibody level is detected by usual methods.

5 Polyclonal antibodies against the protein of the invention can be obtained as follows. After verifying that the desired serum antibody level has been reached, blood is withdrawn from the mammal sensitized with the antigen. Serum is isolated from this blood using well-known methods. The serum containing the polyclonal antibody may
10 be used as the polyclonal antibody, or according to needs, the polyclonal antibody-containing fraction may be further isolated from the serum.

To obtain monoclonal antibodies, after verifying that the desired serum antibody level has been reached in the mammal sensitized with
15 the above-described antigen, immunocytes are taken from the mammal and used for cell fusion. At this instance, immunocytes that are preferably used for cell fusion are splenocytes. As parent cells fused with the above immunocytes, preferable are mammalian myeloma cells, more preferable are, myeloma cells that have attained the
20 feature of distinguishing fusion cells by agents.

For the cell fusion between the above immunocytes and myeloma cells, for example, the method of Milstein et al. (Galfre, G. and Milstein, C., Methods Enzymol., 1981, 73, 3-46) is basically well known.

The hybridoma obtained from cell fusion is selected by culturing
25 in a usual selective culture medium, for example, HAT culture medium (hypoxanthine, aminopterin, thymidine-containing culture medium). The culture in this HAT medium is continued for a period sufficient enough for cells (non-fusion cells) other than the objective hybridoma to perish, usually from a few days to a few weeks. Next, the usual
30 limiting dilution method is carried out, and the hybridoma producing the objective antibody is screened and cloned.

Other than the above method of obtaining a hybridoma by immunizing an animal other than humans with the antigen, a hybridoma producing the objective human antibodies comprising the activity to bind to
35 proteins can be obtained by the method of sensitizing human lymphocytes, for example, human lymphocytes infected with the EB virus, with proteins, protein-expressing cells, or lysates thereof *in vitro*, fusing the sensitized lymphocytes with myeloma cells derived from

human, for example U266, having the capacity of permanent cell division (Unexamined Published Japanese Patent Application (JP-A) No. Sho 63-17688).

Moreover, human antibody against the protein can be obtained using
 5 a hybridoma made by fusing myeloma cells with antibody-producing cells obtained by immunizing a transgenic animal comprising a repertoire of human antibody genes with an antigen such as a protein, protein-expressing cells, or a cell lysate thereof WO92/03918, WO93/2227, WO94/02602, WO94/25585, WO96/33735, and WO96/34096).

10 Other than producing antibodies by using hybridoma, antibody-producing immunocytes such as sensitized lymphocytes that are immortalized by oncogenes may also be used.

Such monoclonal antibodies can also be obtained as recombinant antibodies produced by using the gene engineering technique (for
 15 example, Borrebaeck, C.A.K. and Larrick, J.W., THERAPEUTIC MONOCLONAL ANTIBODIES, Published in the United Kingdom by MACMILLAN PUBLISHERS LTD, 1990). Recombinant antibodies are produced by cloning the encoding DNA from immunocytes such as hybridoma or antibody-producing sensitized lymphocytes, incorporating this into a suitable vector, and introducing this vector into a host to produce the antibody. The
 20 present invention encompasses such recombinant antibodies as well.

The antibody of the present invention may be an antibody fragment or a modified-antibody as long as it binds to the protein of the invention. For example, Fab, F(ab')₂, Fv, or single chain Fv in which
 25 the H chain Fv and the L chain Fv are suitably linked by a linker (scFv, Huston, J.S. et al., Proc. Natl. Acad. Sci. U.S.A., 1988, 85, 5879-5883) can be given as antibody fragments. Specifically, antibody fragments are produced by treating an antibody with an enzyme, for example, papain, pepsin, etc. or by constructing a gene encoding
 30 an antibody fragment, introducing this into an expression vector, and expressing this vector on suitable host cells (for example, Co, M.S. et al., J. Immunol., 1994, 152, 2968-2976; Better, M. and Horwitz, A.H., Methods Enzymol., 1989, 178, 476-496; Pluckthun, A. and Skerra, A., Methods Enzymol., 1989, 178, 497-515; Lamoyi, E., Methods Enzymol.,
 35 1986, 121, 652-663; Rousseaux, J. et al., Methods Enzymol., 1986, 121, 663-669; Bird, R.E. and Walker, B.W., Trends Biotechnol., 1991, 9, 132-137).

As a modified antibody, an antibody bound to various molecules

such as polyethylene glycol (PEG) can be used. The present antibody encompasses such modified antibodies as well. To obtain such a modified antibody, chemical modifications are done to the obtained antibody. These methods are already established in the field.

5 The antibody of the invention may be obtained as a chimeric antibody comprising non-human antibody-derived variable region and a human antibody-derived constant region, or as a humanized antibody comprising non-human antibody-derived complementarity determining region (CDR), and human antibody-derived framework region (FR) and
10 a constant region.

Antibodies thus obtained can be purified till uniform. The separation and purification methods for separating and purifying the antibody used in the present invention may be any method usually used for proteins, and is not in the least limited. Antibody concentration
15 of the above mentioned antibody can be assayed by measuring the absorbance, or by the enzyme-linked immunosorbent assay (ELISA), etc.

Also, as methods that assay the antigen-binding activity of the antibody of the invention, ELISA, enzyme immunoassay (EIA), radio immunoassay (RIA), or fluorescent antibody method can be given. For
20 example, when using ELISA, the protein of the invention is added to a plate coated with the antibody of the invention, and next, the objective antibody sample, for example, culture supernatants of antibody-producing cells, or purified antibodies are added. Then, secondary antibody recognizing the antibody, which is labeled by
25 alkaline phosphatase and such enzymes, is added, the plate is incubated and washed, and absorbance is measured to evaluate the antigen-binding activity after adding an enzyme substrate such as p-nitrophenyl phosphate. As the protein, a protein fragment, for example, a fragment comprising a C terminus, or a fragment comprising
30 an N terminus may be used. To evaluate the activity of the antibody of the invention, BIAcore (Pharmacia) may be used.

By using these methods, the antibody of the invention and a sample presumed to contain the protein of the invention are contacted, and the protein of the invention is detected or assayed by detecting or
35 assaying the immune complex of the above-mentioned antibody and protein.

A method of detecting or assaying the protein of the invention is useful in various experiments using proteins as it can specifically

detect or assay the proteins.

The present invention also encompasses a DNA specifically hybridizing to a DNA comprising a nucleotide sequence of any one of SEQ ID NOS: 2, 4, 6, 8, 20, and 22 to 27 or its complementary DNA, and comprising at least 15 nucleotides. Namely, a probe that can selectively hybridize to the DNA encoding the protein of the invention, or a DNA complementary to the above DNA, a nucleotide or nucleotide derivative, for example, antisense oligonucleotide, ribozyme, and such are included.

The present invention also encompasses an antisense oligonucleotide that hybridizes to any portion of any one of the nucleotide sequences shown in, for example, SEQ ID NOS: 2, 4, 6, 8, 20, and 22 to 27. This antisense oligonucleotide is preferably one against at least 15 continuous nucleotides in any one of the nucleotide sequences of SEQ ID NOS: 2, 4, 6, 8, 20, and 22 to 27. More preferable is the above-mentioned antisense oligonucleotide against the above-mentioned at least 15 continuous nucleotides containing a translation start codon.

Derivatives or modified products of antisense oligonucleotides can be used as antisense oligonucleotides. As such modified products, for example, lower alkyl phosphonate modifications such as methyl-phosphonate-type or ethyl-phosphonate-type, phosphorothioate or phosphoroamidate-modified products, etc. may be used.

The term "antisense oligonucleotide(s)" as used herein means, not only those in which the nucleotides corresponding to those constituting a specified region of a DNA or mRNA are entirely complementary, but also those having a mismatch of one or more nucleotides, as long as the DNA or mRNA and the oligonucleotide can selectively and stably hybridize with the nucleotide sequence of SEQ ID NO: 1.

"Selectively and stably hybridize" means that significant cross hybridization with DNA encoding other proteins does not occur under usual hybridization conditions, preferably under stringent hybridization conditions. Such DNAs are indicated as those having, in the "at least 15 continuous nucleotide" sequence region, a homology of at least 70% or higher, preferably 80% or higher, more preferably 90% or higher, even more preferably 95% or higher nucleotide sequence

homology. The algorithm stated herein can be used to determine homology. Such DNA is useful as a probe for detecting or isolating DNA encoding the protein of the invention, or as a primer for amplification as described in Examples below.

5 The antisense oligonucleotide derivative of the present invention acts upon cells producing the protein of the invention by binding to the DNA or mRNA encoding the protein to inhibit its transcription or translation, and to promote the degradation of mRNA, and has an effect of suppressing the function of the protein of the
10 invention by suppressing the expression of the protein.

 The antisense oligonucleotide derivative of the present invention can be made into an external preparation such as a liniment and a poultice by mixing with a suitable base material, which is inactive against the derivatives.

15 Also, as needed, the derivatives can be formulated into tablets, powders, granules, capsules, liposome capsules, injections, solutions, nose-drops, and freeze-dried agents by adding excipients, isotonic agents, solubilizers, stabilizers, preservatives, pain-killers, etc. These can be prepared using the usual methods.

20 The antisense oligonucleotide derivative is given to the patient by directly applying onto the ailing site, by injecting into a blood vessel, etc. so that it will reach the ailing site. An antisense-mounting material can also be used to increase durability and membrane-permeability. Examples are, liposome, poly-L lysine,
25 lipid, cholesterol, lipofectin, or derivatives of these.

 The dosage of the antisense oligonucleotide derivative of the present invention can be adjusted suitably according to the patient's condition and used in desired amounts. For example, a dose range of 0.1 to 100 mg/kg, preferably 0.1 to 50 mg/kg can be administered.

30 The antisense oligonucleotide derivative of the present invention is useful in inhibiting the expression of the protein of the invention, and therefore is useful in suppressing the biological activity of the protein of the invention. Also, expression-inhibitors comprising the antisense oligonucleotide derivative of
35 the present invention are useful because of their capability to suppress the biological activity of the protein of the invention.

Brief Description of the Drawings

Figure 1 is a schematic diagram showing the results of BlastX search where the query was 180 nucleotides of 40861-41040 including 40952-40966, the only probe sequence within the AC002303. "#": For only NR8 the number was indicated by the nucleotide number. The underline of the NR8 sequence shows the portion corresponding to the exon. Other underlined sequences show identical amino acids.

Figure 2 is a schematic diagram showing the results of BlastX scanning of 180 nucleotides in both the 5' and 3' directions, where the search centered on the 180 nucleotides of 40861-41040 containing 40952-40966, the only probe sequence within the AC002303.

Figure 3 shows the electrophoresis results of the amplification done by the RT-PCR method for the combinations of SN1/AS1, SN1/AS2, SN2/AS1, and SN2/AS2 primers using human fetal liver and skeletal muscle cDNA as templates.

Figure 4 shows the electrophoretic results of the 5'-RACE method and 3'-RACE method using human fetal liver cDNA as the template.

Figure 5 shows the nucleotide sequence and the amino acid sequence of NR8 α cDNA. The arrows show the positions of primers used for RT-PCR. They are, SN1 (798-827), SN2 (894-923), AS2 (1055-1026), and AS1 (1127-1098) from the 5' side, in their order. For two bases at the 5' end of AS1, AC, which is derived from the genomic sequence, was used in place of CT.

Figure 6 is the continuation of Fig. 5 showing the nucleotide sequence and the amino acid sequence of NR8 α cDNA.

Figure 7 shows the nucleotide sequence and the amino acid sequence of NR8 β cDNA. Two possible open reading frames (ORF) are shown.

Figure 8 is the continuation of Fig. 7 showing the nucleotide sequence and the amino acid sequence of NR8 β cDNA.

Figure 9 shows the nucleotide sequence and the amino acid sequence of NR8 γ cDNA. The 177 amino acids inserted by selective splicing are underlined.

Figure 10 is the continuation of Fig. 9 showing the nucleotide sequence and the amino acid sequence of NR8 γ cDNA. The 177 amino acids inserted by selective splicing are underlined.

Figure 11 is the continuation of Fig. 10 showing the nucleotide sequence and the amino acid sequence of NR8 γ cDNA.

Figure 12 shows the results of Northern blot analysis of NR8 expression in each organ.

Figure 13 is a schematic diagram showing the structure of the NR8 gene. Other repetitives include, (CA)_n, (CAGA)_n, (TGGA)_n, (CATA)_n, (TA)_n, (GA)_n, (GGAA)_n, (CATG)_n, (GAAA)_n, MSTA, AT-rich, MLT1A1, LINE2, FLAM_C, MER63A, and MSTB.

5 Figure 14 is a schematic diagram showing the structure of expressible proteins constructed in the expression vector.

Figure 15 shows the results of cross PCR, in which the human NR8 primer set was used against a mouse cDNA library. As the size marker, 100 bp DNA Ladder (NEB#323-1L) was used.

10 Figure 16 shows a comparison between amino acid sequences of human and mouse NR8 β . The amino acid sequences where the two coincide are shadowed. Also, cysteine residues conserved in other hemopoietin receptors are displayed in boldface type within the sequence.

15 Figure 17 shows a comparison between amino acid sequences of human and mouse NR8 γ . The amino acid sequences where the two coincide are shadowed. Also, cysteine residues conserved in other hemopoietin receptors and the WSXWS-Box are displayed in boldface type within the sequence.

20 Figure 18 shows the results of NR8 gene expression analysis in each mouse organ using the RT-PCR method. The size marker, 100 bp DNA Ladder (NEB#323-1L), is shown on the either sides of the lane. A 320 bp target gene has been detected in all organs.

25 Figure 19 shows the results of NR8 gene expression analysis in each mouse organ using the Northern blotting method (left). An approximately 4.2 kb transcript was intensely detected in the testis only. Mouse β -actin was detected in the same blot as a positive control (right).

Best Mode for Carrying Out the Invention

30 The present invention shall be described in detail below with reference to examples, but is not be construed as being limited thereto.

Example 1: Two step Blast Search

35 Probe sequences (256 types) comprising the tggag(t/c)nnntggag(t/c) (where n is an arbitrary nucleotide) as the oligonucleotide encoding the Trp-Ser-Xaa-Trp-Ser motif were designed. These sequences enable the detection of almost all known hemopoietin

receptors, except for the EPO receptor, TPO receptor, and the mouse IL6 receptor. Using each sequence as the query, the GenBank nr database was searched using the BlastN (Advanced BlastN 2.0.4) program. Default values (Descriptions=100, Alignments=100) were used as
5 parameters for the search, except for making the expectation value 100.

Since approximately 500 clones that completely matched the probe sequences were obtained as a result of the primary search, among these, a 180-residue nucleotide sequence of human genome-derived clones
10 (cosmid, BAC, and PAC) containing the probe sequence in approximately the center was excised. Next, using this 180-residue nucleotide sequence as the query, the nr database was searched again using the BlastX (Advanced BlastX 2.0.4) program to search the homology of the amino acid sequence around the probe sequence with known hemopoietin
15 receptors.

Default values were used as parameters for the search, except for making the expectation value 100. However, when extremely large number of hits were obtained (caused by the Alu sub family that is a high repetitive sequence), it was often difficult to observe hits
20 for known hemopoietic receptors. Therefore, to maximize the sensitivity in such cases, a value of "Expect=1000, Descriptions=500, Alignments=500" was used.

As a result of the secondary search by BlastX, 28 clones hit one or more known hemopoietin receptors (Table 1 to Table 8).
25

Table 1

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TGGAGTAATTGGAGC	Asn	AL009181	30692 tggagtaattggagc 30678	1p34.1-1p35	mLL11R(opposite), FOBR
TGGAGCTGATGGAGC	***	Z97987	140006 tggagctgatggagc 139992	1p36.2-36.3	line1, Leu Zip p40,
TGGAGCAGCTGGAGC	Ser	AF023268	39931 tggagcagctggagc 39917	1q21	metaxin
TGGAGCTGCTGGAGC	Cys	AL009051	78023 tggagctgctggagc 78037	1q23-24	HP-10, semaphorin F,G
TGGAGCACGTGGAGT	Thr	Z97200	112905 tggagcacgtggagt 112891	1q24	AFP enhancer BP, RAR
TGGAGTGCCTGGAGC	Ala	U95626	101031 tggagtgcctggagc 101017	3	CFTC, TcR
TGGAGTAGATGGAGT	Arg	Z84495	2547 tggagtagatggagt 2538	3p21.3	trithorax
TGGAGCTGATGGAGT	***	Z74023	5255 tggagctgatggagt 5241	3p21.3	E2ABP, fibronectin, nidgen
TGGAGTTTCTGGAGT	Phe	Z68275	7291 tggagtttctggagt 7277	4p16.3	mena, NMDAR
TGGAGTGCCTGGAGT	Ala	Z54072	21277 tggagtgcctggagt 21291	4p16.3	crk, AchR, HER3
TGGAGCTGCTGGAGC	Cys	Z69837	30266 tggagctgctggagc 30252	4p16.3	KIT, FLT3, PDGFRa
TGGAGTTACTGGAGT	Tyr	AC003951	27290 tggagtactggagt 27304	5	collagen
TGGAGCCTGTGGAGT	Leu	AC004502	48334 tggagcctgtggagt 48320	5	ADAMTS-1, properdin, etc
TGGAGTTGATGGAGC	***	L81613	2418 tggagtgtatggagc 2404	5	APC, bat2, p53
TGGAGTGTATGGAGT	Val	AC002122	43679 tggagtgtatggagt 43665	5p15.2	Met tRNA synthase
TGGAGTCCATGGAGT	Pro	AC002380	34646 tggagtccatggagt 34632	5p15.2	N-WASP, enigma
TGGAGCAACTGGAGC	Asn	AC002479	80443 tggagcaactggagc 80457	5p15.2	NEU, glycoprotein C
TGGAGCTGCTGGAGT	Cys	AC004592	125445 tggagctgctggagt 125431	5q31	CD22-B
TGGAGTAGCTGGAGT	Ser	AC002393	3721 tggagtagctggagt 3735	6	glycoprotein
TGGAGTTGCTGGAGT	Cys	AC002326	114578 tggagttgctggagt 114564	6	G3P REGULON
TGGAGTGCATGGAGT	Ala	Z84490	20244 tggagtgcattggagt 20230	6	Alu, adrenergic receptor

Table 2

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TTGGAGTTTCTGGAGC	Phe	AC002112	68699 tggagtttctggagc 68685	6	IgHv, MYD116
TGGAGCGGCTGGAGC	Gly	U89336	35829 tggagcggctggagc 35815	6p21	myosin HC, cep250,
TGGAGCGTCTGGAGC	Val	U53588	3558 tggagcgcctggagc 3672	6p21.3	ring finger, BRCA1
TGGAGTGCATGGAGT	Ala	Z98744	38358 tggagtcacatggagt 38344	6p21.3-22.3	Alu, AD7c-NTP
TGGAGTTGCTGGAGT	Cys	AL009031	104325 tggagtgtctggagt 104311	6p22.3-24.1	ACC synthase
TGGAGTGTCTGGAGT	Val	AL008729	21325 tggagtgtctggagt 21339	6p24	E1A, DUB-2
TGGAGTTGTTGGAGT	Cys	Z98755	69825 tggagtgtctggagt 69811	6q16.1-21	dynein
TGGAGCTTCTGGAGC	Phe	Z98172	35554 tggagctctggagc 35540	6q21	HGXPRT
TGGAGCAGGTGGAGC	Arg	Z97989	79116 tggagcaggtggagc 79102	6q21-22	syn fyn, slk, yes, src
TGGAGCTAATGGAGT	***	Z95326	16562 tggagcctatggagc 16576	6q22.1-6q22.33	tyrosinase
TGGAGCTCTTGGAGC	Ser	Z98049	25800 tggagctctggagc 25786	6q26-q27	collagen, AT3, C1Qb
TGGAGCTCCTGGAGT	Ser	AC003090	22068 tggagctctggagt 22082	7p15	ICE
TGGAGTATATGGAGC	Ile	AC004744	22740 tggagtatatggagc 22754	7p15-p21	TSH-R, RNABP
TGGAGTAGCTGGAGC	Ser	AC004485	86356 tggagttagctggagc 86370	7p15-p21	Hox 2.4, mLL1Ra(stop*)
TGGAGTCTTTGGAGT	Leu	AC004141	3130 tggagctttggagt 3144	7p21-p22	polyprotein
TGGAGCAGATGGAGC	Arg	AC004548	62876 tggagcagatggagc 62862	7q11.23-q21.1	NCAM
TGGAGCAACTGGAGT	Asn	AC002456	69500 tggagcaactggagt 69514	7q21	glycoprotein A
TGGAGTAACTGGAGT	Asn	AC000064	9170 tggagtaactggagt 9184	7q21-22	GA3PD
TGGAGTTATTGGAGT	Tyr	AC003085	87341 tggagtatttggagt 87355	7q21-22	Nmyc, FGFR
TGGAGTTGTTGGAGT	Cys	AC000119	65235 tggagtgttggagt 65221	7q21-7q22	FVIII, TopoIII
TGGAGTTGTTGGAGT	Cys	AC002458	44435 tggagtgttggagt 44421	7q21-q22	telomerase, NFAT
TGGAGTACATGGAGC	Thr	AC000059	9977 tggagtacatggagc 9963	7q21-7q22	Alu, Notch4

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TGGAGTCAGTGGAGC	Glu	AC000119	103655 tggagcattggagc 103659	7q21.7q22	reverse transcriptase
TTGGAGTATTGGAGT	Ile	AC002384	52216 tggagtatttggagt 52202	7q22	pol, GHR(another frame)
TGGAGCAGCTGGAGT	Ser	AC004522	55291 tggagcagctggagt 55277	7q22-q31.1	hemoglobin beta
TGGAGTGTTTGGAGT	Val	AC002466	43273 tggagtgttggagt 43287	7q31	ryanodine receptor, mTPO
TGGAGTGGCTGGAGC	Gly	AC002543	112948 tggagtggctggagc 112962	7q31.2	EGF, P-selectin
TGGAGCTGATGGAGC	***	AC000061	79564 tggagctgatggagc 79550	7q31.2	laminin B1, tubulin
TGGAGTTTGTGGAGT	Phe	AC000125	13750 tggagtttggagt 13736	7q31.3	p150
TGGAGTTGTGGAGT	Cys	AC002498	20166 tggagtgttggagt 20152	7q31.3	IL3Rb(homnosaite)
TGGAGCGGGTGGAGC	Gly	U66059	158491 tggagcgggtggagc 158477	7q35(TcRb)	properdin
TGGAGCATTTGGAGC	Ile	AC003109	4761 tggagcatttggagc 4775	7q36	CD2, HOX-2.6
TGGAGTTATTGGAGT	Tyr	AF027390	174448 tggagtatttggagt 174434	7q tel	IkB, V2R
TGGAGCATATGGAGT	Ile	AC002052	28882 tggagcatatggagt 28896	9p22	myosin VIIA, OSMR
TGGAGCAACTGGAGT	Asn	AC001643	27345 tggagcaactggagt 27331	9q34	hox1.4, gastrinR
TGGAGCGGATGGAGC	Gly	AC000396	16394 tggagcggatggagc 16380	9q34	vWf, laminin a3
TGGAGTGAGTGGAGT	Glu	U73649	16850 tggagtggatggagt 16836	11	zinc finger
TGGAGTGGGTGGAGT	Glu	U73649	16850 tggagtggatggagt 16845	11	zinc finger
TGGAGTGCCCTGGAGT	Ala	U73629	31027 tggagtgcctggagt 31041	11	Alu, gp2b, BCGF-12
TGGAGTCCCTGGAGT	Pro	U73629	3673 tggagtccctggagt 36745	11	E2FRN, hemagglutinin
TGGAGTCCCTGGAGC	Pro	U73643	14550 tggagtccctggagc 14564	11	reverse transcriptase
TGGAGCAACTGGAGC	Asn	AK015116	65621 tggagcaactggagc 65635	11p16.5	Nasopressin R, OSMR
TGGAGTGCATGGAGT	Ala	AC002350	23543 tggagtgcattggagt 23529	12q24	Alu, IFNaR

Table 4

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TGGAGTGCATGGAGT	Ala	AC004217	88822 tggagtcactggagt 88808	12q24.1	Alu, HPK
TTGGAGTTACTGGAGC	Tyr	AC002978	65893 tggagttactggagc 65907	12q24	clathrin LC, EPOR(nonWS)
TGGAGTTGTTGGAGT	Cys	AC000403	91715 tggagttgtggagt 91729	13	VHL, inhibin B
TGGAGCCGTTGGAGC	Gly	X97051	73621 tggagcgttggagc 73607	14q32.33 (IgD)	polycystic kidney
TGGAGTAGGTGGAGC	Arg	AC003024	16596 tggagtagtggagc 16582	15q26	pksF
TGGAGTTTCTGGAGC	Phe	AC002492	93356 tggagtttctggagc 93370	16	pol, PRER
TGGAGTTCATGGAGT	Ser	U91318	102406 tggagttcatggagt 102392	16	ICAM1, MBP1
TGGAGTGATGGAGT	Val	AC002289	10631 tggagtgatggagt 10645	16	Alu
TGGAGCCACTGGAGT	His	U91318	152212 tggagccactggagt 152238	16	laminin alpha5
TGGAGTTAATGGAGT	***	AC002519	81768 tggagttaatggagt 81754	16	Rho, Notch
TGGAGCTGCTGGAGT	Cys	U91326	84127 tggagctgctggagt 84113	16p11.2	NIPI-like, IL2Rr(nonWS)
TGGAGTCAATGGAGT	Glu	AC002303	10952 tggagtcactggagt 10976	16p12	TPOR, ORR, and many
TGGAGCACTGGAGC	Thr	AC002551	82245 tggagcactggagc 82259	16p12.1	envelope, androgen R
TGGAGTCCCTGGAGC	Pro	AC002299	162 tggagtcctggagc 148	16p12-p13.1	CYCLIN H, FN
TGGAGCTATTGGAGC	Tyr	AC002299	84540 tggagctattggagc 84526	16p12-p13.1	Alu, RNA adase
TGGAGTCACTGGAGT	His	U95737	16180 tggagtcactggagt 16144	16p13.1	TcRa, HLAA
			16374 tggagtcactggagt 16388		Notch, Pro-rich
			16599 tggagtcactggagt 16613		phosphatase, ORFB
TGGAGTCCTTGGAGC	Pro	U91318	112272 tggagtccttggagc 112286	16p13.1	CD80, collagen, MAP1a
TGGAGCACTTGGAGC	Thr	AC004509	26031 tggagcacttggagc 26045	16p13.3	TcRb
TGGAGCCGTTGGAGC	Arg	AC004496	28217 tggagccgttggagc 28231	16p13.3	mucin, ET1, IL12R(nonWS)

Table 5

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TGGAGCGCTGGAGC	Arg	AC004232	34550 tggagcgcctggagc 34564	16p13.3	IgLk, AGPR
TTGGAGCTACTTGGAGC	Thr	AJ003147	151180 tggagctacttggagc 151166	16p13.3	RanBP2
TGGAGCGGTGGAGC	Val	X71874	11520 tggagcgttggagc 11534	16q22.1	collagen a5IV
TGGAGCAAAATGGAGT	Lys	AC003663	114346 tggagcaaatggagt 114360	17	beta-D-glucosidase
TGGAGTCTCTGGAGC	Leu	AC003957	52898 tggagtctctggagc 52884	17	TIE-1, SEX, Rho,
TGGAGCAGATGGAGC	Arg	AC003971	76277 tggagcagatggagc 76263	18	LIMK-1, TcR
TGGAGTGCATGGAGT	Ala	AD000812	30891 tggagtgcattggagt 30905	19	Alu
TGGAGTGCATGGAGT	Ala	AC002126	85832 tggagtgcattggagt 85846	19	Alu, AD7cNTP
TGGAGCTGCTGGAGT	Cys	AC004660	10008 tggagctgctggagt 10022	19	Reps1
TGGAGCCCTGGAGT	Pro	AC004490	14389 tggagccctggagt 14403	19	mucin, ataxin-2, N-WASP
TGGAGCTGAGTGGAGC	Gln	AC003112	18315 tggagctgagtgagc 18301	19p12(NIR)	TPOR, PRLR, OBR, etc
TGGAGCAGATGGAGC	Arg	AC004004	39010 tggagcagatggagc 38996	19p12	PRLR, IL12R, GM-
		presumably a pseudogene.....		CSERb, IL11R(+stop codon)
TGGAGCACCTGGAGT	Thr	AD000685	39177 tggagcacatggagc 39163		IL3Ra(weak, 22, nonWS)
TGGAGCTGATGGAGC	***	AC002115	21015 tggagcacctggagt 21001	19p13.1	GM-CSFRb(nonWS+stop)
TGGAGCCAGTGGAGC	Gln	M63796	37164 tggagctgagtgagc 37178	19q13.1	Mpc2, Pro rich protein
TGGAGTTACTGGAGT	Tyr	AC004505	7622 tggagccagtgagc 7636	19q13.3	NFCP, titin, Jagged 2
TGGAGTTGATGGAGC	***	Z93016	31711 tggagtacttggagt 31725	20	Gap junction
TGGAGTCAATGGAGT	Gln	[35567]	31093 tggagttagtgagc 31079	20q12-13.2	smaphorin F, GHS-R, JAK2
TGGAGTGCCTGGAGT	Ala	AF039907	579 tggagtcaatggagt 565	21(MX1)	GLI, [131], IL7R(nonWS)
TGGAGTGTCTGGAGT	Val	AG000937	29892 tggagtgcctggagt 29906	21	IgV, Cyt.Oxidase
			105 tggagtgtctggagt 91	21q	peroxidase

Table 6

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TGGAGTAAATGGAGT	Lys	AP000034	28803 tggagtaaatggagt 28789	21q11.1	Na/Ca exchanger
TTGGAGTAGGTGGAGT	Arg	AP000039	24900 tggagtaggtggagt 24914	21q11.1	RNA polymerase
TGGAGTGAGTGGAGT	Glu	AP000035	21721 tggagtgtggagt 21707	21q11.1	semaphorin F
TGGAGTGCTGGAGT	Val	AG000038	26164 tggagtgtctggagt 26150	21q11.1	Glycoprotein
TGGAGTGCTGGAGT	Val	AP000038	26164 tggagtgtctggagt 26150	21q11.1	PKC
TGGAGTGCTGGAGT	Ala	AP000045	7204 tggagtgcctggagt 7218	21q11.1	IgV,
TGGAGCATTTGGAGC	Ile	AP000052	93726 tggagcatttggagc 93740	21q11.1	Ig H, TCF-3, CERP
TGGAGCCTCTGGAGC	Leu	AP000037	17581 tggagcctctggagc 17567	21q11.1	Alu, BCGF
TGGAGTGGGTGGAGT	Gly	AP000015	48480 tggagtgggtggagt 48494	21q22.2	TPO
TGGAGTGAGTGGAGT	Glu	Z97055	151632 tggagtgtgtggagt 151618	22	semaphorin H, CD44
TGGAGCTGGTGGAGT	Trp	Z83856	8503 tggagctgttggagt 8489	22	ERF
TGGAGTGGGTGGAGT	Gly	Z95113	69325 tggagtgggtggagt 69311	22q11.2-qter	factor H
TGGAGTGCTGGAGT	Ala	Z93784	36348 tggagtgcctggagt 36362	22q11.2-qter	Alu, NF2
TGGAGCCTCTGGAGT	Leu	AC002308	130741 tggagcctctggagt 130727	22q11.2	collagen a1, Na channel
TGGAGTCCCTGGAGC	Pro	AC000086	40705 tggagtccctggagc 40691	22q11.2	ADH, collagen
TGGAGCATCTGGAGC	Ile	L77569	21088 tggagcatctggagc 21074	22q11.2	Georgelathrin heavy chain 2
TGGAGCAGCTGGAGC	Ser	AC000072	9817 tggagcagctggagc 9803	22q11.2	clathrin heavy chain 2
TGGAGCAACTGGAGC	Asn	Z95116	64481 tggagcaactggagc 64495	22q11.2	IgHv, PC binding
TGGAGCTAGTGGAGC	***	AC003071	114780 tggagctagtggagc 114794	22q12.1	p150, IL4RWSNWSF*)
TGGAGCCCTGGAGC	Pro	Z80902	2675 tggagcccttggagc 2661	22q12.1-qter	FGFRb
TGGAGCTCTGGAGT	Ser	Z79999	40825 tggagctcttggagt 40839	22q12-qter	collagen a1, collagen a1,

Table 7

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TGGAGCCATTGGAGT	His	Z81308	12575 tggagccattggagt 12561	22q12-qter	MYF-5, p53, INK4a
TGGAGCGCAATGGAGT	Glu	AL008637	85322 tggagcgcaatggagt 85316	22q12.3-13.2	GM-CSFR β , IL3R, EPOR, etc
TTGGAGTGGAGTGGAGT	Glu	U62317	77740 tggagtggagtggagt 77726	22q13	latrophilin-related
TGGAGTGCATGGAGT	Ala	Z93013	31082 tggagtgcattggagt 31068	22q13	Alu, <u>G-CSFR</u> , AD7c-NTP
TGGAGTTGTTGGAGT	Cys	AC002422	19151 tggagtgtggagt 19137	X	cGMP PDase
TGGAGTGCTCGGAGT	Val	Z73418	31830 tggagtgtcaggagt 31816	X	WNT-8D, Mi-2
TGGAGTCTTTGGAGT	Leu	Z83843	114972 tggagtctttggagt 114958	X	reverse transcriptase
TGGAGTCTCTGGAGT	Leu	Z99706	7749 tggagtctcaggagt 7735	X*	Selenoprotein
TGGAGCAACTGGAGT	Asn	AC002420	70704 tggagcaactggagt 70690	X	homeoprotein, OBR(stop)
TGGAGCATGTGGAGT	Met	Z77249	5702 tggagcatgtggagt 5688	X	TcR β , <u>PLINR1</u>
TGGAGTTCCTGGAGC	Ser	Z83131	4904 tggagtctcaggagt 4890	X	VPS41 homolog
TGGAGTGGCTGGAGC	Gly	AC004388	239975 tggagtggctggagc 239989	X	GAP, mJER(stop)
TGGAGTGGCTGGAGC	Gly	AC004478	73509 tggagtggctggagc 73485	X	RNAse, mJER(stop)
TGGAGTCTATGGAGC	Leu	Z70050	9934 tggagtctatggagc 9948	X	complement C8, C7
TGGAGTCTATGGAGC	Leu	Z73986	40768 tggagtctatggagc 40750	X	complement C8, C7
TGGAGCTGTTGGAGC	Cys	L44140	112657 tggagctgttggagc 112671	X	rab GDI alpha, BDGF
TGGAGCTCATGGAGC	Ser	AC004383	144906 tggagctcatggagc 144892	X	RTase, transposon
TGGAGTAAATGGAGC	Lys	Z69732	31681 tggagtaaatggagc 31695	Xp11	OT-R, acrosin
TGGAGTTCGTGGAGC	Ser	Z92545	88703 tggagtctcaggagc 88717	Xp11	PMK1
TGGAGCTTCTGGAGC	Phe	AL008709	46089 tggagcttctggagc 46075	Xp11.23-Xp11.4	rMHC class 1a, HLA-C
TGGAGTTTCTGGAGT	Phe	U96409	116332 tggagtttctggagt 116346	Xp22	myosin H
TGGAGTTGCTGGAGT	Cys	AC003106	89544 tggagtgtcaggagt 89530	Xp22	<u>IL9R</u>

Table 8

Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TTGGAGTCACTGGAGT	His	AL021706	11982 tggagtcactggagt 11968	Xq21.1-21.33	dopamine receptor
TGGAGCTGGTGGAGT	Trp	AC000113	119188 tggagctggaggagt 119202	Xq23	DNA repair protein, MHC
TGGAGCAAGTGGAGT	Lys	AF007262	98212 tggagcaagtggagt 98226	Xq28	RNA polymerase
TGGAGCTGCTGGAGT	Cys	U82671	35792 tggagctgctggagt 35806	Xq28	XTCF-3c
TGGAGTCAGTGGAGC	Gln	AF011899	144465 tggagtcagtggagc 144451	Xq28	GHRHR, Werner Synd.
TGGAGCTAATGGAGC	***	AF030876	107409 tggagctaatggagc 107395	Xq28	gp41, clk3
TGGAGCTTCTGGAGT	Phe	AC002531	106698 tggagcttctggagt 106712	Xq28	Alu, hpk
TGGAGCAGTGGAGC	Ser	AC004474	124745 tggagcagtggagc 124731	Y	EGFR, Smad6
TGGAGTTTGTGGAGT	Leu	U26425	12899 tggagtttgtggagt 12913	PLCb2	PRLR(opposite)
TGGAGCAACTGGAGT	Asn	U96726	61672 tggagcaactggagt 61658	mouse DNA	envelope mIL11R(opposite)
TGGAGTCCCTGGAGC	Pro	<u>U35323</u>	22244 tggagtcctcggagc 22230	MHC class II	CFTC, <u>IL6R</u>
TGGAGCAGATGGAGC	Arg	AC002482	14276 tggagcagatggagc 14290	RG208O03	I-309, TcR, IL9R(nonWS)
TGGAGCTCTTGGAGC	Ser	U34879	24914 tggagctcttggagc 24928	EDH17B2	Large tegument protein
TGGAGCCTTTGGAGC	Leu	Z15025	6359 tggagcctttggagc 6373	Bat2	commonR(opposit.nonWS)
GM-CSERb(opposite.stop)					bat2,mucin,

Redundant clones are shadowed. White and underlined letters indicate hits and pseudo-hits, respectively.

Four clones out of these 28 clones (AC002303, AC003112, AL008637, and AC004004) hit several known hemopoietin receptors, however, AC004004 was excluded as it has a stop codon downstream three amino acids of the Trp-Ser-Xaa-Trp-Ser motif. Among the three remaining clones, AL008637 was thought to be a known receptor, GM-CSF receptor β . AC002303 is the BAC clone CIT987-SKA-670B5 derived from the 16p12 region of human chromosome no. 16 registered by TIGR group on June 19, 1997 and comprises the full-length of 131530 base pairs (Lamerdin, J.E., et al., GenBank Report on AC003112, 1997).

As shown in Fig. 1, a BlastX search (query: 180 nucleotides of 40861-41040 including tggagtgaatggagt (40952-40966), the only probe sequence within the AC002303) revealed that numerous hemopoietin receptors starting with the TPO receptor and leptin receptor show an evident homology, however, there were no known, database-registered hemopoietin receptors that completely matched the query sequence. Also, a BlastX scanning was done under the above conditions, by excising a sequential 180-residue nucleotide sequence in both the 5' and 3' directions, centering on the 180-residue nucleotide sequence mentioned above, and when this was used as a query, two sequences having a homology to known hemopoietin receptors were found in the regions 39181-39360 and 42301-42480, and were thought to be other exons of the same gene (Fig. 2).

A Pro-rich motif PAPPF was conserved in the 39181-39360 site, and a Box 1 motif in the 42301-42480 site. The 3' side exon adjacent to the exon containing the Trp-Ser-Xaa-Trp-Ser motif has a transmembrane domain, and this domain has a low homology with other hemopoietin receptors, and was not detected by the BlastX scan. These results suggested the possibility of a novel hemopoietin receptor gene existing in the above-described BAC clone CIT987-SKA-670B5.

Example 2: Search for NR8 expressing tissues using RT-PCR

Pseudogenes have been reported to exist in several hemopoietin receptors (Kermouni, A. et al., Genomics, 1995, 29 (2) 371-382; Fukunaga, R. and Nagata, S., Eur. J. Biochem., 1994, 220, 881-891).

To verify that NR8 is not a pseudogene, and with the objective of identifying NR8 expressing tissues, transcripts of the NR8 gene were searched by RT-PCR method.

In the AC002303 sequence of the above-described BAC clone, several

exon regions widely conserved at the amino acid translation level in known cytokine receptors were surmised, and on the sequence of the surmised exon region, the following primers were synthesized. (See Fig. 5 for the location of each primer.)

- 5 NR8-SN1; 5'- CCG GCT CCC CCT TTC AAC GTG ACT GTG ACC -3' (SEQ ID NO: 9)
 NR8-SN2; 5'- GGC AAG CTT CAG TAT GAG CTG CAG TAC AGG -3' (SEQ ID NO: 10)
 NR8-AS1; 5'- ACC CTC TGA CTG GGT CTG AAA GAT GAC CGG -3' (SEQ ID NO: 11)
 10 NR8-AS2; 5'- CAT GGG CCC TGC CCG CAC CTG CAG CTC ATA -3' (SEQ ID NO: 12)

Using the Human Fetal Multiple Tissue cDNA Panel (Clontech #K1425-1) as the template, RT-PCR was attempted using combinations
 15 of the above primers. Advantage cDNA Polymerase Mix (Clontech #8417-1) was used for the PCR, which was conducted under the conditions below using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler.

Namely, the PCR conditions were, 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 3 min," 5 cycles of "94°C for 20 sec, 70°C for 3 min," 28 cycles of "94°C for 20 sec, 68°C for 3 min," 72°C for 4 min,
 20 and completed at 4°C.

From the primer locations shown in Fig. 5, amplifications of bands sized 330 bp, 258 bp, 234 bp, and 162 bp can be expected from the combinations of SN1/AS1, SN1/AS2, SN2/AS1, and SN2/AS2. When
 25 evaluated using human fetal liver, brain, and skeletal muscle cDNA as the template, clear bands having the anticipated sizes were obtained in the fetal liver only with the respective primer combinations (Fig. 3).

An amplification was not seen at all for fetal brain cDNA, and
 30 a band of about 650 bp and a broad band of 400 to 500 bp were observed for fetal skeletal muscle cDNA. However, since the band sizes for skeletal muscle cDNA remained constant even when different combinations of primers were used, it is thought that these bands were non-specific amplifications due to some reason.

35 The obtained PCR product was subcloned to pGEM-T Easy vector (Promega #A1360), and the nucleotide sequence was determined. The recombination of PCR products to the pGEM-T Easy vector was done by T4 DNA Ligase (Promega #A1360) reacted at 4°C for 12 hr. The genetic

recombinant between the PCR product and pGEM-T Easy vector was obtained by transforming *E. coli* strain DH5 α (Toyobo #DNA-903).

For the selection of the genetic recombinant, Insert Check Ready (Toyobo #PIK-101) was used. The dRhodamine Terminator Cycle
 5 Sequencing Kit (ABI/Perkin Elmer #4303141) was used for determining the nucleotide sequence, and analysis was done using the ABI PRISM 377 DNA Sequencer. As a result of determining the nucleotide sequences of all inserts of the 10 independent clones of genetic recombinants, all clones were found to comprise a single nucleotide
 10 sequence. These obtained sequences were verified to be partial nucleotide sequences of NR8.

Example 3: Full-length cDNA cloning by the 5' and 3'-RACE methods

Using the thus-obtained fetal liver-derived cDNA, 5' and 3'-RACE
 15 methods were conducted to obtain full-length cDNA (Fig. 4).

3-1) 5'-RACE method

5'-RACE PCR was performed using the above-mentioned NR8-AS1 primer for primary PCR, and NR8-AS2 primer for secondary PCR. Human Fetal Liver Marathon-Ready cDNA Library (Clontech #7403-1) was used as the
 20 template and Advantage cDNA Polymerase Mix for the PCR experiment. As a result of PCR under the following conditions using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler, two types of PCR products were obtained, which have different sizes through selective splicing.

Primary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 4 min," 5 cycles of "94°C for 20 sec, 70°C for 4 min,"
 25 28 cycles of "94°C for 20 sec, 68°C for 4 min," 72°C for 4 min, and completed at 4°C.

Secondary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 70°C for 3 min 30 sec," 28 cycles of "94°C for 20 sec, 68°C
 30 for 3 min 30 sec," 72°C for 4 min, and completed at 4°C.

Both types of PCR products obtained were subcloned to pGEM-T Easy vector as mentioned earlier, and the nucleotide sequences of all inserts were determined for the 16 independent clones of genetic transformants. As before, the dRhodamine Terminator Cycle
 35 Sequencing Kit was used for determining the nucleotide sequence, and analysis was done using the ABI PRISM 377 DNA Sequencer. As a result, the clones can be divided into two groups, one having 14 clones, and the other having 2 clones, by the length of the base pairs and the

differences in sequence (though described later, the differences lie in the products due to selective splicing, and the group of 14 independent clones comprises the sequence corresponding to exon 5 in the genomic sequence, and the remaining group of two independent clones does not have this sequence).

3-2) 3'-RACE method

3'-RACE PCR was performed using the above-mentioned NR8-SN1 primer for primary PCR, and NR8-SN2 primer for secondary PCR. Human Fetal Liver Marathon-Ready cDNA Library was used as the template similar to 5'-RACE PCR, and Advantage cDNA Polymerase Mix for the PCR experiment. As a result of conducting PCR under the conditions shown in 3-1), a single band PCR product was obtained.

The obtained PCR product was subcloned to pGEM-T Easy vector as above, and the nucleotide sequences of all inserts of the 12 independent clones of genetic recombinants were determined. As before, the dRhodamine Terminator Cycle Sequencing Kit was used for determining the nucleotide sequence, and the sequences determined were analyzed using the ABI PRISM 377 DNA Sequencer. As a result, all 12 independent clones showed a single nucleotide sequence.

As a result of analyzing the nucleotide sequence of the fragments (approximately 1.1 kb and 1.2 kb) amplified by 5'-RACE and 3'-RACE, respectively, it was conceived that the approximately 260 bp of each fragment overlap and extend to the 5' side and 3' side, and contain almost the full-length of NR8 mRNA. These were joined to make a full-length cDNA (NR8 α) (Fig. 5 and Fig. 6). The plasmid containing the NR8 α cDNA (SEQ ID NO: 2) was named pGEM-NR8 α , and *E. coli* containing the plasmid has been internationally deposited at the National Institute of BioScience and Human-Technology, Agency of Industrial Science and Technology (1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) under the accession number FERM BP-6543 since October 9, 1998 according to the Budapest Treaty.

As shown in Fig. 5 and Fig. 6, in the ORF of NR8 α cDNA, the Met starting from nucleotide no. 441 is thought to be the start codon due to the presence of an inframe stop codon 39 bp upstream, and completes with two stop codons starting from nucleotide no. 1524. It has the features of, from the N terminus in order, a typical secretion signal sequence, a domain thought to be the ligand binding

site containing a Cys residue conserved in other hemopoietic receptor members, a Pro-rich motif, Trp-Ser-Xaa-Trp-Ser motif, a transmembrane domain, a Box 1 motif thought to be involved in signal transduction, and such features of hemopoietin receptors. From the above results,
5 the NR8 gene was thought to encode a novel hemopoietin receptor.

Analysis of fragments amplified by the RACE method suggested the presence of a splice variant. As a result of nucleotide sequence analysis, this variant was revealed to be lacking approximately 150 bp including the above-described Pro-rich motif of NR8 α . Moreover,
10 as a result of comparing AC002303 sequence with NR8 α , and carrying out analogy of exons/introns (Table 9), the above-described variant was thought to be deficient of the 5th exon due to selective splicing.

Table 9

Exon	# in AC002303	# in NR8	Characteristics
1	<1	: 1-424	inframe stop codon
2	26334-26398	: 425-489	start codon, signal peptide
3	30625-30727	: 490-592	conserved Cys residue
4	33766-33965	: 593-792	conserved Cys residue, N-glycosylation site
5	39240-39394	: 793-947	Pro-rich motif (PAPPF), N-glycosylation site
6	40820-40997	: 948-1125	gtWSEWSdp motif
7	41455-41554	: 1126-1225	transmembrane domain
8	42285-42366	: 1226-1307	Box1 (IWAVPSP)
9a	44812-44909	: 1308-1405*	connects to exon 10, Box2-like sequence (PSTLEVYSCH), nontypical exon/intron boundary
9b	44812-45922<	: 1308-2465**	double stop codons, Box2-like sequence (PSTLEVYSCH, PAELVESDG), polyA
10	45441-45922<	: 1406-1934*	double stop codons, polyA
NR8 α^* : exons 1+2+3+4+5+6+7+8+9a+10			
NR8 β : exons 1+2+3+4+6+7+8+9a+10			
(two alternative reading frames for soluble-type and transmembrane(-signal)-type)			
NR8 γ^{**} : exons 1+2+3+4+5+6+7+8+9b			

This variant (NR8 β) can encode a soluble receptor in the truncated form by the joining of the 6th exon directly to the 4th exon and causing a frame shift. The boundary between the exons and the introns takes a consensus sequence in most cases, but the boundary between the 9th exon (Exon 9a) and the 9th intron is the only boundary that takes a different sequence from the consensus sequence (nag/gtgagt, etc.), being acc/acggag. The plasmid comprising NR8 β cDNA (SEQ ID NO: 4) was named pGEM-NR8 β , and *E.coli* comprising the plasmid has been internationally deposited at the National Institute of BioScience and Human-Technology, Agency of Industrial Science and Technology (1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) under the accession number FERM BP-6544 since October 9, 1998 according to the Budapest Treaty.

15 Example 4: Northern blotting

In order to analyze the distribution and mode of NR8 gene expression in each human organ and human cancer cell lines, Northern blot analysis was done using the cDNA encoding the full-length NR8 α protein prepared based on all the cDNA fragments obtained in Example 3 as a probe. The probe was prepared using Mega Prime Kit (Amersham, cat#RPN1607) by radiolabeling it with [α -³²P] dCTP (Amersham, cat#AA0005).

As Northern blots, Human Multiple Tissue Northern (MTN) Blot (Clontech #7760-1), Human MTN Blot IV (Clontech #7766-1), and Human Cancer Cell Line MTN Blot (Clontech #7757-1) were used. Express Hyb Hybridization Solution (Clontech #8015-2) was used for hybridization.

Hybridization conditions were: a prehybridization at 68°C for 30 min, followed by hybridization at 68°C for 14 hr. After washing under the following conditions, the blots were exposed to Imaging Plate (FUJI#BAS-III), and the gene expression of NR8 mRNA was detected by the Image Analyzer (FUJIX, BAS-2000 II). Washing conditions were: (1) 1x SSC/0.1% SDS, at room temperature for 5 min; (2) 1x SSC/0.1% SDS, at 50°C 30 min; and (3) 0.1x SSC/0.1% SDS, at 50°C 30 min.

Fig. 12 shows the results of Northern blot analysis of NR8 expression in each organ. A total of three different-sized mRNA, one 5kb-sized and two 3 to 4kb sized, were detected in human adult lung, spleen, thymus, skeletal muscle, pancreas, small intestines, peripheral leucocytes, and uterus. A similar examination of various cell lines including hemopoietic cell lines showed similar sized bands

in two cell lines, the promyeloid leukemic cell line HL60 and Burkett's lymphoma-derived Raji.

Example 5: Plaque screening

5 Northern blot analysis of NR8 gene expression detected at least three types of specific mRNA bands with different sizes in each human organ and in each human cancer cell line for which NR8 gene expression was seen. However, the inventors had succeeded in isolating only two types of selective splicing variants, namely NR8 α and NR8 β genes, in
10 the above-described Examples. Therefore, the inventors performed plaque screening with the objective of isolating the gene of the third selective splicing variant. Human Lymph Node (Clontech, cat#HL5000a) that showed a strong NR8 gene expression in the above-mentioned Northern analysis results, was used as the cDNA
15 library. The probe used was NR8 α cDNA fragment, which was radio-labeled by [α -³²P] dCTP (Amersham, cat#AA0005) using the Mega Prime Kit (Amersham, cat#RPN1607). Approximately 7.2×10^5 plaques of Human Lymph Node cDNA Library were blotted onto a Hybond N (+) (Amersham, cat#RPN303B) charged nylon membrane to conduct primary screening.
20 Rapid Hybridization Buffer (Amersham, cat#RPN1636) was used for the hybridization. Hybridization conditions were: a prehybridization at 65°C for 1 hr, followed by hybridization at 65°C for 14 hr. After washing under the conditions, (1) 1x SSC/0.1% SDS, at room temperature for 15 min; (2) 1x SSC/0.1% SDS, at 58°C 30 min; and (3) 0.1x SSC/0.1%
25 SDS, at 58°C 30 min, the membrane was exposed to an X-ray film (Kodak, cat#165-1512) to detect NR8 positive plaques.

As a result, positive or pseudo-positive 16 independent clones were obtained. A similar secondary screening was done for the 16 clones obtained from the primary screening to successfully isolate
30 plaques of NR8 positive 15 independent clones. The inserts of these 15 clones were amplified by PCR through a pair of primers located in both ends of the λ gt10 vector cloning site. Advantage cDNA polymerase Mix (Clontech #8417-1) was used for the PCR reaction conducted using the Perkin Elmer Gene Amp PCR System 2400
35 Thermalcycler, under the following experiment conditions. Namely, 94°C for 4 min, 5 cycles of "94°C for 20 sec, 70°C for 4 min," 30 cycles of "94°C for 20 sec, 68°C for 4 min," 72°C for 4 min, and completed at 4°C.

Similar to above, the obtained PCR products were subcloned to pGEM-T Easy vector, and the nucleotide sequence of the inserts were determined using the BigDye Terminator Cycle Sequencing SF Ready Reaction Kit (ABI/Perkin Elmer#4303150), and analyzed by the ABI PRISM 377 DNA Sequencer. As a result, among the 15 clones obtained, at least two clones showed an insertion of 177 amino acids flanking the NR8 α C terminus, and since this portion derives from the 9th intron of the NR8 gene and is removed by splicing in NR8 α , this 3rd selective splicing variant was named NR8 γ . The plasmid containing the NR8 γ cDNA (SEQ ID NO: 8) was named pGEM-NR8 γ , and *E.coli* containing the plasmid has been internationally deposited at the National Institute of BioScience and Human-Technology, Agency of Industrial Science and Technology (1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) under the accession number FERM BP-6545 since October 9, 1998 according to the Budapest Treaty.

Among the 15 clones obtained here, four clones other than the two mentioned above were further selected, and their nucleotide sequences were analyzed. As a result, among the six clones selected, two clones had the NR8 β nucleotide sequence, and all the remaining four clones had the NR8 γ nucleotide sequence. Therefore, the six clones for which the nucleotide sequence was analyzed did not contain the NR8 α sequence. The NR8 γ cDNA clones for which the nucleotide sequences were determined included those having 3'-UTR (3UTR-2) in which a poly-A tail is added to the site elongated 483 bp from the 3'-UTR of NR8 α obtained by the 3'-RACE method (3UTR-1), and those having 3'-UTR (3UTR-3) in which a poly-A tail is added to the site elongated 2397 bp from the 3'-UTR of NR8 α . On the other hand, the two clones of NR8 β for which the nucleotide sequence was decided above, both contained the nucleotide sequence of 3UTR-3. In Table 10 below, the 3' end non-translation region sequences contained in the cDNA clones thus far obtained are summarized. Also, the nucleotide sequences of 3UTR-1, 3UTR-2, and 3UTR-3 following the translation stop codon of NR8 γ cDNA sequence are shown in SEQ ID NO: 23, SEQ ID NO: 24, and SEQ ID NO: 25, respectively.

Moreover, the nucleotide sequences of 3UTR-B1 and 3UTR-B3 following the translation stop codon of NR8 β cDNA sequence are shown in SEQ ID NO: 26 and SEQ ID NO: 27, respectively.

Table 10

	NR8 cDNA clone	3'-UTR sequence
5	NR8 α	3UTR-1
	NR8 β	3UTR-B1, 3UTR-B3
	NR8 γ	3UTR-1, 3UTR-2, 3UTR-3

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The nucleotide sequences thus obtained revealed that the gene transcripts of NR8 can encode various different sizes not only due to the differences in selective splicing, but also due to the length of the 3' end non-translation region sequence. This may adequately explain the presence of various-sized transcripts detected by Northern blot analysis.

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Example 6: Ligand screening

6-1) Construction of NR8 chimeric receptor

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A screening system was constructed for searching a ligand that can specifically bind to NR8, namely, a novel hemopoietin. First, the cDNA sequence encoding the extracellular region of NR8 α (the amino acid sequence of SEQ ID NO: 1; from the 1st Met to the 228th Glu) was amplified by PCR, and this DNA fragment was bound to DNA fragments encoding the transmembrane region and the intracellular region of a known hemopoietin receptor to prepare a fusion sequence encoding a chimeric receptor. As described above, there were several candidates for the partner, the known hemopoietin receptor, and among them, the human TPO receptor (Human MPL-P) was selected. Namely, after amplifying the DNA sequence encoding the intracellular region that includes the transmembrane region of the human TPO receptor by PCR, this sequence was bound to the cDNA sequence encoding the extracellular region of NR8 α in frame, and inserted into a plasmid vector expressible in mammalian cells. The expression vector constructed was named pEF-NR8/TPO-R. A schematic diagram of the structure of the constructed NR8/TPO-R chimeric receptor is shown in Fig. 14, and the nucleotide sequence of the chimeric receptor and the expressible amino acid sequence encoded by it are shown in SEQ

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ID NOs: 13 and 14, respectively. Together with an expression vector pSV2bsr (Kaken Pharmaceutical Co., Ltd.) containing Blastcidin S resistant gene, the NR8/TPO-R chimeric receptor-expressing vector was introduced into the growth factor-dependent cell line Ba/F3, and
 5 forcedly expressed. Gene-introduced cells were selected by culturing with 8 µg/ml of Blastcidin S hydrochloride (Kaken Pharmaceutical Co., Ltd.) and IL-3. By transferring the obtained chimeric receptor-introduced cells to an IL-3-free medium, adding a material expected to contain a target ligand, and culturing, it
 10 is possible to conduct a screening that uses the fact that survival/proliferation will be possible only when a ligand that specifically binds to NR8 is present.

6-2) Preparation of NR8/IgG1-Fc soluble fusion protein

15 NR8/IgG1-Fc soluble fusion protein was prepared to be used for searching cell membrane-bound ligands, or the detection of soluble ligands through BIAcore (Pharmacia) and West-western blotting. A fusion sequence encoding the soluble fusion protein was prepared by binding a DNA fragment encoding the extracellular region of NR8α
 20 (amino acid sequence; from the 1st Met to the 228th Glu) prepared in 5-1) with the DNA fragment encoding the Fc region of human immunoglobulin IgG1 in frame. A schematic diagram of the structure of the soluble fusion protein encoding the NR8/IgG1-Fc is shown in Fig. 14, and the nucleotide sequence and the expressible amino acid
 25 sequence encoded by it in SEQ ID NOs: 15 and 16, respectively. This fusion gene fragment was inserted into a plasmid vector expressible in mammalian cells, and the constructed expression vector was named pEF-NR8/IgG1-Fc. If this pEF-NR8/IgG1-Fc is forcedly expressed in mammalian cells, and after selecting stable gene-introduced cells,
 30 the recombinant protein secreted into the culture supernatant can be purified by immunoprecipitation using anti-human IgG1-Fc antibody, or by affinity columns, etc.

35 6-3) Construction of an expression system of NR8β and purification of recombinant NR8β protein

The recombinant NR8β protein was prepared to be used for searching cell membrane-bound ligands, or the detection of soluble ligands using BIAcore (Pharmacia) or West-western-blotting. Using the amino acid

coding sequence of NR8 β cDNA, the stop codon was replaced by point mutation to a nucleotide sequence encoding an arbitrary amino acid residue, and then, was bound to the nucleotide sequence encoding the FLAG peptide in frame. This bound fragment was inserted into a plasmid vector expressible within mammalian cells, and the constructed expression vector was named pEF-BOS/NR8 β FLAG. Fig. 14 shows a schematic diagram of the structure of the insert NR8 β FLAG within the constructed expression vector. Moreover, the nucleotide sequence of NR8 β FLAG and the expressible amino acid sequence encoded by it are shown in SEQ ID NOs: 17 and 18, respectively. If this pEF-BOS/NR8 β FLAG is forcedly expressed in mammalian cells, and after selecting stable gene-introduced cells, the recombinant protein secreted into the culture supernatant can be immunoprecipitated using anti-FLAG peptide antibody, or may be purified by affinity columns, etc.

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Example 7: Isolation of mouse NR8 (mNR8) gene

7-1) The mouse homologous gene using human NR8 primers

Xenogeneic cross PCR cloning was isolated using the oligonucleotide primers, NR8-SN1 and NR8-SN2 (SEQ ID NOs: 9 and 10) at the sense side (downstream direction) and NR8-AS1 and NR8-AS2 (SEQ ID NOs: 11 and 12) at the antisense side (upstream direction), which were used for isolating full-length cDNA of human NR8. By combining the above-mentioned human NR8 primers, four types of primer sets can be constructed. Namely, using the combinations of "NR8-SN1 vs. NR8-AS1," "NR8-SN1 vs. NR8-AS2," "NR8-SN2 vs. NR8-AS1," and "NR8-SN2 vs. NR8-AS2," and a mouse brain cDNA library (Clontech #7450-1) and a mouse testis cDNA library (Clontech #7455-1) as templates, amplification of cross PCR products was expected. Advantage cDNA Polymerase Mix (Clontech #8417-1) was used for the PCR that was conducted under the conditions below using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler to amplify partial nucleotide sequence that could encode a mouse homologous gene of this receptor.

Namely, the cross PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 1 min," 5 cycles of "94°C for 20 sec, 70°C for 1 min," 28 cycles of "94°C for 20 sec, 68°C for 1 min," 72°C for 4 min, and completed at 4°C.

As a result, as shown in Fig. 15, an amplification of the cross PCR product was seen when any primer set was used. Also, a much clearer

amplification product can be obtained when mouse brain cDNA was used as the template than when mouse testis cDNA was used.

- 7-2) Determination of the partial nucleotide sequence of the mouse homologous gene corresponding to NR8

Among the amplification products obtained in 7-1), mouse brain cDNA-derived product was subcloned to pGEM-T Easy vector (Promega #A1360), and the nucleotide sequence was determined. Namely, the PCR product was recombined into pGEM-T Easy vector by using T4 DNA ligase (Promega #A1360) at 4°C for 12 hr, and the resulting product was transfected into *E.coli* strain DH5α (Toyobo #DNA-903) to obtain the genetic recombinants of the PCR product and pGEM-T Easy vector. For the selection of genetic recombinant, Insert Check Ready Blue (Toyobo #PIK-201) was used. The nucleotide sequence was determined by using the BigDye Terminator Cycle Sequencing Ready Reaction Kit (ABI/Perkin Elmer #4303154), and sequence analysis was done by the ABI PRISM 377 DNA Sequencer. As a result of determining the nucleotide sequence of all inserts of eight independent clones of genetic recombinants, nucleotide sequences derived from the same transcript were obtained, and they were verified to be partial nucleotide sequences of mNR8. The obtained partial nucleotide sequence is shown in SEQ ID NO: 28.

- 7-3) Design of oligonucleotide primers specific to the mouse NR8 gene

Based on the partial nucleotide sequence of mNR8 obtained in 7-2), oligonucleotide primers specific to the mouse NR8 were designed. As shown in the sequence given below, mNR8-SN3 was synthesized in the sense side (downstream direction), and, mNR8-AS3 was synthesized in the antisense side (upstream direction). ABI's 394 DNA/RNA Synthesizer was used for primer synthesis, which was done under 5'-end trityl residue addition conditions. After that, the complete length of the synthesized product was purified by using an OPC column (ABI #400771). These primers contributed towards the 5'-RACE method and the 3'-RACE method described later on.

mNR8-SN3; 5'- TCC AGG CGC TCA GAT TAC GAA GAC CCT GCC -3' (SEQ ID NO: 29)

mNR8-AS3; 5'- ACT CCA GGT CCC CTG GTA GGA GGA GCC AGG -3' (SEQ ID NO: 30)

7-4) Cloning of cDNA corresponding to N terminus by the 5'-RACE method

To isolate full-length cDNA of mNR8, 5'-RACE PCR was performed using the NR8-AS2 primer (SEQ ID NO: 12) for the primary PCR, and the above-mentioned mNR8-AS3 primer (SEQ ID NO: 30) for secondary

- 5 PCR. Mouse Brain Marathon-Ready cDNA Library (Clontech #7450-1) was used as the template, and Advantage cDNA Polymerase Mix for PCR experiment. As a result of conducting PCR under the following conditions using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler, PCR products of two different sizes were obtained.
- 10 Primary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 100 sec," 5 cycles of "94°C for 20 sec, 70°C for 100 sec," 28 cycles of "94°C for 20 sec, 68°C for 100 sec," 72°C for 3 min, and completed at 4°C.

- 15 Secondary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 70°C for 100 sec," 25 cycles of "94°C for 20 sec, 68°C for 100 sec," 72°C for 3 min, and completed at 4°C.

- Both types of PCR products obtained were subcloned to pGEM-T Easy vector as described above, and the nucleotide sequences were determined. Namely, the PCR products were recombined into the pGEM-T
- 20 Easy vector with T4 DNA ligase at 4°C for 12 hr, and the resulting product was transfected into *E.coli* strain DH5 α to obtain the genetic recombinant between the PCR product and pGEM-T Easy vector. Also, as mentioned earlier, Insert Check Ready Blue was used for the selection of the genetic recombinant. For the determination of the
- 25 nucleotide sequence, the BigDye Terminator Cycle Sequencing Ready Reaction Kit was used, and the nucleotide sequence was analyzed by the ABI PRISM 377 DNA Sequencer. The result of determining the nucleotide sequences of all inserts of eight independent clones of genetic recombinants suggests that they could be divided into two
- 30 groups of four clones each by the base pair length and differences in the sequence. This difference of the products was caused by selective splicing, and both of the obtained sequences were verified to contain the sequence of full-length mNR8 cDNA clone corresponding to the N terminal sequence. The cDNA clone comprising the long ORF
- 35 containing the exon encoding the Pro-rich region was named mNR8 γ , and the cDNA clone encoding the short ORF that does not have the Pro-rich region was named mNR8 β . These clones correspond to xenogeneic homologous genes of human NR8 γ and human NR8 β , respectively.

7-5) Cloning of cDNA corresponding to C terminus using the 3'-RACE method

To isolate full-length cDNA of mNR8, 3'-RACE PCR was performed using the NR8-SN1 primer (SEQ ID NO: 9) for the primary PCR, and the mNR8-SN3 primer (SEQ ID NO: 29) for secondary PCR. Mouse Brain Marathon-Ready cDNA Library was used as the template, and Advantage cDNA Polymerase Mix for PCR experiment. As a result of conducting PCR under the above-mentioned conditions using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler, a PCR product of a single size was obtained. The PCR product obtained was subcloned to pGEM-T Easy vector as before according to 7-2), and the nucleotide sequence was determined. As a result of determining the nucleotide sequences of all inserts of four independent clones of genetic recombinants, it was found to contain the sequence of full-length mNR8 cDNA corresponding to the C terminal sequence. By combining the resulting nucleotide sequence determined through this 3'-RACE PCR, and the nucleotide sequence of 5'-RACE PCR products determined in 7-4), the complete nucleotide sequences of the full-length of mNR8 γ and mNR8 β cDNA were finally determined. The determined mNR8 γ cDNA nucleotide sequence and the amino acid sequence encoded by it are shown in SEQ ID NOs: 22 and 21, respectively. The determined mNR8 β cDNA nucleotide sequence and the amino acid sequence encoded by it are shown in SEQ ID NOs: 20 and 19, respectively.

When the human and mouse NR8 amino acid sequences were compared, a high homology of 98.9% was seen for NR8 γ , and the homology was 97.2% even for NR8 β . This result strongly suggests the possibility that the same receptor gene has a vital functional responsibility that exceeds species. Fig. 16 shows a comparison between human and mouse NR8 β amino acid sequences. Fig. 17 shows a comparison between human and mouse NR8 γ amino acid sequences.

Both the full-length cDNAs of mNR8 γ and mNR8 β finally isolated were able to encode the transmembrane receptor protein comprising 538 amino acids, and the soluble receptor-like protein comprising 144 amino acids, respectively, through a selective splicing similar to human NR8. The structure below shows the characteristics of mNR8 γ . First, it is presumed that from amino acid no. 1 Met to amino acid no. 19 Gly is a typical secretion signal sequence. Here, since an inframe

stop codon exists in the minus 13 position from the 1st Met, this Met residue is presumed to be the translation start codon. Next, from the 25th Cys to the 35th Cys residue is a typical ligand binding site sequence, and the 65th and 109th Cys residues also show the repetitive Cys residue structure conserved in other hemopoietin receptors as well. Next, the Pro-rich region is conserved by the Pro residues repeating at the 120th, 122nd, and 123rd positions. From the 214th Trp to 218th Ser residue is a typical WSXWS-Box (WS motif). Following these structural characteristics in the extracellular region, a typical transmembrane domain is seen in the 23 amino acids from the 233rd Gly to the 255th Leu. In the intracellular region that follows, the 271st and 273rd Pro residues are Box-1 consensus sequence (PXP motif) conserved in other hemopoietin receptor members, and these are thought to be deeply involved in signal transduction. Thus, mNR8 γ adequately satisfies the characteristics of hemopoietin receptor members.

On the other hand, for mNR8 β , among the structural characteristics for the above-mentioned extracellular region, the exon sequence encoding the Pro-rich region has been skipped by selective splicing, and directly joins the next exon encoding the WS motif. However, the WSXWS-Box sequence has been excluded from the reading frame by frame shift, and after coding up to 144th Leu, the translation frame completed the next stop codon. Thus, a soluble hemopoietin receptor-like protein that does not have a transmembrane domain is encoded.

25 Example 8: Expression analysis of mouse NR8 gene

8-1) Analysis of mouse NR8 gene expression by the RT-PCR method

To analyze the distribution and mode of NR8 gene expression in each mouse organ, the mRNA was detected by RT-PCR analysis. As primers for this RT-PCR analysis, NR8-SN1 primer (SEQ ID NO: 9) was used as the sense side (downstream direction) primer, and NR8-AS1 primer was used as the antisense side (upstream direction) primer. Mouse Multiple Tissue cDNA Panel (Clontech #K1423-1) was used as the template. Advantage cDNA Polymerase Mix (Clontech #8417-1) and the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler were used for PCR. The target genes were amplified by the PCR reaction under the cycle condition given below.

PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 1 min," 5 cycles of "94°C for 20 sec, 70°C for 1 min," 24 cycles

of "94°C for 20 sec, 68°C for 1 min," 72°C for 3 min, and completed at 4°C.

The results of RT-PCR are shown in Fig. 18. The NR8 gene was strongly detected in the testis and day 17 embryo, and a constitutive gene expression was seen in all mouse organs and in all mouse tissue-derived mRNA analyzed. By detecting the expression of the house keeping gene G3PDH under the above-mentioned PCR conditions using the mouse G3PDH primer for all the templates used in the analysis, it has been verified beforehand that the number of copies of template mRNA has been normalized (standardized) between samples. The detected RT-PCR product size herein was 320 bp, and this coincides with the size calculated by the determined nucleotide sequence. Therefore, it was thought to be the product of the mouse NR8 specific PCR amplification reaction. To further verify this, the PCR product amplified in the day 17 embryo was subcloned to pGEM-T Easy vector according to 7-2), and the nucleotide sequence was analyzed. The result verified that the PCR product could be a partial nucleotide sequence of mouse NR8, and the possibility that it might be the product of a non-specific PCR amplification was denied.

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8-2) Analysis of mouse NR8 gene expression by Northern blotting

In order to analyze NR8 gene expression in each mouse organ, and with the objective of identifying the NR8 transcription size, gene expression analysis by the Northern blotting method was conducted.

Mouse Multiple Tissue Northern Blot (Clontech #7762-1) was used as the blot. Among the 5'-RACE products obtained in 7-4), the mNR8 β cDNA fragment was used as the probe. The probe was radiolabeled with [α -³²P] dCTP (Amersham, cat#AA0005) using Mega Prime Kit (Amersham, cat#RPN1607). Express Hyb Hybridization Solution (Clontech #8015-2) was used for hybridization. After a prehybridization at 68°C for 30 min, the heat-denatured labeled probe was added, and hybridization was conducted at 68°C for 16 hr. After washing under the following conditions, the blot was exposed to Imaging Plate (FUJI #BAS-III), and a mouse NR8 specific signal was detected by the Image Analyzer (FUJIX, BAS-2000 II).

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Washing conditions were: (1) 1x SSC/0.1% SDS, at room temperature for 5 min; (2) 1x SSC/0.1% SDS, at 50°C 30 min; and (3) 0.5x SSC/0.1% SDS, at 50°C 30 min.

As a result, as shown in Fig. 19, a strong expression was seen in the mouse testis only, and no gene expression of the same gene was detected in other organs. Here, there is a difference between the results of RT-PCR analysis and Northern blot analysis. Since the
 5 detection sensitivity of the Northern method is much lower than RT-PCR, it is thought that mRNA with low expression levels could not be detected. However, results of both analyses coincide in the point that a strong gene expression was detected in the testis. Also, the size of the detected transcript was about 4.2 kb.

10 Although there was a deviation of the expression levels in each mouse organ analyzed by the Northern method and RT-PCR, the gene expression was widely distributed, being detectable in all the organs analyzed especially when using RT-PCR. This result contrasts with the human NR8 gene in which the expression was strong only in
 15 immunocompetent tissues, hemopoietic tissues, and specific leukemic cell lines, and the significance of this expression is extremely interesting. This means namely the possibilities that in mouse, the NR8 molecule not only is involved in systemic hemopoietic functions, or in immunological responses, and hemopoiesis, but also may be
 20 involved in various physiological regulatory mechanisms of the body. Namely, its ligand may be able to function as a hormone-like factor.

Example 9: Isolation of the NR8 mouse genomic gene by plaque screening

The present inventors analyzed the genomic structure of mouse NR8
 25 gene and performed plaque hybridization against the mouse genomic DNA library. 129SVJ strain Genomic DNA (Stratagene #946313) constructed in LambdaFIX II was used as the library. This genomic library of approximately 5.0×10^5 plaques was developed and blotted to a Hybond N (+) (Amersham #RPN303B) charged nylon membrane to perform
 30 primary screening. NR8 β cDNA fragment of 5'-RACE products obtained in 7-4) was used as the probe. The probe was radiolabeled with [α - 32 P] dCTP prepared as above-mentioned in 8-2) using the Mega Prime Kit. Express Hyb Hybridization Solution was used for hybridization, and after a prehybridization at 65°C for 30 min, a heat-denatured labeled
 35 probe was added, and hybridization was done at 65°C for 16 hr. After washing under the following conditions, the membrane was exposed to an X-ray film (Kodak, cat#165-1512) to detect mouse NR8 positive plaques.

Washing conditions were: (1) 1x SSC/0.1% SDS, at room temperature for 5 min; (2) 1x SSC/0.1% SDS, at 58°C 30 min; and (3) 0.5x SSC/0.1% SDS, at 58°C 30 min.

As a result, positive, or pseudo-positive 16 independent clones were obtained. When a secondary screening was similarly conducted against these 16 clones obtained by the primary screening, the inventors succeeded in isolating NR8 positive, nine independent plaque clones.

10 Industrial Applicability

The present invention provides a novel hemopoietin receptor protein "NR8," and the encoding DNA. The present invention also provides, a vector into which the DNA has been inserted, a transformant harboring the DNA, and a method of producing a recombinant protein using the transformant. It also provides a method of screening a compound or a natural ligand that binds to the protein. The NR8 protein of the invention is thought to be related to hemopoiesis, and therefore, is useful in analyzing hemopoietic functions. The protein would also be applied in the diagnosis and treatment of hemopoiesis-associated diseases.

Since the expression of mouse NR8 gene was widely distributed in mouse organs, mouse NR8 protein would be involved in various physiological regulatory mechanisms of the body, including the above-mentioned hemopoiesis. Furthermore, by using mouse NR8 protein, it is possible to isolate first the mouse NR8 ligand, and next, the human homologue of the NR8 ligand using the conserved structure of the mouse NR8 ligand. Specifically, after determining the nucleotide sequence of mouse NR8 ligand cDNA, an oligonucleotide primer is designed on this sequence, and using this to conduct cross PCR using the human-derived cDNA library as the template, human NR8 ligand cDNA can be obtained. Alternatively, human NR8 ligand cDNA can be obtained by conducting cross hybridization against human-derived cDNA library using mouse NR8 ligand cDNA as the probe. It is also possible to analyze biological function of the NR8 receptor protein by creating a mouse NR8 gene-deficient mouse using the mouse NR8 gene.

CLAIMS

1. A protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1, or a protein
5 comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO:
10 1.
2. A protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3, or a protein
15 comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO:
3.
- 20 3. A protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from
25 the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO:
5.
- 30 4. A protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO:
7.
- 35 5. A protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added

and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19.

- 5 6. A protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21.
- 10 7. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 2, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1.
- 15 8. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 4, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3.
- 20 9. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 6, which is functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5.
- 25 10. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 8, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7.
- 30 11. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 20, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19.
- 35 12. A protein encoded by a DNA hybridizing to a DNA comprising the

nucleotide sequence of SEQ ID NO: 22, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21.

- 5 13. A fusion protein comprising the protein of any one of claims 1 to 12 and another peptide or polypeptide.
14. A DNA encoding the protein of any one of claims 1 to 13.
15. A vector comprising the DNA of claim 14.
- 10 16. A transformant harboring the DNA of claim 14 in an expressible manner.
17. A method of producing the protein of any one of claims 1 to 13, comprising the step of culturing the transformant of claim 16.
18. A method of screening a compound that binds to the protein of any one of claims 1 to 13 comprising the steps of:
 - 15 (a) contacting a test sample with the protein of any one of claims 1 to 13; and
 - (b) selecting a compound that comprises an activity to bind to the protein of any one of claims 1 to 13.
19. An antibody that specifically binds to the protein of any one of claims 1 to 12.
- 20 20. A method of detecting or measuring the protein of any one of claims 1 to 13 comprising the steps of contacting a test sample presumed to contain said protein with the antibody of claim 19, and detecting or measuring the formation of the immune complex between the antibody and the protein.
- 25 21. A DNA specifically hybridizing to a DNA comprising a nucleotide sequence of any one of SEQ ID NOs: 2, 4, 6, 8, 20, and 22 to 27 comprising at least 15 nucleotides, and comprising at least 15 nucleotides.

30

ABSTRACT

The present invention provides novel hemopoietin receptor proteins (proteins comprising the amino acid sequence of SEQ ID NOs: 1, 3, 5, 7, 19, or 21), proteins comprising a modified amino acid sequence of the amino acid sequence of the above protein in which one or more amino acids have been deleted, added, and/or replaced with another amino acid, genes encoding these proteins, methods of producing the proteins, as well as uses of these proteins and genes.

Figure 1

NR8 ⁺	40862	<u>SLLPLEFRKDSSVELQVRAGMPGSSYQGTWSEWSDPVIFQIQSEGRCEAGMDTPLL</u>	41032
hTPOR	442	<u>LELRPRSRYLQLRAR-LNGPTYQGPWSSWSDPTRVEIATE</u>	481
hOBR	292	<u>SLLVDSILPGSSYEVRGKRLDGP----</u> <u>GIWSDWSTPRVFTIQ</u>	331
hIL2Rb	201	<u>DTQVEFQVRVKPLQGEFT--TWSPWSQPLAFRIK</u>	232
hIL7R	189	<u>TLLQRKLQPAAMYEIKVRS--IPDHYFKGFWEWSPSYFRIPENNSSGEMDPILL</u>	243
hGM-CSFRb	196	<u>TLGPEHLMPSSTYVARVTRLAPGSRLSGRPSKWSPEVCWDSQ</u>	238
	419	<u>TGYNGIWSEWSEARSWDIES</u>	438
mIL3Rb	200	<u>NLEPKLFLPNSIYAARVTRLSAGSSLSGRPSRWSPVEVHWSQ</u>	242
	404	<u>QLEPDTSYCARVRVKPI--SDYDGIWSEWSNEYTWII</u>	438
hIL5Ra	302	<u>SKYDVQVRAAVSSMCREAGLWSEWSQPI</u>	329
hIL9R	241	<u>YTGQWSEWSQPVCFQ</u>	255
hEPOR	211	<u>RGRTRYTFAVRAR-MAEPSFGGFWSAWSEPVSLLIPSD</u>	247
hIL2Rr	209	<u>SLPSVDGQKRYTFRVRSRFNPLCGSAQH--WSEWSHPPI</u>	244
hIL12R	197	<u>LCPLEMNVAQEFQLRRQLGSGQSS-----WSKWSSPV</u>	229
hIL12Rb	282	<u>LDLKPFTYEYEFQISSKL---HLKYGSWSDWSES LRAQTPEE</u>	319

Figure 2

[Query: 39181-39360]

NR8	39233	<u>HQVKPAPPFN</u> — <u>VTVTFSGQYNISWRS-DYEDP</u> —— <u>AFYMLKGKLQY</u>	39355
hIL6Ra	214	<u>LQDPPANI</u> — <u>TVTAVAR-NPRWLSVTWQDPHSWNSSFYRLRFELRY</u>	257
hgp130	218	<u>YKVKPNPPHNL</u> — <u>SVINSEELSSILKLTWT-NPSIKSV</u> — <u>IILKYNIQY</u>	261
rOBRb	234	<u>VKDPPLGLRMEVTDDGNLKISWDS</u> -QTKAP	263

[Query: 42301-42480]

NR8	42307	<u>VPSPERFFMPLYKGCSGDFK</u>	42366
mIL9R	305	<u>IPSPEAFFHPLYSVYHGDFQ</u>	324
hIL9R	305	<u>VPSPAMFFQPLYSVHNGNFQ</u>	324

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Figure 3

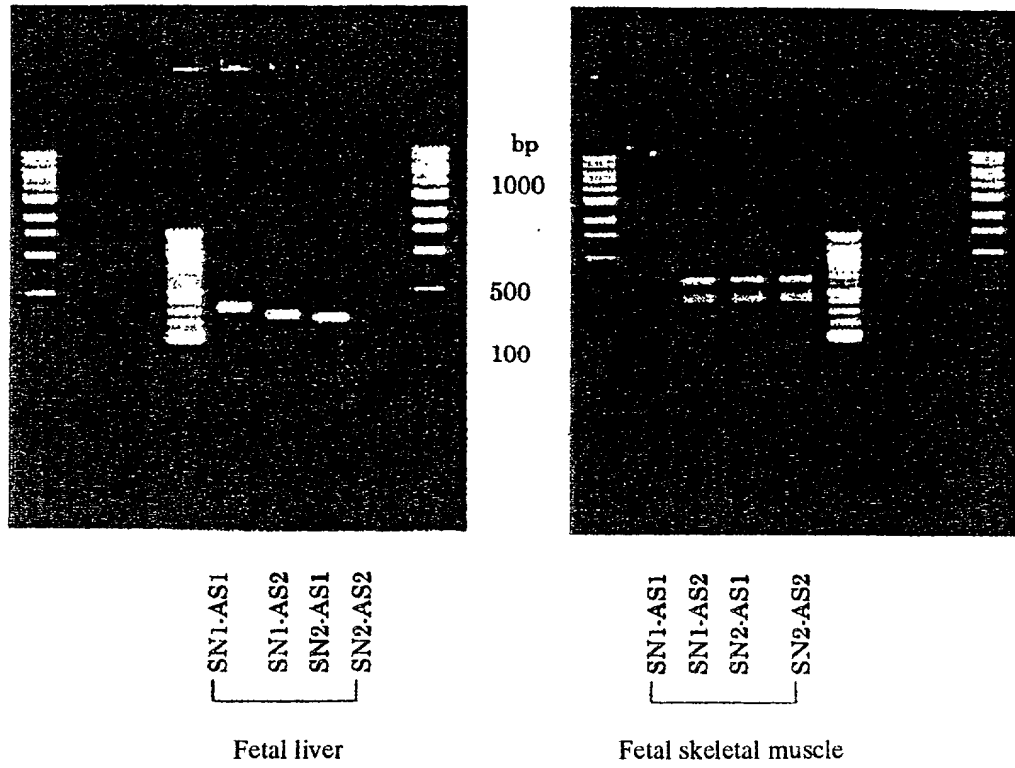


Figure 4

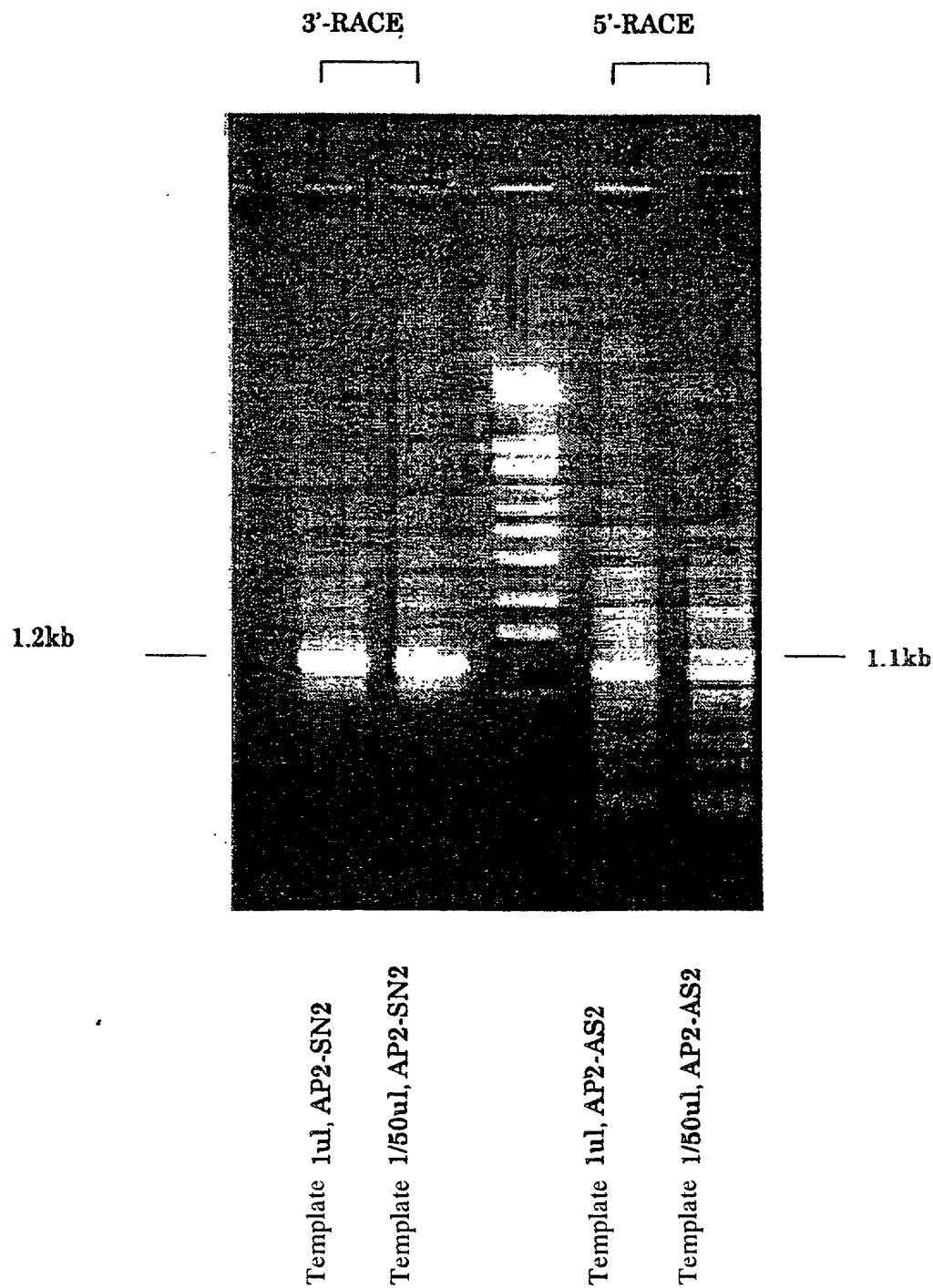


Figure 5

10 20 30 40 50 60 70 80
 GGCAGCCAGCGGCTCAGACAGACCCACTGGCGTCTCTGCTGAGTGACCGTAAGCTCGGCGTCTGGCCCTCTGCCTGC
 90 100 110 120 130 140 150 160
 CTCTCCCTGAGTGTGGCTGACAGCCACGCAGCTGTGTCTGTCTGTCTGCGGCCCGTGCATCCCTGCTCGGCGCGCTGGT
 170 180 190 200 210 220 230 240
 ACCTTCCTTGCCGTCTCTTTCCTGTGTCTGCTCTGTGGGACACCTGCCTGGAGGCCAGCTGCCCGTCATCAGAGTG
 250 260 270 280 290 300 310 320
 ACAGGTCTTATGACAGCCTGATTGGTGACTCGGGCTGGGTGTGGATTCTCAGCCAGGCTCTGCCTGCTTTCTCAGACC
 330 340 350 360 370 380 390 400
 CTCATCTGTACCCCCACGCTGAACCCAGCTGCCACCCCCAGAAGCCCATCAGACTGCCCCAGCACAGGAATGGATT
 410 420 430 440 450 460 470 480
 CTGAGAAAGAAGCCGAAACAGAGGCCCGTGGGAGTCAGCATGCCGCGTGGCTGGGCGGCCCCCTTGCTCCTGCTGCTGC
 M P R G W A A P L L L L L L
 490 500 510 520 530 540 550 560
 TCCAGGGAGGCTGGGGCTGCCCGACCTCGTCTGTCTACACCGATTACCTCCAGACGGTCATCTGCATCCTGGAAATGTGG
 Q G G W G C P D L V C Y T D Y L Q T V I C I L E M W
 570 580 590 600 610 620 630 640
 AACCTCCACCCAGCAGCTCACCTTACCTGGCAAGACCAGTATGAAGAGCTGAAGGACGAGGCCACCTCCTGCAGCCT
 N L H P S T L T L T W Q D Q Y E E L K D E A T S C S L
 650 660 670 680 690 700 710 720
 CCACAGGTGCGCCACAATGCCACGCATGCCACCTACACCTGCCACATGGATGTATTCCACTTCATGGCCGACGACATTT
 H R S A H N A T H A T Y T C H M D V F H F M A D D I F
 730 740 750 760 770 780 790 800
 TCAGTGTCAACATCACAGACCAGTCTGGCAACTACTCCAGGAGTGTGGCAGCTTTCTCCTGGCTGAGAGCATCAAGCCG
 S V N I T D Q S G N Y S Q E C G S F L L A E S I K P
 810 820 830 840 850 860 870 880
 GCTCCCCCTTTCAACGTGACTGTGACCTTCTCAGGACAGTATAATATCTCCTGGCGCTCAGATTACGAAGACCCTGCCTT
 A P P F N V T V T F S G Q Y N I S W R S D Y E D P A F
 890 900 910 920 930 940 950 960
 CTACATGCTGAAGGGCAAGCTTCAGTATGAGCTGCAGTACAGGAACCGGGAGACCCCTGGGCTGTGAGTCCGAGGAGAA
 Y M L K G K L Q Y E L Q Y R N R G D P W A V S P R R K
 970 980 990 1000 1010 1020 1030 1040
 AGCTGATCTCAGTGGACTCAAGAAGTGTCTCCCTCCTCCCGTGGAGTTCGCAAGACTCGAGCTATGAGCTGCAGGTG
 L I S V D S R S V S L L P L E F R K D S S Y E L Q V
 1050 1060 1070 1080 1090 1100 1110 1120
 CGGGCAGGGCCCATGCCTGGCTCCTCCTACCAGGGGACCTGGAGTGAATGGAGTGACCCGGTCATCTTTCAGACCCAGTC
 R A G P M P G S S Y Q G T W S E W S D P V I F Q T Q S

Figure 6

1130	1140	1150	1160	1170	1180	1190	1200
AGAGGAGTTAAAGGAAGGCTGGAACCCCTCACCTGCTGCTTCTCCTCCTGCTTGTGCATAGTCTTCATTCCTGCCTTCTGGA							
E	E	L	K	E	G	W	N
1210	1220	1230	1240	1250	1260	1270	1280
GCCTGAAGACCCATCCATTGTGGAGGCTATGGAAGAAGATATGGGCCGTCCCAGCCCTGAGCGGTTCTTCATGCCCTTG							
L	K	T	H	P	L	W	R
1290	1300	1310	1320	1330	1340	1350	1360
TACAAGGGCTGCAGCGGAGACTTCAAGAAATGGGTGGGTGCACCCTTCACTGGCTCCAGCCTGGAGCTGGGACCCTGGAG							
Y	K	G	C	S	G	D	F
1370	1380	1390	1400	1410	1420	1430	1440
CCCAGAGGTGCCCTCCACCTTGGAGGTGTACAGCTGCCACCCACCCAGCAGCCCTGTGGAGTGTGACTTCACCAGCCCCG							
P	E	V	P	S	T	L	E
1450	1460	1470	1480	1490	1500	1510	1520
GGGACGAAGGACCCCCCGGAGCTACCTCCGCCAGTGGGTGGTCACTTCTCCGCCACTTTCGAGCCCTGGACCCCAGGCC							
D	E	G	P	P	R	S	Y
1530	1540	1550	1560	1570	1580	1590	1600
AGCTAATGAGGCTGACTGGATGTCCAGAGCTGGCCAGGCCACTGGGCCCTGAGCCAGAGACAAGGTACCTGGGCTGTGA							
S	*	*					
1610	1620	1630	1640	1650	1660	1670	1680
TGTGAAGACACCTGCAGCCTTTGGTCTCCTGGATGGGCCTTTGAGCCTGATGTTACAGTGTCTGTGTGTGTGCATAT							
1690	1700	1710	1720	1730	1740	1750	1760
GTGTGTGTGTGCATATGCATGTGTGTGTGTGTGTGTGTCTTAGGTGCGCAGTGGCATGTCCACGTGTGTGTGATTGCACG							
1770	1780	1790	1800	1810	1820	1830	1840
TGCTGTGGGCTGGGATAATGCCCATGGTACTCCATGCATTACCTGCCCTGTGCATGTCTGGACTCACGGAGCTCACC							
1850	1860	1870	1880	1890	1900	1910	1920
CATGTGCACAAGTGTGCACAGTAAACGTGTTTGTGGTCAACAGAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA							
1930							
AAAAAAAAAAAAAA							

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Figure 7

10 20 30 40 50 60 70 80
GGCAGCCAGCGGCTCAGACAGACCCACTGGCGTCTCTGCTGAGTGACCGTAAGCTCGGCGTCTGGCCCTCTGCCTGC

90 100 110 120 130 140 150 160
CTCTCCCTGAGTGTGGCTGACAGCCACGCAGCTGTGTCTGTCTGTCTGCGGCCGTGCATCCCTGCTGCGGCCGCTGGT

170 180 190 200 210 220 230 240
ACCTTCCTTGCCGTCTCTTTCTCTGTCTGCTGCTCTGTGGGACACCTGCCTGGAGGCCAGCTGCCCGTCATCAGAGTG

250 260 270 280 290 300 310 320
ACAGGTCTTATGACAGCCTGATTGGTGACTCGGGCTGGGTGTGGATTCTACCCCAGGCCTCTGCCTGCTTTCTCAGACC

330 340 350 360 370 380 390 400
CTCATCTGTACCCCCACGCTGAACCCAGCTGCCACCCCAGAAGCCCATCAGACTGCCCCAGCACACGGAATGGATT

410 420 430 440 450 460 470 480
CTGAGAAAGAAGCCGAAACAGAAGGCCGTGGGAGTCAGCATGCCCGTGGCTGGGCCGCCCTTGTCTCTGCTGCTGC
M P R G W A A P L L L L L L

490 500 510 520 530 540 550 560
TCCAGGGAGGCTGGGGCTGCCCCGACCTCGTCTGCTACACCGATTACCTCCAGACGGTCATCTGCATCCTGGAAATGTGG
Q G G W G C P D L V C Y T D Y L Q T V I C I L E M W

570 580 590 600 610 620 630 640
AACCTCCACCCAGCAGCTCACCTTACCTGGCAAGACAGTATGAAGAGCTGAAGGACGAGGCCACCTCCTGCAGCCT
N L H P S T L T L T W Q D Q Y E E L K D E A T S C S L

650 660 670 680 690 700 710 720
CCACAGGTGGGCCACAATGCCACGCATGCCACCTACACCTGCCACATGGATGTATTCCACTTCATGGCCGACGACATTT
H R S A H N A T H A T Y T C H M D V F H F M A D D I F
M P R M P P T P A T - W M Y S T S W P T T F

730 740 750 760 770 780 790 800
TCAGTGTCAACATCACAGACCAGTCTGGCAACTACTCCCAGGAGTGTGGCAGCTTTCTCCTGGCTGAGAGCAAGTCCGAG
S V N I T D Q S G N Y S Q E C G S F L L A E S K S E
S V S T S Q T S L A T T P R S V A A F S W L R A S P R

810 820 830 840 850 860 870 880
GAGAAAGCTGATCTCAGTGGACTCAAGAAAGTGTCTCCCTCCTCCCCCTGGAGTTCGCAAAGACTCGAGCTATGAGCTGC
E K A D L S G L K K C L P P P P G V P Q R L E L *
R K L I S V D S R S V S L L P L E F R K D S S Y E L Q

890 900 910 920 930 940 950 960
AGGTGCGGGCAGGGCCCATGCCTGGCTCCTCCTACCAGGGACCTGGAGTGAATGGAGTGACCCGGTCATCTTTCAGACC
V R A G P M P G S S Y Q G T W S E W S D P V I F Q T

Figure 8

970 980 990 1000 1010 1020 1030 1040
CAGTCAGAGGAGTTAAAGGAAGGCTGGAACCCTCACCTGCTGCTTCTCCTCCTGCTTGCATAGTCTTCATTCTGCCTT

Q S E E L K E G W N P H L L L L L L V I V F I P A F
1050 1060 1070 1080 1090 1100 1110 1120
CTGGAGCCTGAAGACCCATCCATTGTGGAGGCTATGGAAGAAGATATGGGCCGTCCCAGCCCTGAGCGTTCTTCATGC

W S L K T H P L W R L W K K I W A V P S P E R F F M P
1130 1140 1150 1160 1170 1180 1190 1200
CCCTGTACAAGGGCTGCAGCGGAGACTTCAAGAAATGGGTGGGTGCACCCTTCACTGGCTCCAGCCTGGAGCTGGGACCC

L Y K G C S G D F K K W V G A P F T G S S L E L G P
1210 1220 1230 1240 1250 1260 1270 1280
TGGAGCCCAGAGGTGCCCTCCACCCTGGAGGTGTACAGCTGCCACCCACCCAGCAGCCCTGTGGAGTGTGACTTCACCAG

W S P E V P S T L E V Y S C H P P S S P V E C D F T S
1290 1300 1310 1320 1330 1340 1350 1360
CCCCGGGGACGAAGGACCCCCGGGAGCTACCTCCGCCAGTGGGTGGTTCATTCTCCGCCACTTTCGAGCCCTGGACCCC

P G D E G P P R S Y L R Q W V V I P P P L S S P G P Q
1370 1380 1390 1400 1410 1420 1430 1440
AGGCCAGCTAATGAGGCTGACTGGATGTCCAGAGCTGGCCAGGCCACTGGGCCCTGAGCCAGAGACAAGGTCACCTGGGC

A S * *
1450 1460 1470 1480 1490 1500 1510 1520
TGTGATGTGAAGACACCTGCAGCCTTTGGTCTCCTGGATGGGCCTTTGAGCCTGATGTTTACAGTGTCTGTGTGTGTG

1530 1540 1550 1560 1570 1580 1590 1600
CATATGTGTGTGTGTGCATATGCATGTGTGTGTGTGTGTCTTAGGTGCGCAGTGGCATGTCCACGTGTGTGTGATT

1610 1620 1630 1640 1650 1660 1670 1680
GCACGTGCCTGTGGGCCTGGGATAATGCCCATGGTACTCCATGCATTACCTGCCCTGTGCATGTCTGGACTCACGGAGC

1690 1700 1710 1720 1730 1740 1750 1760
TCACCCATGTGCACAAGTGTGCACAGTAAACGTGTTTGTGTTCAACAGAAAAAAAAAAAAAAAAAAAAAAAAAAAAA

1770 1780
AAAAAAAAAAAAAAAAAAAA

Figure 9

10 20 30 40 50 60 70 80
GGCAGCCAGCGGCTCAGACAGACCCACTGGCGTCTCTCTGCTGAGTGACCGTAAGCTCGGCGTCTGGCCCTCTGCCTGC

90 100 110 120 130 140 150 160
CTCTCCCTGAGTGTGGCTGACAGCCAGCGAGCTGTGTCTGTCTGTCTGGGCGCGTGCATCCCTGCTGCGGCCGCTGCT

170 180 190 200 210 220 230 240
ACCTTCCTTGCCGTCTCTTTCTCTGTCTGCTGCTCTGTGGGACACCTGCCTGGAGGCCAGCTGCCGTCATCAGAGTG

250 260 270 280 290 300 310 320
ACAGGTCTTATGACAGCCTGATTGGTGACTCGGGCTGGGTGTGGATTCTCAGCCAGGCTCTGCCTGCTTTCTCAGACC

330 340 350 360 370 380 390 400
CTCATCTGTACCCCCAGCTGAACCCAGCTGCCACCCCAAGAAGCCATCAGACTGCCCCAGCACAGGAATGGATT

410 420 430 440 450 460 470 480
CTGAGAAAGAAGCCGAACAGAAGGCCCGTGGGAGTCAGCATGCCGCGTGGCTGGGCCGCCCCCTTGCTCCTGCTGCTGC
M P R G W A A P L L L L L L

490 500 510 520 530 540 550 560
TCCAGGGAGGCTGGGGCTGCCCGACCTCGTCTGTACACCGATTACCTCCAGACGGTCATCTGCATCCTGGAAATGTGG
Q G G W G C P D L V C Y T D Y L Q T V I C I L E M W

570 580 590 600 610 620 630 640
AACCTCCACCCAGCAGCCTCACCTTACCTGGCAAGACCAGTATGAAGAGCTGAAGGACGAGGCCACCTCCTGCAGCCT
N L H P S T L T L T W Q D Q Y E E L K D E A T S C S L

650 660 670 680 690 700 710 720
CCACAGGTGGGCCACAATGCCAGCATGCCACCTACACCTGCCACATGGATGTATTCCACTTCATGGCCGACGACATTT
H R S A H N A T H A T Y T C H M D V F H F M A D D I F

730 740 750 760 770 780 790 800
TCAGTGTCAACATCACAGACCAGTCTGGCAACTACTCCAGGAGTGTGGCAGCTTTCTCCTGGCTGAGAGCATCAAGCCG
S V N I T D Q S G N Y S Q E C G S F L L A E S I K P

810 820 830 840 850 860 870 880
GCTCCCCCTTTCAACGTGACTGTGACCTTCTCAGGACAGTATAATATCTCCTGGCGCTCAGATTACGAAGACCCTGCCTT
A P P F N V T V T F S G Q Y N I S W R S D Y E D P A F

890 900 910 920 930 940 950 960
CTACATGCTGAAGGGCAAGCTTCAGTATGAGCTGAGTACAGGAACCGGGGAGACCCCTGGGCTGTGAGTCCGAGGAGAA
Y M L K G K L Q Y E L Q Y R N R G D P W A V S P R R K

970 980 990 1000 1010 1020 1030 1040
AGCTGATCTCAGTGGACTCAAGAAGTGTCTCCCTCCTCCCCCTGGAGTTCGCAAGACTCGAGCTATGAGCTGCAGGTG
L I S V D S R S V S L L P L E F R K D S S Y E L Q V

1050 1060 1070 1080 1090 1100 1110 1120
CGGGCAGGGCCCATGCCTGGCTCCTCCTACCAGGGACCTGGAGTGAATGGAGTGACCCGGTCATCTTCAGACCCAGTC
R A G P M P G S S Y Q G T W S E W S D P V I F Q T Q S

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Figure 10

1130 1140 1150 1160 1170 1180 1190 1200
 AGAGGAGTTAAAGGAAGGCTGGAACCCCTCACCTGCTGCTTCTCCTCCTGCTGTGCATAGTCTTCATTCTGCCTTCTGGA
 E E L K E G W N P H L L L L L L L V I V F I P A F W S
 1210 1220 1230 1240 1250 1260 1270 1280
 GCCTGAAGACCCATCCATTGTGGAGGCTATGGAAGAAGATATGGGCCGTCCCAGCCCTGAGCGGTTCTTCATGCCCTG
 L K T H P L W R L W K K I W A V P S P E R F F M P L
 1290 1300 1310 1320 1330 1340 1350 1360
 TACAAGGGCTGCAGCGGAGACTTCAAGAAATGGGTGGGTGCACCCCTTCACTGGCTCCAGCCTGGAGCTGGGACCCCTGGAG
 Y K G C S G D F K K W V G A P F T G S S L E L G P W S
 1370 1380 1390 1400 1410 1420 1430 1440
 CCCAGAGGTGCCCTCCACCCTGGAGGTGTACAGCTGCCACCCACCGAGCCCGCCAAGAGGCTGCAGCTCAGGAGC
 P E V P S T L E V Y S C H P P R S P A K R L Q L T E L
 1450 1460 1470 1480 1490 1500 1510 1520
 TACAAGAACCAGCAGAGCTGGTGGAGTCTGACGGTGTGCCAAGCCAGCTTCTGGCCGACAGCCAGAAGCTCGGGGGG
 Q E P A E L V E S D G V P K P S F W P T A Q N S G G

1530 1540 1550 1560 1570 1580 1590 1600
 TCAGCTTACAGTGAGGAGAGGGATCGGCCATACGGCCTGGTGTCCATTGACACAGTGACTGTGCTAGATGCAGAGGGGCC
 S A Y S E E R D R P Y G L V S I D T V T V L D A E G P

1610 1620 1630 1640 1650 1660 1670 1680
 ATGCACCTGGCCCTGCAGCTGTGAGGATGACGGCTACCCAGCCCTGGACCTGGATGCTGGCCTGGAGCCAGCCAGGCC
 C T W P C S C E D D G Y P A L D L D A G L E P S P G L

1690 1700 1710 1720 1730 1740 1750 1760
 TAGAGGACCACTCTTGGATGCAGGGACCACAGTCTGTCTGTGGCTGTGTCTCAGCTGGCAGCCCTGGGCTAGGAGGG
 E D P L L D A G T T V L S C G C V S A G S P G L G G

1770 1780 1790 1800 1810 1820 1830 1840
 CCCCTGGGAAGCCTCCTGGACAGACTAAAGCCACCCCTTGACAGATGGGGAGGACTGGGCTGGGGGACTGCCCTGGGGTGG
 P L G S L L D R L K P P L A D G E D W A G G L P W G G

1850 1860 1870 1880 1890 1900 1910 1920
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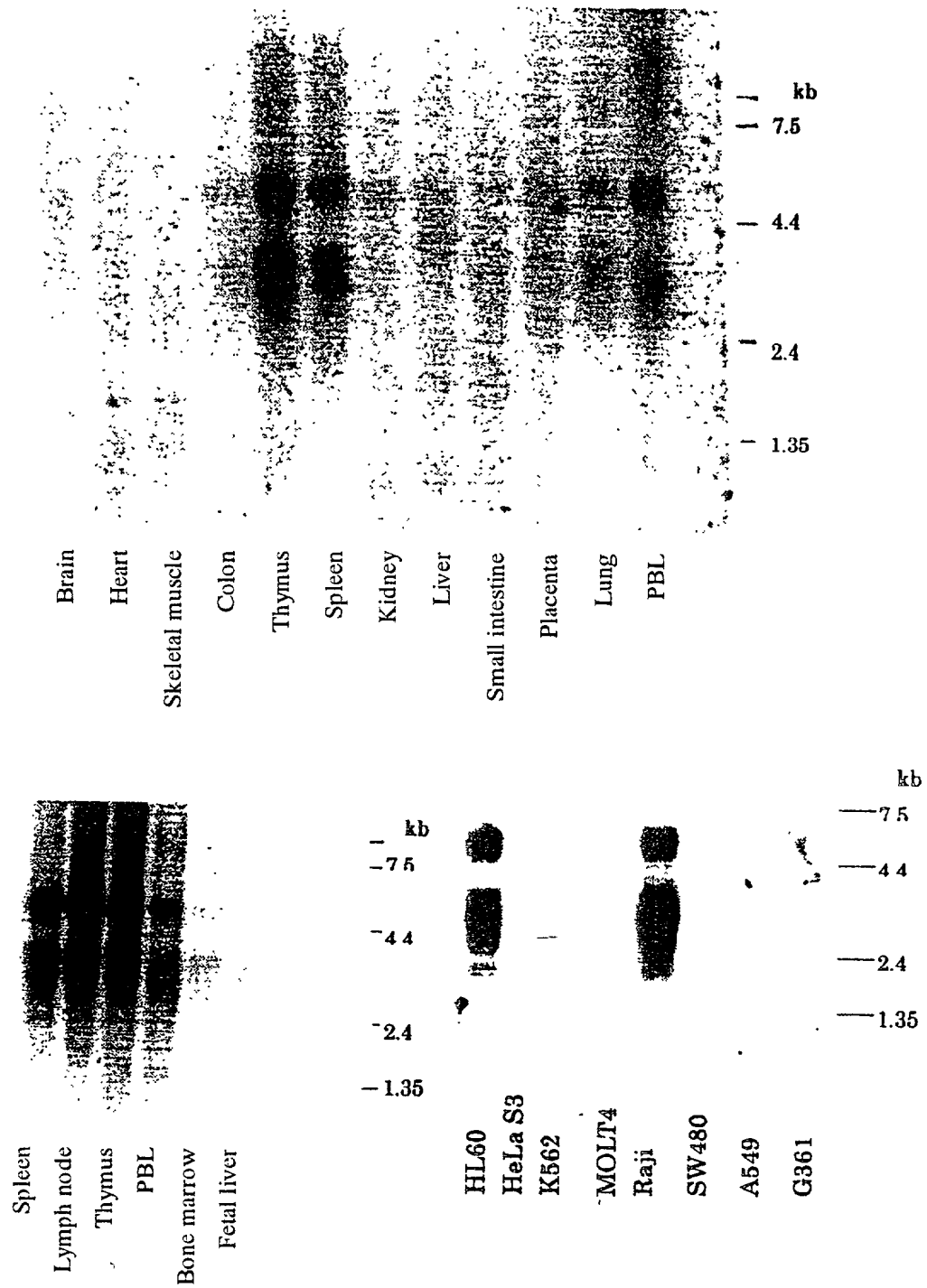
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2010 2020 2030 2040 2050 2060 2070 2080
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 R Q W V V I P P P L S S P G P Q A S * *

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TGGATGGGCCTTTGAGCCTGATGTTACAGTGTCTGTGTGTGTGTCATATGTGTGTGTGTGTCATATGCATGTGTGTGTG							
2250	2260	2270	2280	2290	2300	2310	2320
TGTGTGTGTCTTAGGTGCGCAGTGGCATGTCCACGTGTGTGTGATTGCACGTGCCTGTGGGCCTGGGATAATGCCCATGG							
2330	2340	2350	2360	2370	2380	2390	2400
TACTCCATGCATTACCTGCCCTGTGCATGTCTGGACTCAGGAGCTCACCCATGTGCACAAGTGTGCACAGTAAACGTG							
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Figure 12



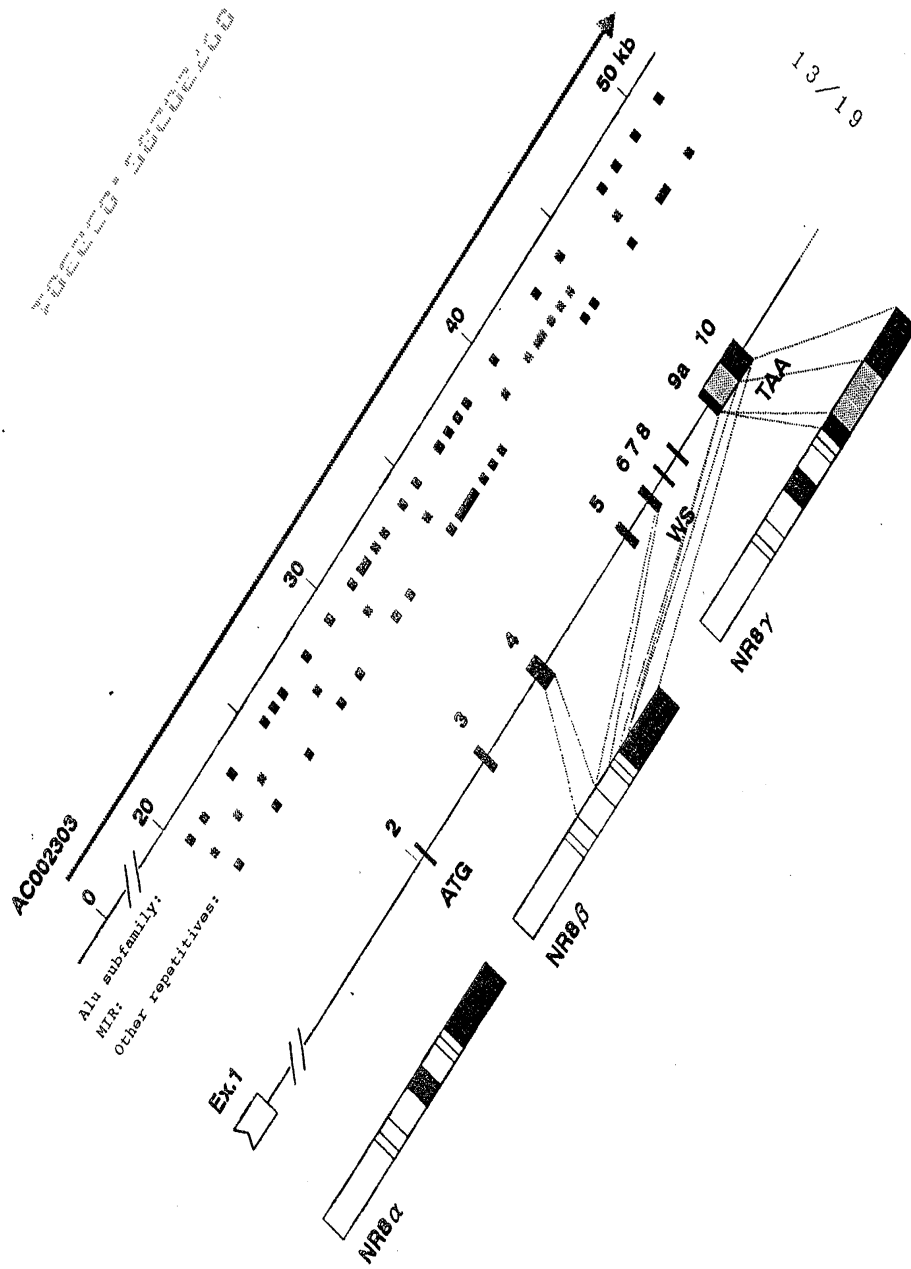


Figure 13

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Figure 14

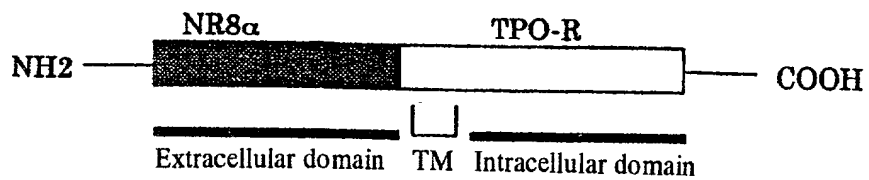
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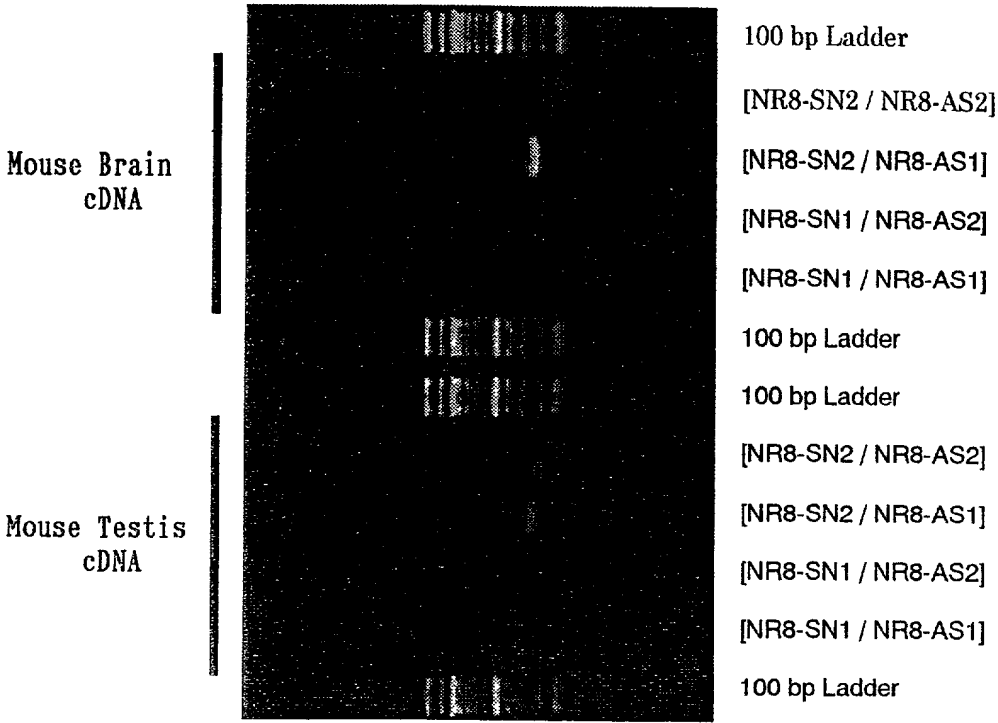


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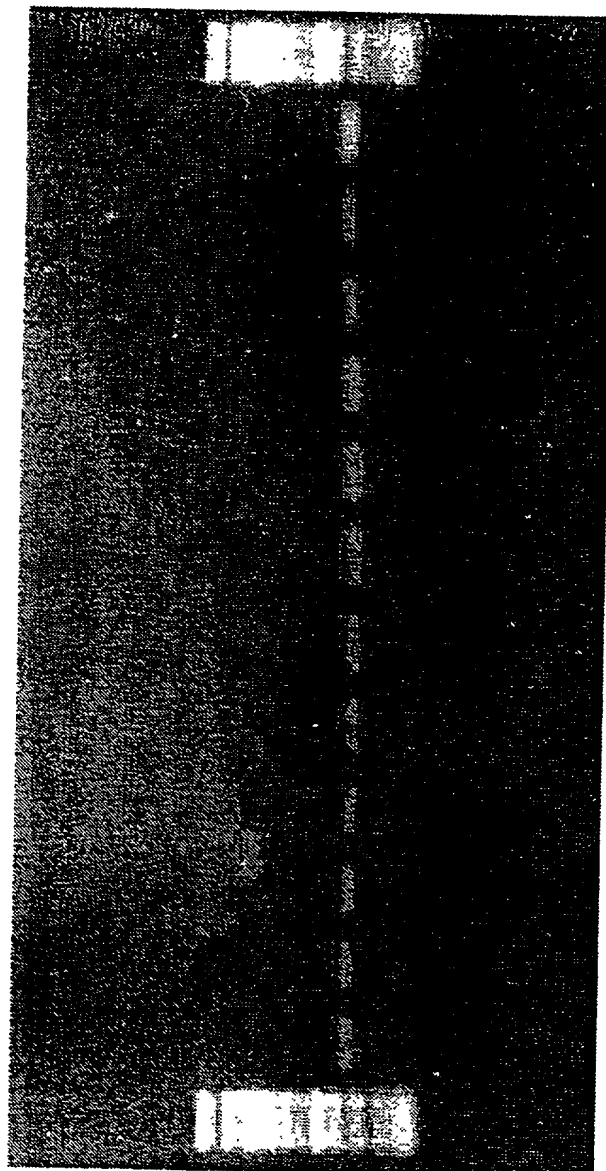
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mNR8BETA	NLHPSTLTLT	WDDQYEELKD	EATSCSLHRS	AHNATHATYT	80
hNR8BETA	CHMDVFHMA	DDIFSVNITD	OSGNYSQECG	SFLAESKSE	120
mNR8BETA	SHMDVFHMA	DDIFSVNITD	OSGNYSQECG	SFLAESKSE	120
hNR8BETA	EKADLSGLKK	CLPPPPGVPO	RLEL		144
mNR8BETA	EKADLSGLKK	CLPPPPGVPO	RLEL		144

Figure 17

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mNR8G	SHMDVFHMA	DDIFSVMITD	QSGNYFQECG	SFLRAESIKP	120
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mNR8G	APPFNVTVTF	SGQYNISRRS	DYEDPAFYML	KGKLOYELOQ	160
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mNR8G	YKGCSDFKK	WVGAFFTGSS	LELGPWSPEV	PSTLEVYSCH	320
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mNR8G	PPRSPAKRLQ	LTELQEPABL	VESDGVPKPS	FWPTAQNSGG	360
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mNR8G	ALDLDAGLEP	SPGLEDPILL	AGTTVLSCGC	VSAGSPGLGG	440
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mNR8G	PLGSLIDRLK	PPLADGEDWA	GGLPWGGRSP	GGVSESEAGS	480
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mNR8G	PLAGLMDTF	DSGFVGSDCS	SPVECDFTSP	GDEGPFRSYL	520
hNR8G	RQWVIPPPL	SSPGEQAS			538
mNR8G	RQWVIPPPL	SSPGEQAS			538

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Figure 18



100 bp Ladder

E17-day

E15-day

E11-day

E7-day

Testis

Kidney

Skeletal muscle

Liver

Lung

Spleen

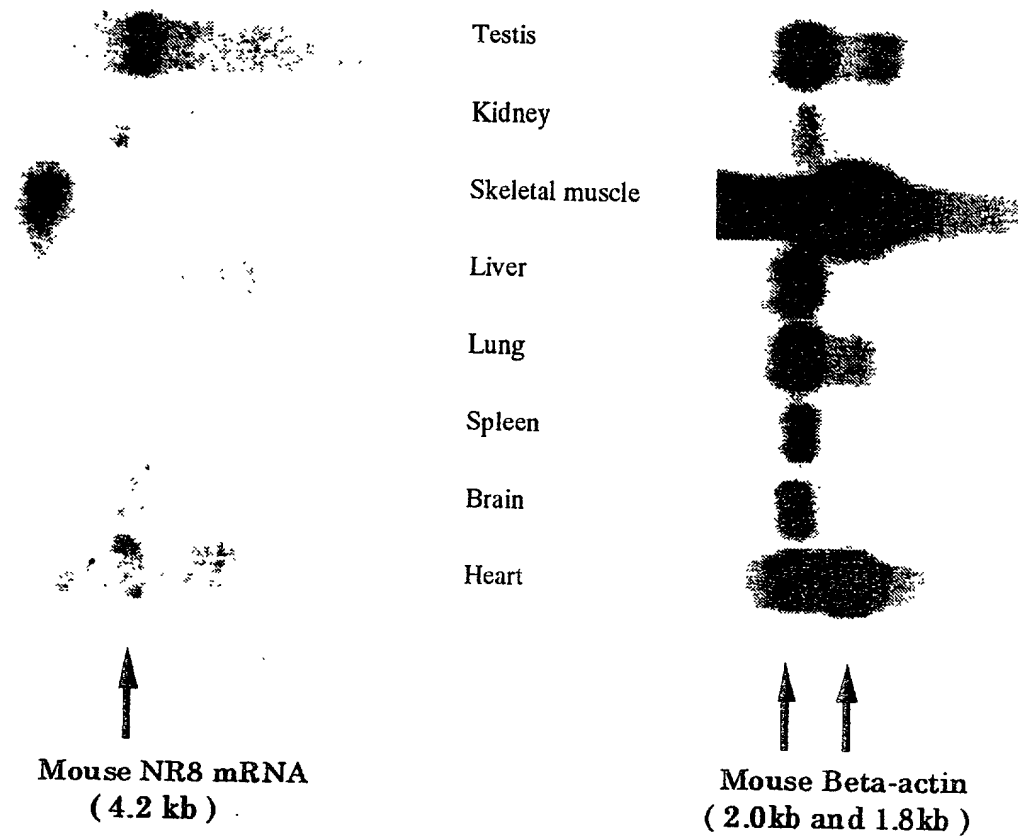
Brain

Heart

100 bp Ladder

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Figure 19



09/720285

JC01 Rec'd PCT/PTO 21 DEC 2000

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Pro Val Ile Phe Gln Thr Gln Ser Glu Glu Leu Lys Glu Gly Trp Asn		220		225		230
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Cys Ser Gly Asp Phe Lys Lys Trp Val Gly Ala Pro Phe Thr Gly Ser		285		290		295
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gcc gtc ccc agc cct gag cgg ttc ttc atg ccc ctg tac aag ggc tgc Ala Val Pro Ser Pro Glu Arg Phe Phe Met Pro Leu Tyr Lys Gly Cys 145 150 155 160	1137
agc gga gac ttc aag aaa tgg gtg ggt gca ccc ttc act ggc tcc agc Ser Gly Asp Phe Lys Lys Trp Val Gly Ala Pro Phe Thr Gly Ser Ser 165 170 175	1185
ctg gag ctg gga ccc tgg agc cca gag gtg ccc tcc acc ctg gag gtg Leu Glu Leu Gly Pro Trp Ser Pro Glu Val Pro Ser Thr Leu Glu Val 180 185 190	1233
tac agc tgc cac cca ccc agc agc cct gtg gag tgt gac ttc acc agc Tyr Ser Cys His Pro Pro Ser Ser Pro Val Glu Cys Asp Phe Thr Ser 195 200 205	1281
ccc ggg gac gaa gga ccc ccc cgg agc tac ctc cgc cag tgg gtg gtc Pro Gly Asp Glu Gly Pro Pro Arg Ser Tyr Leu Arg Gln Trp Val Val 210 215 220	1329
att cct ccg cca ctt tcg agc cct gga ccc cag gcc agc taatgaggct	1378

95	100	105
Glu Cys Gly Ser Phe Leu Leu Ala Glu Ser Ile Lys Pro Ala Pro Pro		
110	115	120
Phe Asn Val Thr Val Thr Phe Ser Gly Gln Tyr Asn Ile Ser Trp Arg		
125	130	135
Ser Asp Tyr Glu Asp Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln		
140	145	150
155		
Tyr Glu Leu Gln Tyr Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro		
160	165	170
Arg Arg Lys Leu Ile Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro		
175	180	185
Leu Glu Phe Arg Lys Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly		
190	195	200
Pro Met Pro Gly Ser Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp		
205	210	215
Pro Val Ile Phe Gln Thr Gln Ser Glu Glu Leu Lys Glu Gly Trp Asn		
220	225	230
235		
Pro His Leu Leu Leu Leu Leu Leu Val Ile Val Phe Ile Pro Ala		
240	245	250
Phe Trp Ser Leu Lys Thr His Pro Leu Trp Arg Leu Trp Lys Lys Ile		
255	260	265
Trp Ala Val Pro Ser Pro Glu Arg Phe Phe Met Pro Leu Tyr Lys Gly		
270	275	280
Cys Ser Gly Asp Phe Lys Lys Trp Val Gly Ala Pro Phe Thr Gly Ser		
285	290	295
Ser Leu Glu Leu Gly Pro Trp Ser Pro Glu Val Pro Ser Thr Leu Glu		
300	305	310
315		

Val Tyr Ser Cys His Pro Pro Arg Ser Pro Ala Lys Arg Leu Gln Leu
320 325 330

Thr Glu Leu Gln Glu Pro Ala Glu Leu Val Glu Ser Asp Gly Val Pro
335 340 345

Lys Pro Ser Phe Trp Pro Thr Ala Gln Asn Ser Gly Gly Ser Ala Tyr
350 355 360

Ser Glu Glu Arg Asp Arg Pro Tyr Gly Leu Val Ser Ile Asp Thr Val
365 370 375

Thr Val Leu Asp Ala Glu Gly Pro Cys Thr Trp Pro Cys Ser Cys Glu
380 385 390 395

Asp Asp Gly Tyr Pro Ala Leu Asp Leu Asp Ala Gly Leu Glu Pro Ser
400 405 410

Pro Gly Leu Glu Asp Pro Leu Leu Asp Ala Gly Thr Thr Val Leu Ser
415 420 425

Cys Gly Cys Val Ser Ala Gly Ser Pro Gly Leu Gly Gly Pro Leu Gly
430 435 440

Ser Leu Leu Asp Arg Leu Lys Pro Pro Leu Ala Asp Gly Glu Asp Trp
445 450 455

Ala Gly Gly Leu Pro Trp Gly Gly Arg Ser Pro Gly Gly Val Ser Glu
460 465 470 475

Ser Glu Ala Gly Ser Pro Leu Ala Gly Leu Asp Met Asp Thr Phe Asp
480 485 490

Ser Gly Phe Val Gly Ser Asp Cys Ser Ser Pro Val Glu Cys Asp Phe
495 500 505

Thr Ser Pro Gly Asp Glu Gly Pro Pro Arg Ser Tyr Leu Arg Gln Trp
510 515 520

Val Val Ile Pro Pro Pro Leu Ser Ser Pro Gly Pro Gln Ala Ser
525 530 535

<220>
<221> CDS
<222> (441)..(2054)

<400> 8	
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tctgtctgcg gcccgatcat cctgctgcg gccgcctggt accttccttg ccgtctcttt	180
cctctgtctg ctgctctgtg ggacacctgc ctggaggccc agctgcccgat catcagagtg	240
acaggtctta tgacagcctg attggtgact cgggctgggt gtggattctc accccaggcc	300
tctgcctgct ttctcagacc ctcatctgtc acccccacgc tgaaccacgc tgccaccccc	360
agaagcccat cagactgccc ccagcacacg gaatggattt ctgagaaaga agccgaaaca	420
gaaggcccgat gggagtcagc atg ccg cgt ggc tgg gcc gcc ccc ttg etc ctg	473
Met Pro Arg Gly Trp Ala Ala Pro Leu Leu Leu	
1 5 10	
ctg ctg etc cag gga ggc tgg ggc tgc ccc gac etc gtc tgc tac acc	521
Leu Leu Leu Gln Gly Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr	
15 20 25	
gat tac etc cag acg gtc atc tgc atc ctg gaa atg tgg aac etc cac	569
Asp Tyr Leu Gln Thr Val Ile Cys Ile Leu Glu Met Trp Asn Leu His	
30 35 40	
ccc agc acg etc acc ctt acc tgg caa gac cag tat gaa gag ctg aag	617
Pro Ser Thr Leu Thr Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys	
45 50 55	

gac gag gcc acc tcc tgc agc etc cac agg tgc gcc cac aat gcc acg Asp Glu Ala Thr Ser Cys Ser Leu His Arg Ser Ala His Asn Ala Thr 60 65 70 75	665
cat gcc acc tac acc tgc cac atg gat gta ttc cac ttc atg gcc gac His Ala Thr Tyr Thr Cys His Met Asp Val Phe His Phe Met Ala Asp 80 85 90	713
gac att ttc agt gtc aac atc aca gac cag tct ggc aac tac tcc cag Asp Ile Phe Ser Val Asn Ile Thr Asp Gln Ser Gly Asn Tyr Ser Gln 95 100 105	761
gag tgt ggc agc ttt ctc ctg gct gag agc atc aag ccg gct ccc cct Glu Cys Gly Ser Phe Leu Leu Ala Glu Ser Ile Lys Pro Ala Pro Pro 110 115 120	809
ttc aac gtg act gtg acc ttc tca gga cag tat aat atc tcc tgg cgc Phe Asn Val Thr Val Thr Phe Ser Gly Gln Tyr Asn Ile Ser Trp Arg 125 130 135	857
tca gat tac gaa gac cct gcc ttc tac atg ctg aag ggc aag ctt cag Ser Asp Tyr Glu Asp Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln 140 145 150 155	905
tat gag ctg cag tac agg aac cgg gga gac ccc tgg gct gtg agt ccg Tyr Glu Leu Gln Tyr Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro 160 165 170	953
agg aga aag ctg atc tca gtg gac tca aga agt gtc tcc ctc ctc ccc Arg Arg Lys Leu Ile Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro 175 180 185	1001
ctg gag ttc cgc aaa gac tgc agc tat gag ctg cag gtg cgg gca ggg Leu Glu Phe Arg Lys Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly 190 195 200	1049
ccc atg cct ggc tcc tcc tac cag ggg acc tgg agt gaa tgg agt gac Pro Met Pro Gly Ser Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp 205 210 215	1097

ccg gtc atc ttt cag acc cag tca gag gag tta aag gaa ggc tgg aac	1145
Pro Val Ile Phe Gln Thr Gln Ser Glu Glu Leu Lys Glu Gly Trp Asn	
220 225 230 235	
cct cac ctg ctg ctt ctc ctc ctg ctt gtc ata gtc ttc att cct gcc	1193
Pro His Leu Leu Leu Leu Leu Leu Val Ile Val Phe Ile Pro Ala	
240 245 250	
ttc tgg agc ctg aag acc cat cca ttg tgg agg cta tgg aag aag ata	1241
Phe Trp Ser Leu Lys Thr His Pro Leu Trp Arg Leu Trp Lys Lys Ile	
255 260 265	
tgg gcc gtc ccc agc cct gag cgg ttc ttc atg ccc ctg tac aag ggc	1289
Trp Ala Val Pro Ser Pro Glu Arg Phe Phe Met Pro Leu Tyr Lys Gly	
270 275 280	
tgc agc gga gac ttc aag aaa tgg gtg ggt gca ccc ttc act ggc tcc	1337
Cys Ser Gly Asp Phe Lys Lys Trp Val Gly Ala Pro Phe Thr Gly Ser	
285 290 295	
agc ctg gag ctg gga ccc tgg agc cca gag gtg ccc tcc acc ctg gag	1385
Ser Leu Glu Leu Gly Pro Trp Ser Pro Glu Val Pro Ser Thr Leu Glu	
300 305 310 315	
gtg tac agc tgc cac cca cca cgg agc ccg gcc aag agg ctg cag ctc	1433
Val Tyr Ser Cys His Pro Pro Arg Ser Pro Ala Lys Arg Leu Gln Leu	
320 325 330	
acg gag cta caa gaa cca gca gag ctg gtg gag tct gac ggt gtg ccc	1481
Thr Glu Leu Gln Glu Pro Ala Glu Leu Val Glu Ser Asp Gly Val Pro	
335 340 345	
aag ccc agc ttc tgg ccg aca gcc cag aac tcg ggg ggc tca gct tac	1529
Lys Pro Ser Phe Trp Pro Thr Ala Gln Asn Ser Gly Gly Ser Ala Tyr	
350 355 360	
agt gag gag agg gat cgg cca tac ggc ctg gtg tcc att gac aca gtg	1577
Ser Glu Glu Arg Asp Arg Pro Tyr Gly Leu Val Ser Ile Asp Thr Val	
365 370 375	
act gtg cta gat gca gag ggg cca tgc acc tgg ccc tgc agc tgt gag	1625

Thr Val Leu Asp Ala Glu Gly Pro Cys Thr Trp Pro Cys Ser Cys Glu	
380 385 390 395	
gat gac ggc tac cca gcc ctg gac ctg gat gct ggc ctg gag ccc agc	1673
Asp Asp Gly Tyr Pro Ala Leu Asp Leu Asp Ala Gly Leu Glu Pro Ser	
400 405 410	
cca ggc cta gag gac cca ctc ttg gat gca ggg acc aca gtc ctg tcc	1721
Pro Gly Leu Glu Asp Pro Leu Leu Asp Ala Gly Thr Thr Val Leu Ser	
415 420 425	
tgt ggc tgt gtc tca gct ggc agc cct ggg cta gga ggg ccc ctg gga	1769
Cys Gly Cys Val Ser Ala Gly Ser Pro Gly Leu Gly Gly Pro Leu Gly	
430 435 440	
agc ctc ctg gac aga cta aag cca ccc ctt gca gat ggg gag gac tgg	1817
Ser Leu Leu Asp Arg Leu Lys Pro Pro Leu Ala Asp Gly Glu Asp Trp	
445 450 455	
gct ggg gga ctg ccc tgg ggt ggc cgg tca cct gga ggg gtc tca gag	1865
Ala Gly Gly Leu Pro Trp Gly Gly Arg Ser Pro Gly Gly Val Ser Glu	
460 465 470 475	
agt gag gcg ggc tca ccc ctg gcc ggc ctg gat atg gac acg ttt gac	1913
Ser Glu Ala Gly Ser Pro Leu Ala Gly Leu Asp Met Asp Thr Phe Asp	
480 485 490	
agt ggc ttt gtg ggc tct gac tgc agc agc cct gtg gag tgt gac ttc	1961
Ser Gly Phe Val Gly Ser Asp Cys Ser Ser Pro Val Glu Cys Asp Phe	
495 500 505	
acc age ccc ggg gac gaa gga ccc ccc cgg age tac etc cgc cag tgg	2009
Thr Ser Pro Gly Asp Glu Gly Pro Pro Arg Ser Tyr Leu Arg Gln Trp	
510 515 520	
gtg gtc att cct ccg cca ctt teg age cct gga ccc cag gcc age taa	2057
Val Val Ile Pro Pro Pro Leu Ser Ser Pro Gly Pro Gln Ala Ser	
525 530 535	
tgaggctgac tggatgtcca gagctggcca ggccactggg ccctgagcca gagacaaggt	2117

cacctgggct gtgatgtgaa gacacctgca gcctttggtc tcttgatgg gcctttgagc 2177
 ctgatgttta cagtgtctgt gtgtgtgtgc atatgtgtgt gtgtgcatat gcatgtgtgt 2237
 gtgtgtgtgt gtcttaggtg cgcagtggca tgtccacgtg tgtgtgattg cacgtgcctg 2297
 tgggcctggg ataatgcccata tggtaactcca tgcattcacc tgcctgtgc atgtctggac 2357
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<210> 9

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Artificially Synthesized Primer Sequence

<400> 9

ccggctcccc ctttcaacgt gactgtgacc 30

<210> 10

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Artificially Synthesized Primer Sequence

<400> 10

ggcaagcttc agtatgagct gcagtacagg 30

<210> 11

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Artificially Synthesized Primer Sequence

<400> 11

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30

<210> 12

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Artificially Synthesized Primer Sequence

<400> 12

catgggccct gcccgcacct gcagtcata

30

<210> 13

<211> 1128

<212> DNA

<213> Homo sapiens

<220>

<221> CDS

<222> (1)..(1125)

<400> 13

atg	ccg	cgt	ggc	tgg	gcc	gcc	ccc	ttg	ctc	ctg	ctg	ctc	cag	gga	48
Met	Pro	Arg	Gly	Trp	Ala	Ala	Pro	Leu	Leu	Leu	Leu	Leu	Gln	Gly	
1				5				10					15		

ggc	tgg	ggc	tgc	ccc	gac	ctc	gtc	tgc	tac	acc	gat	tac	ctc	cag	acg	96
Gly	Trp	Gly	Cys	Pro	Asp	Leu	Val	Cys	Tyr	Thr	Asp	Tyr	Leu	Gln	Thr	
			20					25					30			

gtc	atc	tgc	atc	ctg	gaa	atg	tgg	aac	ctc	cac	ccc	agc	acg	ctc	acc	144
Val	Ile	Cys	Ile	Leu	Glu	Met	Trp	Asn	Leu	His	Pro	Ser	Thr	Leu	Thr	
		35					40					45				

ctt	acc	tgg	caa	gac	cag	tat	gaa	gag	ctg	aag	gac	gag	gcc	acc	tcc	192
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys Asp Glu Ala Thr Ser	
50 55 60	
tgc agc ctc cac agg tgc gcc cac aat gcc acg cat gcc acc tac acc	240
Cys Ser Leu His Arg Ser Ala His Asn Ala Thr His Ala Thr Tyr Thr	
65 70 75 80	
tgc cac atg gat gta ttc cac ttc atg gcc gac gac att ttc agt gtc	288
Cys His Met Asp Val Phe His Phe Met Ala Asp Asp Ile Phe Ser Val	
85 90 95	
aac atc aca gac cag tct ggc aac tac tcc cag gag tgt ggc agc ttt	336
Asn Ile Thr Asp Gln Ser Gly Asn Tyr Ser Gln Glu Cys Gly Ser Phe	
100 105 110	
ctc ctg gct gag agc atc aag ccg gct ccc cct ttc aac gtg act gtg	384
Leu Leu Ala Glu Ser Ile Lys Pro Ala Pro Pro Phe Asn Val Thr Val	
115 120 125	
acc ttc tca gga cag tat aat atc tcc tgg cgc tca gat tac gaa gac	432
Thr Phe Ser Gly Gln Tyr Asn Ile Ser Trp Arg Ser Asp Tyr Glu Asp	
130 135 140	
cct gcc ttc tac atg ctg aag ggc aag ctt cag tat gag ctg cag tac	480
Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln Tyr Glu Leu Gln Tyr	
145 150 155 160	
agg aac cgg gga gac ccc tgg gct gtg agt ccg agg aga aag ctg atc	528
Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro Arg Arg Lys Leu Ile	
165 170 175	
tca gtg gac tca aga agt gtc tcc ctc ctc ccc ctg gag ttc cgc aaa	576
Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro Leu Glu Phe Arg Lys	
180 185 190	
gac tgc agc tat gag ctg cag gtg cgg gca ggg ccc atg cct ggc tcc	624
Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly Pro Met Pro Gly Ser	
195 200 205	
tcc tac cag ggg acc tgg agt gaa tgg agt gac ccg gtc atc ttt cag	672
Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp Pro Val Ile Phe Gln	

210	215	220	
acc cag tea gag acc gcc tgg atc tcc ttg gtg acc gct ctg cat cta			720
Thr Gln Ser Glu Thr Ala Trp Ile Ser Leu Val Thr Ala Leu His Leu			
225	230	235	240
gtg ctg ggc ctc agc gcc gtc ctg ggc ctg ctg ctg ctg agg tgg cag			768
Val Leu Gly Leu Ser Ala Val Leu Gly Leu Leu Leu Leu Arg Trp Gln			
	245	250	255
ttt cct gca cac tac agg aga ctg agg cat gcc ctg tgg ccc tca ctt			816
Phe Pro Ala His Tyr Arg Arg Leu Arg His Ala Leu Trp Pro Ser Leu			
	260	265	270
cca gac ctg cac cgg gtc cta ggc cag tac ctt agg gac act gca gcc			864
Pro Asp Leu His Arg Val Leu Gly Gln Tyr Leu Arg Asp Thr Ala Ala			
	275	280	285
ctg agc ccg ccc aag gcc aca gtc tca gat acc tgt gaa gaa gtg gaa			912
Leu Ser Pro Pro Lys Ala Thr Val Ser Asp Thr Cys Glu Glu Val Glu			
	290	295	300
ccc agc ctc ctt gaa atc ctc ccc aag tcc tca gag agg act cct ttg			960
Pro Ser Leu Leu Glu Ile Leu Pro Lys Ser Ser Glu Arg Thr Pro Leu			
305	310	315	320
ccc ctg tgt tcc tcc cag gcc cag atg gac tac cga aga ttg cag cct			1008
Pro Leu Cys Ser Ser Gln Ala Gln Met Asp Tyr Arg Arg Leu Gln Pro			
	325	330	335
tct tgc ctg ggg acc atg ccc ctg tct gtg tgc cca ccc atg gct gag			1056
Ser Cys Leu Gly Thr Met Pro Leu Ser Val Cys Pro Pro Met Ala Glu			
	340	345	350
tca ggg tcc tgc tgt acc acc cac att gcc aac cat tcc tac cta cca			1104
Ser Gly Ser Cys Cys Thr Thr His Ile Ala Asn His Ser Tyr Leu Pro			
	355	360	365
cta agc tat tgg cag cag cct tga			1128
Leu Ser Tyr Trp Gln Gln Pro			
370	375		

<210> 14
 <211> 375
 <212> PRT
 <213> Homo sapiens

<400> 14

Met Pro Arg Gly Trp Ala Ala Pro Leu Leu Leu Leu Leu Leu Gln Gly
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Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr Asp Tyr Leu Gln Thr
 20 25 30

Val Ile Cys Ile Leu Glu Met Trp Asn Leu His Pro Ser Thr Leu Thr
 35 40 45

Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys Asp Glu Ala Thr Ser
 50 55 60

Cys Ser Leu His Arg Ser Ala His Asn Ala Thr His Ala Thr Tyr Thr
 65 70 75 80

Cys His Met Asp Val Phe His Phe Met Ala Asp Asp Ile Phe Ser Val
 85 90 95

Asn Ile Thr Asp Gln Ser Gly Asn Tyr Ser Gln Glu Cys Gly Ser Phe
 100 105 110

Leu Leu Ala Glu Ser Ile Lys Pro Ala Pro Pro Phe Asn Val Thr Val
 115 120 125

Thr Phe Ser Gly Gln Tyr Asn Ile Ser Trp Arg Ser Asp Tyr Glu Asp
 130 135 140

Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln Tyr Glu Leu Gln Tyr
 145 150 155 160

Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro Arg Arg Lys Leu Ile
 165 170 175

Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro Leu Glu Phe Arg Lys
 180 185 190

Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly Pro Met Pro Gly Ser
 195 200 205

Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp Pro Val Ile Phe Gln
 210 215 220

Thr Gln Ser Glu Thr Ala Trp Ile Ser Leu Val Thr Ala Leu His Leu
 225 230 235 240

Val Leu Gly Leu Ser Ala Val Leu Gly Leu Leu Leu Leu Arg Trp Gln
 245 250 255

Phe Pro Ala His Tyr Arg Arg Leu Arg His Ala Leu Trp Pro Ser Leu
 260 265 270

Pro Asp Leu His Arg Val Leu Gly Gln Tyr Leu Arg Asp Thr Ala Ala
 275 280 285

Leu Ser Pro Pro Lys Ala Thr Val Ser Asp Thr Cys Glu Glu Val Glu
 290 295 300

Pro Ser Leu Leu Glu Ile Leu Pro Lys Ser Ser Glu Arg Thr Pro Leu
 305 310 315 320

Pro Leu Cys Ser Ser Gln Ala Gln Met Asp Tyr Arg Arg Leu Gln Pro
 325 330 335

Ser Cys Leu Gly Thr Met Pro Leu Ser Val Cys Pro Pro Met Ala Glu
 340 345 350

Ser Gly Ser Cys Cys Thr Thr His Ile Ala Asn His Ser Tyr Leu Pro
 355 360 365

Leu Ser Tyr Trp Gln Gln Pro
 370 375

<211> 1383
 <212> DNA
 <213> Homo sapiens

<220>
 <221> CDS
 <222> (1)..(1380)

<400> 15

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Met	Pro	Arg	Gly	Trp	Ala	Ala	Pro	Leu	Leu	Leu	Leu	Leu	Leu	Gln	Gly	
1				5				10						15		

ggc	tgg	ggc	tgc	ccc	gac	ctc	gtc	tgc	tac	acc	gat	tac	ctc	cag	acg	96
Gly	Trp	Gly	Cys	Pro	Asp	Leu	Val	Cys	Tyr	Thr	Asp	Tyr	Leu	Gln	Thr	
			20					25					30			

gtc	atc	tgc	atc	ctg	gaa	atg	tgg	aac	ctc	cac	ccc	agc	acg	ctc	acc	144
Val	Ile	Cys	Ile	Leu	Glu	Met	Trp	Asn	Leu	His	Pro	Ser	Thr	Leu	Thr	
		35					40					45				

ctt	acc	tgg	caa	gac	cag	tat	gaa	gag	ctg	aag	gac	gag	gcc	acc	tcc	192
Leu	Thr	Trp	Gln	Asp	Gln	Tyr	Glu	Glu	Leu	Lys	Asp	Glu	Ala	Thr	Ser	
		50					55				60					

tgc	agc	ctc	cac	agg	tgc	gcc	cac	aat	gcc	acg	cat	gcc	acc	tac	acc	240
Cys	Ser	Leu	His	Arg	Ser	Ala	His	Asn	Ala	Thr	His	Ala	Thr	Tyr	Thr	
65					70					75	—				80	

tgc	cac	atg	gat	gta	ttc	cac	ttc	atg	gcc	gac	gac	att	ttc	agt	gtc	288
Cys	His	Met	Asp	Val	Phe	His	Phe	Met	Ala	Asp	Asp	Ile	Phe	Ser	Val	
				85					90				95			

aac	atc	aca	gac	cag	tct	ggc	aac	tac	tcc	cag	gag	tgt	ggc	agc	ttt	336
Asn	Ile	Thr	Asp	Gln	Ser	Gly	Asn	Tyr	Ser	Gln	Glu	Cys	Gly	Ser	Phe	
				100				105					110			

ctc	ctg	gct	gag	agc	atc	aag	ccg	gct	ccc	cct	ttc	aac	gtg	act	gtg	384
Leu	Leu	Ala	Glu	Ser	Ile	Lys	Pro	Ala	Pro	Pro	Phe	Asn	Val	Thr	Val	
				115				120					125			

acc ttc tca gga cag tat aat atc tcc tgg cgc tca gat tac gaa gac	432
Thr Phe Ser Gly Gln Tyr Asn Ile Ser Trp Arg Ser Asp Tyr Glu Asp	
130 135 140	
cct gcc ttc tac atg ctg aag ggc aag ctt cag tat gag ctg cag tac	480
Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln Tyr Glu Leu Gln Tyr	
145 150 155 160	
agg aac cgg gga gac ccc tgg gct gtg agt ccg agg aga aag ctg atc	528
Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro Arg Arg Lys Leu Ile	
165 170 175	
tca gtg gac tca aga agt gtc tcc ctc ctc ccc ctg gag ttc cgc aaa	576
Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro Leu Glu Phe Arg Lys	
180 185 190	
gac tgg agc tat gag ctg cag gtg cgg gca ggg ccc atg cct ggc tcc	624
Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly Pro Met Pro Gly Ser	
195 200 205	
tcc tac cag ggg acc tgg agt gaa tgg agt gac ccg gtc atc ttt cag	672
Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp Pro Val Ile Phe Gln	
210 215 220	
acc cag tca gag gag ccc aaa tct tgt gac aaa act cac aca tgc cca	720
Thr Gln Ser Glu Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro	
225 230 235 240	
ccg tgc cca gca cct gaa ctc ctg ggg gga ccg tca gtc ttc ctc ttc	768
Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe	
245 250 255	
ccc cca aaa ccc aag gac acc ctc atg atc tcc cgg acc cct gag gtc	816
Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val	
260 265 270	
aca tgc gtg gtg gtg gac gtg agc cac gaa gac cct gag gtc aag ttc	864
Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe	
275 280 285	
aac tgg tac gtg gac ggc gtg gag gtg cat aat gcc aag aca aag ccg	912

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro	
290 295 300	
cgg gag gag cag tac aac agc acg tac cgg gtg gtc agc gtc ctc acc	960
Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr	
305 310 315 320	
gtc ctg cac cag gac tgg ctg aat ggc aag gag tac aag tgc aag gtc	1008
Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val	
325 330 335	
tcc aac aaa gcc ctc cca gcc ccc atc gag aaa acc atc tcc aaa gcc	1056
Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala	
340 345 350	
aaa ggg cag ccc cga gaa cca cag gtg tac acc ctg ccc cca tcc cgg	1104
Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg	
355 360 365	
gat gag ctg acc aag aac cag gtc agc ctg acc tgc ctg gtc aaa ggc	1152
Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly	
370 375 380	
ttc tat ccc agc gac atc gcc gtg gag tgg gag agc aat ggg cag ccg	1200
Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro	
385 390 395 400	
gag aac aac tac aag acc acg cct ccc gtg ctg gac tcc gac ggc tcc	1248
Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser	
405 410 415	
ttc ttc ctc' tac agc aag ctc acc gtg gac aag agc agg tgg cag cag	1296
Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser-Arg Trp Gln Gln	
420 425 430	
ggg aac gtc ttc tca tgc tcc gtg atg cat gag gct ctg cac aac cac	1344
Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His	
435 440 445	
tac acg cag aag agc ctc tcc ctg tct ccg ggt aaa tga	1383
Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys	

450

455

460

<210> 16

<211> 460

<212> PRT

<213> Homo sapiens

<400> 16

Met Pro Arg Gly Trp Ala Ala Pro Leu Leu Leu Leu Leu Leu Gln Gly
 1 5 10 15

Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr Asp Tyr Leu Gln Thr
 20 25 30

Val Ile Cys Ile Leu Glu Met Trp Asn Leu His Pro Ser Thr Leu Thr
 35 40 45

Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys Asp Glu Ala Thr Ser
 50 55 60

Cys Ser Leu His Arg Ser Ala His Asn Ala Thr His Ala Thr Tyr Thr
 65 70 75 80

Cys His Met Asp Val Phe His Phe Met Ala Asp Asp Ile Phe Ser Val
 85 90 95

Asn Ile Thr Asp Gln Ser Gly Asn Tyr Ser Gln Glu Cys Gly Ser Phe
 100 105 110

Leu Leu Ala Glu Ser Ile Lys Pro Ala Pro Pro Phe Asn Val Thr Val
 115 120 125

Thr Phe Ser Gly Gln Tyr Asn Ile Ser Trp Arg Ser Asp Tyr Glu Asp
 130 135 140

Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln Tyr Glu Leu Gln Tyr
 145 150 155 160

Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro Arg Arg Lys Leu Ile
 165 170 175

Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro Leu Glu Phe Arg Lys
 180 185 190
 Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly Pro Met Pro Gly Ser
 195 200 205
 Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp Pro Val Ile Phe Gln
 210 215 220
 Thr Gln Ser Glu Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro
 225 230 235 240
 Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe
 245 250 255
 Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val
 260 265 270
 Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe
 275 280 285
 Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro
 290 295 300
 Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr
 305 310 315 320
 Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val
 325 330 335
 Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala
 340 345 350
 Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg
 355 360 365
 Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
 370 375 380
 Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro

385		390		395		400
Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser						
	405			410		415
Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln						
	420		425			430
Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His						
	435		440			445
Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys						
	450		455			460

<210> 17
 <211> 477
 <212> DNA
 <213> Homo sapiens

<220>
 <221> CDS
 <222> (1)..(474)

<400> 17

atg ccg cgt ggc tgg gcc gcc ccc ttg ctc ctg ctg ctg ctc cag gga	48
Met Pro Arg Gly Trp Ala Ala Pro Leu Leu Leu Leu Leu Leu Gln Gly	
1 5 10 15	
ggc tgg ggc tgc ccc gac ctc gtc tgc tac acc gat tac ctc cag acg	96
Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr Asp Tyr Leu Gln Thr	
20 25 30	
gtc atc tgc atc ctg gaa atg tgg aac ctc cac ccc agc acg ctc acc	144
Val Ile Cys Ile Leu Glu Met Trp Asn Leu His Pro Ser Thr Leu Thr	
35 40 45	
ctt acc tgg caa gac cag tat gaa gag ctg aag gac gag gcc acc tcc	192
Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys Asp Glu Ala Thr Ser	
50 55 60	

tgc agc ctc cac agg tgc gcc cac aat gcc acg cat gcc acc tac acc 240
 Cys Ser Leu His Arg Ser Ala His Asn Ala Thr His Ala Thr Tyr Thr
 65 70 75 80

tgc cac atg gat gta ttc cac ttc atg gcc gac gac att ttc agt gtc 288
 Cys His Met Asp Val Phe His Phe Met Ala Asp Asp Ile Phe Ser Val
 85 90 95

aac atc aca gac cag tct gcc aac tac tcc cag gag tgt ggc agc ttt 336
 Asn Ile Thr Asp Gln Ser Gly Asn Tyr Ser Gln Glu Cys Gly Ser Phe
 100 105 110

ctc ctg gct gag agc aag tcc gag gag aaa gct gat ctc agt gga ctc 384
 Leu Leu Ala Glu Ser Lys Ser Glu Glu Lys Ala Asp Leu Ser Gly Leu
 115 120 125

aag aag tgt ctc cct cct ccc cct gga gtt ccg caa aga ctc gag cta 432
 Lys Lys Cys Leu Pro Pro Pro Gly Val Pro Gln Arg Leu Glu Leu
 130 135 140

agg gcg cgc cag gac tac aag gac gac gat gac aag acg cgt taa 477
 Arg Ala Arg Gln Asp Tyr Lys Asp Asp Asp Lys Thr Arg
 145 150 155

<210> 18

<211> 158

<212> PRT

<213> Homo sapiens

<400> 18

Met Pro Arg Gly Trp Ala Ala Pro Leu Leu Leu Leu Leu Leu Gln Gly
 1 5 10 15

Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr Asp Tyr Leu Gln Thr
 20 25 30

Val Ile Cys Ile Leu Glu Met Trp Asn Leu His Pro Ser Thr Leu Thr
 35 40 45

Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys Asp Glu Ala Thr Ser

50

55

60

Cys Ser Leu His Arg Ser Ala His Asn Ala Thr His Ala Thr Tyr Thr
 65 70 75 80

Cys His Met Asp Val Phe His Phe Met Ala Asp Asp Ile Phe Ser Val
 85 90 95

Asn Ile Thr Asp Gln Ser Gly Asn Tyr Ser Gln Glu Cys Gly Ser Phe
 100 105 110

Leu Leu Ala Glu Ser Lys Ser Glu Glu Lys Ala Asp Leu Ser Gly Leu
 115 120 125

Lys Lys Cys Leu Pro Pro Pro Gly Val Pro Gln Arg Leu Glu Leu
 130 135 140

Arg Ala Arg Gln Asp Tyr Lys Asp Asp Asp Asp Lys Thr Arg
 145 150 155

<210> 19

<211> 144

<212> PRT

<213> Mus musculus

<400> 19

Met Pro Arg Gly Trp Ala Ala Ser Leu Leu Leu Leu Leu Gln Gly
 1 5 10 15

Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr Asp Tyr Leu Gln Thr
 20 25 30

Val Ile Cys Ile Leu Glu Met Trp Asn Leu His Pro Ser Thr Leu Thr
 35 40 45

Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys Asp Glu Ala Thr Ser
 50 55 60

Cys Ser Leu His Arg Ser Ala His Asn Ala Thr His Ala Thr Tyr Thr
 65 70 75 80

Ser His Met Asp Val Phe His Phe Met Ala Asp Asp Ile Phe Ser Val
 85 90 95

Asn Ile Thr Asp Gln Ser Gly Asn Tyr Phe Gln Glu Cys Gly Ser Phe
 100 105 110

Leu Arg Ala Glu Ser Lys Ser Glu Glu Lys Ala Asp Leu Ser Gly Leu
 115 120 125

Lys Lys Cys Leu Pro Pro Pro Gly Val Pro Gln Arg Leu Glu Leu
 130 135 140

<210> 20

<211> 1960

<212> DNA

<213> Mus musculus

<400> 20

cagccagcgg cctcagacag acccaactggc gtctctctgc tgagtgaccg taagctcggc 60

gtctggccct ctgcctgcct ctccctgagt gtggctgaca gccacgcagc tgtgtctgtc 120

tgtctgcggc cegtgcaccc ctgctgcggc cgcctgttac cttccttgcc gtctctttcc 180

tctgtctgct gctctgtggg acacctgcct ggaggcccag ctgcccgta tcagagtgc 240

aggtcttatg acagcctgat tggtagctcg ggtgggtgt ggattctcac cccaggcctc 300

tgctgtcttt ctacagacct catcggtcac cccacgctg aaccagctg ccacccccag 360

aagcccatca gactgcccc agcacacgga atggatttct gagaaagaag ccgaaacaga 420

aggcccggtg gagtcagc atg ccg cgt ggc tgg gcc gcc tcc ttg ctc ctg 471

Met Pro Arg Gly Trp Ala Ala Ser Leu Leu Leu

1

5

10

ctg ctg ctc cag gga ggc tgg ggc tgc ccc gac ctc gtc tgc tac acc 519

Leu Leu Leu Gln Gly Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr

15

20

25

gat tac ctc cag acg gtc atc tgc atc ctg gaa atg tgg aac ctc cac 567
 Asp Tyr Leu Gln Thr Val Ile Cys Ile Leu Glu Met Trp Asn Leu His
 30 35 40

ccc agc acg ctc acc ctt acc tgg caa gac cag tat gaa gag ctg aag 615
 Pro Ser Thr Leu Thr Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys
 45 50 55

gac gag gcc acc tcc tgc agc ctc cac agg tgc gcc cac aat gcc acg 663
 Asp Glu Ala Thr Ser Cys Ser Leu His Arg Ser Ala His Asn Ala Thr
 60 65 70 75

cat gcc acc tac acc agc cac atg gat gta ttc cac ttc atg gcc gac 711
 His Ala Thr Tyr Thr Ser His Met Asp Val Phe His Phe Met Ala Asp
 80 85 90

gac att ttc agt gtc aac atc aca gac cag tct ggc aac tac ttc cag 759
 Asp Ile Phe Ser Val Asn Ile Thr Asp Gln Ser Gly Asn Tyr Phe Gln
 95 100 105

gag tgt ggc agc ttt ctc cgg gct gag agc aag tcc gag gag aaa gct 807
 Glu Cys Gly Ser Phe Leu Arg Ala Glu Ser Lys Ser Glu Glu Lys Ala
 110 115 120

gat ctc agt gga ctc aag aag tgt ctc cct cct ccc cct gga gtt cgg 855
 Asp Leu Ser Gly Leu Lys Lys Cys Leu Pro Pro Pro Pro Gly Val Pro
 125 130 135

caa aga ctc gag cta tgagctgcag gtgcgggcag ggcccatgcc tggctcctcc 910
 Gln Arg Leu Glu Leu
 140

taccagggga cctggagtga atggagtgc cgggtcatct ttcagaccca gtcagaggag 970

ttaaaggaag gctggaaccc tcacctgtg ettctctccc tgettgtcat agtcttcatt 1030

cctgccttct ggagcctgaa gaccatecca ttgtggaggc tatggaagaa gatattggcc 1090

gtccccagcc ctgagcgggtt cttcatgccc ctgtacaagg gctgcagcgg agacttcaag 1150

aaatgggtgg gtgcaccctt cactggtcc agcctggagc tgggaccctg gagcccagag 1210
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 cagctcacgg agctacaaga accagcagag ctggtggagt ctgacggtgt gccaagccc 1330
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 aagccacccc ttgcagatgg ggaggactgg gctgggggac tgcctgggg tggccggtca 1690
 cctggagggg tctcagagag tgaggcgggc tcaccttg cggcctgga tatggacag 1750
 tttgacagtg gctttgtgtg ctctgactgc agcagccctg tggagtgtga cttaccagc 1810
 cccggggacg aaggaccccc cggagctac ctccgccagt ggggtggtcat tcctccgcca 1870
 ctttcgagcc ctggacccca ggccagctaa tgaggctgac tggatgtcca gagctggcca 1930
 ggccactggg ccctgagcca gaaaaaaaaa 1960

<210> 21

<211> 538

<212> PRT

<213> Mus musculus

<400> 21

Met Pro Arg Gly Trp Ala Ala Ser Leu Leu Leu Leu Leu Leu Gln Gly

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Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr Asp Tyr Leu Gln Thr

20

25

30

Val Ile Cys Ile Leu Glu Met Trp Asn Leu His Pro Ser Thr Leu Thr
 35 40 45
 Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys Asp Glu Ala Thr Ser
 50 55 60
 Cys Ser Leu His Arg Ser Ala His Asn Ala Thr His Ala Thr Tyr Thr
 65 70 75 80
 Ser His Met Asp Val Phe His Phe Met Ala Asp Asp Ile Phe Ser Val
 85 90 95
 Asn Ile Thr Asp Gln Ser Gly Asn Tyr Phe Gln Glu Cys Gly Ser Phe
 100 105 110
 Leu Arg Ala Glu Ser Ile Lys Pro Ala Pro Pro Phe Asn Val Thr Val
 115 120 125
 Thr Phe Ser Gly Gln Tyr Asn Ile Ser Arg Arg Ser Asp Tyr Glu Asp
 130 135 140
 Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln Tyr Glu Leu Gln Tyr
 145 150 155 160
 Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro Arg Arg Lys Leu Ile
 165 170 175
 Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro Leu Glu Phe Arg Lys
 180 185 190
 Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly Pro Met Pro Gly Ser
 195 200 205
 Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp Pro Val Ile Phe Gln
 210 215 220
 Thr Gln Ser Glu Glu Leu Lys Glu Gly Trp Asn Pro His Leu Leu Leu
 225 230 235 240
 Leu Leu Leu Leu Val Ile Val Phe Ile Pro Ala Phe Trp Ser Leu Lys
 245 250 255

Thr His Pro Leu Trp Arg Leu Trp Lys Lys Ile Trp Ala Val Pro Ser
260 265 270

Pro Glu Arg Phe Phe Met Pro Leu Tyr Lys Gly Cys Ser Gly Asp Phe
275 280 285

Lys Lys Trp Val Gly Ala Pro Phe Thr Gly Ser Ser Leu Glu Leu Gly
290 295 300

Pro Trp Ser Pro Glu Val Pro Ser Thr Leu Glu Val Tyr Ser Cys His
305 310 315 320

Pro Pro Arg Ser Pro Ala Lys Arg Leu Gln Leu Thr Glu Leu Gln Glu
325 330 335

Pro Ala Glu Leu Val Glu Ser Asp Gly Val Pro Lys Pro Ser Phe Trp
340 345 350

Pro Thr Ala Gln Asn Ser Gly Gly Ser Ala Tyr Ser Glu Glu Arg Asp
355 360 365

Arg Pro Tyr Gly Leu Val Ser Ile Asp Thr Val Thr Val Leu Asp Ala
370 375 380

Glu Gly Pro Cys Thr Trp Pro Cys Ser Cys Glu Asp Asp Gly Tyr Pro
385 390 395 400

Ala Leu Asp Leu Asp Ala Gly Leu Glu Pro Ser Pro Gly Leu Glu Asp
405 410 415

Pro Leu Leu Asp Ala Gly Thr Thr Val Leu Ser Cys Gly Cys Val Ser
420 425 430

Ala Gly Ser Pro Gly Leu Gly Gly Pro Leu Gly Ser Leu Leu Asp Arg
435 440 445

Leu Lys Pro Pro Leu Ala Asp Gly Glu Asp Trp Ala Gly Gly Leu Pro
450 455 460

Trp Gly Gly Arg Ser Pro Gly Gly Val Ser Glu Ser Glu Ala Gly Ser

465 470 475 480
 Pro Leu Ala Gly Leu Asp Met Asp Thr Phe Asp Ser Gly Phe Val Cys
 485 490 495
 Ser Asp Cys Ser Ser Pro Val Glu Cys Asp Phe Thr Ser Pro Gly Asp
 500 505 510
 Glu Gly Pro Pro Arg Ser Tyr Leu Arg Gln Trp Val Val Ile Pro Pro
 515 520 525
 Pro Leu Ser Ser Pro Gly Pro Gln Ala Ser
 530 535

<210> 22

<211> 2115

<212> DNA

<213> Mus musculus

<400> 22

cagccagcgg cctcagacag acccactggc gtctctctgc tgagtgaccg taagctcggc 60

gtctggccct ctgectgcct ctccctgagt gtggctgaca gccacgcagc tgtgtctgtc 120

tgtctgcggc cegtgcattc ctgctgcggc cgcttggtac ctcccttgcc gtctctttcc 180

tctgtctgct gctctgtggg acacctgcct ggaggcccag ctgcccgtca tcagagtgc 240

aggtcttatg acagcctgat tggtagctcg ggctgggtgt ggattctcac ccaggcctc 300

tgctgtcttt ctacagacct catcggtcac cccacgctg aaccagctg ccacccccag 360

aagcccatca gactgcccc agcacacgga atggatttct gagaaagaag ccgaaacaga 420

aggcccgtgg gactcagc atg ccg cgt ggc tgg gcc gcc tcc ttg etc ctg 471

Met Pro Arg Gly Trp Ala Ala Ser Leu Leu Leu

1

5

10

ctg ctg etc cag gga ggc tgg ggc tgc ccc gac etc gtc tgc tac acc 519

Leu Leu Leu Gln Gly Gly Trp Gly Cys Pro Asp Leu Val Cys Tyr Thr

	15	20	25	
gat tac ctc cag acg gtc atc tgc atc ctg gaa atg tgg aac ctc cac				567
Asp Tyr Leu Gln Thr Val Ile Cys Ile Leu Glu Met Trp Asn Leu His				
	30	35	40	
ccc agc acg ctc acc ctt acc tgg caa gac cag tat gaa gag ctg aag				615
Pro Ser Thr Leu Thr Leu Thr Trp Gln Asp Gln Tyr Glu Glu Leu Lys				
	45	50	55	
gac gag gcc acc tcc tgc agc ctc cac agg tgc gcc cac aat gcc acg				663
Asp Glu Ala Thr Ser Cys Ser Leu His Arg Ser Ala His Asn Ala Thr				
	60	65	70	75
cat gcc acc tac acc agc cac atg gat gta ttc cac ttc atg gcc gac				711
His Ala Thr Tyr Thr Ser His Met Asp Val Phe His Phe Met Ala Asp				
	80	85	90	
gac att ttc agt gtc aac atc aca gac cag tct ggc aac tac ttc cag				759
Asp Ile Phe Ser Val Asn Ile Thr Asp Gln Ser Gly Asn Tyr Phe Gln				
	95	100	105	
gag tgt ggc agc ttt ctc cgg gct gag agc atc aag ccg gct ccc cct				807
Glu Cys Gly Ser Phe Leu Arg Ala Glu Ser Ile Lys Pro Ala Pro Pro				
	110	115	120	
ttc aac gtg act gtg acc ttc tca gga cag tat aat atc tcc agg cgc				855
Phe Asn Val Thr Val Thr Phe Ser Gly Gln Tyr Asn Ile Ser Arg Arg				
	125	130	135	
tca gat tac gaa gac cct gcc ttc tac atg ctg aag ggc aag ctt cag				903
Ser Asp Tyr Glu Asp Pro Ala Phe Tyr Met Leu Lys Gly Lys Leu Gln				
	140	145	150	155
tat gag ctg cag tac agg aac cgg gga gac ccc tgg gct gtg agt ccg				951
Tyr Glu Leu Gln Tyr Arg Asn Arg Gly Asp Pro Trp Ala Val Ser Pro				
	160	165	170	
agg aga aag ctg atc tca gtg gac tca aga agt gtc tcc ctc ctc ccc				999
Arg Arg Lys Leu Ile Ser Val Asp Ser Arg Ser Val Ser Leu Leu Pro				
	175	180	185	

ctg gag ttc cgc aaa gac tgc agc tat gag ctg cag gtg cgg gca ggg	1047
Leu Glu Phe Arg Lys Asp Ser Ser Tyr Glu Leu Gln Val Arg Ala Gly	
190 195 200	
ccc atg cct ggc tcc tcc tac cag ggg acc tgg agt gaa tgg agt gac	1095
Pro Met Pro Gly Ser Ser Tyr Gln Gly Thr Trp Ser Glu Trp Ser Asp	
205 210 215	
ccg gtc atc ttt cag acc cag tca gag gag tta aag gaa ggc tgg aac	1143
Pro Val Ile Phe Gln Thr Gln Ser Glu Glu Leu Lys Glu Gly Trp Asn	
220 225 230 235	
cct cac ctg ctg ctt ctc ctc ctg ctt gtc ata gtc ttc att cct gcc	1191
Pro His Leu Leu Leu Leu Leu Leu Leu Val Ile Val Phe Ile Pro Ala	
240 245 250	
ttc tgg agc ctg aag acc cat cca ttg tgg agg cta tgg aag aag ata	1239
Phe Trp Ser Leu Lys Thr His Pro Leu Trp Arg Leu Trp Lys Lys Ile	
255 260 265	
tgg gcc gtc ccc agc cct gag cgg ttc ttc atg ccc ctg tac aag ggc	1287
Trp Ala Val Pro Ser Pro Glu Arg Phe Phe Met Pro Leu Tyr Lys Gly	
270 275 280	
tgc agc gga gac ttc aag aaa tgg gtg ggt gca ccc ttc act ggc tcc	1335
Cys Ser Gly Asp Phe Lys Lys Trp Val Gly Ala Pro Phe Thr Gly Ser	
285 290 295	
agc ctg gag ctg gga ccc tgg agc cca gag gtg ccc tcc acc ctg gag	1383
Ser Leu Glu Leu Gly Pro Trp Ser Pro Glu Val Pro Ser Thr Leu Glu	
300 305 310 315	
gtg tac agc tgc cac cca cca cgg agc ccg gcc aag agg ctg cag ctc	1431
Val Tyr Ser Cys His Pro Pro Arg Ser Pro Ala Lys Arg Leu Gln Leu	
320 325 330	
acg gag cta caa gaa cca gca gag ctg gtg gag tct gac ggt gtg ccc	1479
Thr Glu Leu Gln Glu Pro Ala Glu Leu Val Glu Ser Asp Gly Val Pro	
335 340 345	

aag ccc agc ttc tgg ccg aca gcc cag aac tcg ggg ggc tca gct tac 1527
Lys Pro Ser Phe Trp Pro Thr Ala Gln Asn Ser Gly Gly Ser Ala Tyr
350 355 360

agt gag gag agg gat cgg cca tac ggc ctg gtg tcc att gac aca gtg 1575
Ser Glu Glu Arg Asp Arg Pro Tyr Gly Leu Val Ser Ile Asp Thr Val
365 370 375

act gtg cta gat gca gag ggg cca tgc acc tgg ccc tgc agc tgt gag 1623
Thr Val Leu Asp Ala Glu Gly Pro Cys Thr Trp Pro Cys Ser Cys Glu
380 385 390 395

gat gac ggc tac cca gcc ctg gac ctg gat gct ggc ctg gag ccc agc 1671
Asp Asp Gly Tyr Pro Ala Leu Asp Leu Asp Ala Gly Leu Glu Pro Ser
400 405 410

cca ggc cta gag gac cca ctc ttg gat gca ggg acc aca gtc ctg tcc 1719
Pro Gly Leu Glu Asp Pro Leu Leu Asp Ala Gly Thr Thr Val Leu Ser
415 420 425

tgt ggc tgt gtc tca gct ggc agc cct ggg cta gga ggg ccc ctg gga 1767
Cys Gly Cys Val Ser Ala Gly Ser Pro Gly Leu Gly Gly Pro Leu Gly
430 435 440

agc ctc ctg gac aga cta aag cca ccc ctt gca gat ggg gag gac tgg 1815
Ser Leu Leu Asp Arg Leu Lys Pro Pro Leu Ala Asp Gly Glu Asp Trp
445 450 455

gct ggg gga ctg ccc tgg ggt ggc cgg tca cct gga ggg gtc tca gag 1863
Ala Gly Gly Leu Pro Trp Gly Gly Arg Ser Pro Gly Gly Val Ser Glu
460 465 470 475

agt gag gcg ggc tca ccc ctg gcc ggc ctg gat atg gac acg ttt gac 1911
Ser Glu Ala Gly Ser Pro Leu Ala Gly Leu Asp Met Asp Thr Phe Asp
480 485 490

agt ggc ttt gtg tgc tct gac tgc agc agc cct gtg gag tgt gac ttc 1959
Ser Gly Phe Val Cys Ser Asp Cys Ser Ser Pro Val Glu Cys Asp Phe
495 500 505

acc agc ccc ggg gac gaa gga ccc ccc cgg agc tac ctc cgc cag tgg 2007

Thr Ser Pro Gly Asp Glu Gly Pro Pro Arg Ser Tyr Leu Arg Gln Trp
 510 515 520

gtg gtc att cct ccg cca ctt tcg agc cct gga ccc cag gcc agc 2052
 Val Val Ile Pro Pro Pro Leu Ser Ser Pro Gly Pro Gln Ala Ser
 525 530 535

taatgaggct gactggatgt ccagagctgg ccaggccact gggccctgag ccagaaaaaa 2112

aaa 2115

<210> 23
 <211> 411
 <212> DNA
 <213> Homo sapiens

<220>
 <221> 3' UTR
 <222> (1)..(411)

<400> 23
 taatgaggct gactggatgt ccagagctgg ccaggccact gggccctgag ccagagacaa 60
 ggtcacctgg gctgtgatgt gaagacacct gcagcctttg gtctcctgga tgggcctttg 120
 agcctgatgt ttacagtgtc tgttgtgttg tgcataatgt tgttgtgtga tatgcatgtg 180
 tgttgtgttg tgtgtcttag gtgcgcagtg gcatgtccac gtgtgtgtga ttgcacgtgc 240
 ctgtgggcct gggataatgc ccatggtact ccatgcattc acctgccctg tgcattgtctg 300
 gactcacgga gtcacccat gtgcacaagt gtgcacagta aacgtgtttg tggtaaacag 360
 aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa a 411

<210> 24
 <211> 877
 <212> DNA
 <213> Homo sapiens

<220>

<221> 3' UTR

<222> (1)..(877)

<400> 24

taatgaggct gactggatgt ccagagctgg ccaggccact gggccctgag ccagagacaa 60
 ggtcacctgg gctgtgatgt gaagacacct gcagcctttg gtctcctgga tgggcctttg 120
 agcctgatgt ttacagtgtc tgtgtgtgtg tgtgcataatg tgtgtgtgtg catatgcatg 180
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COMBINED DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled NOVEL HEMOPOIETIN RECEPTOR PROTEINS, the specification of which:

- ☐ is attached hereto.
☒ was filed on December 21, 2000 Application Serial No. 09/720,285 and was amended on _____.
☐ was described and claimed in PCT International Application No. _____ filed on _____ and as amended under PCT Article 19 on _____.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

Country	Application No.	Filing Date	Priority Claimed	
PCT	PCT/JP99/03351	23 June 1999	<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No
Japan	10/214720	24 June 1998	<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No
Japan	10/297409	19 October 1998	<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No

I hereby appoint the following attorneys and/or agents to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

J. Peter Fasse, Reg. No. 32,983 John W. Freeman, Reg. No. 29,066

John F. Hayden, Reg. No. 37,640 Timothy A. French, Reg. No. 30,175

Janis K. Fraser, Reg. No. 34,819 John T. Li, Reg. No. 44,210

Address all telephone calls to J. PETER FASSE at telephone number (617) 542-5070.

Address all correspondence to J. PETER FASSE at:

FISH & RICHARDSON P.C.
225 Franklin Street
Boston, MA 02110-2804

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patents issued thereon.

Combined Declaration and Power of Attorney

Page 2 of 2 Pages

1-00
Full Name of Inventor: HITOSHI NOMURA

Inventor's Signature: Hitoshi Nomura

Date: Mar. 14 / 01

Residence Address:

c/o Chugai Research Institute For Molecular Medicine, Inc.,
153-2, Nagai, Niihari-mura, Niihari-gun, IBARAKI 300-4101 JAPAN JPX

Citizenship:

Japan

Post Office Address:

c/o Chugai Research Institute For Molecular Medicine, Inc.,
153-2, Nagai, Niihari-mura, Niihari-gun, IBARAKI 300-4101 JAPAN

2-00
Full Name of Inventor: ~~MASATSUGU MAEDA~~

Inventor's Signature: M. Maeda

Date: March 15 / 01

Residence Address:

c/o Chugai Research Institute For Molecular Medicine, Inc.,
153-2, Nagai, Niihari-mura, Niihari-gun, IBARAKI 300-4101 JAPAN JPX

Citizenship:

Japan

Post Office Address:

c/o Chugai Research Institute For Molecular Medicine, Inc.,
153-2, Nagai, Niihari-mura, Niihari-gun, IBARAKI 300-4101 JAPAN